Package 'OmnipathR'

October 24, 2022

Type Package

Title OmniPath web service client and more

Version 3.5.25

Description A client for the OmniPath web service
(https://www.omnipathdb.org) and many other resources.
It also includes functions to transform and pretty print
some of the downloaded data, functions to access a number
of other resources such as BioPlex, ConsensusPathDB, EVEX,
Gene Ontology, Guide to Pharmacology (IUPHAR/BPS), Harmonizome,
HTRIdb, Human Phenotype Ontology, InWeb InBioMap, KEGG Pathway,
Pathway Commons, Ramilowski et al. 2015, RegNetwork, ReMap, TF
census, TRRUST and Vinayagam et al. 2011. Furthermore, OmnipathR
features a close integration with the NicheNet method for
ligand activity prediction from transcriptomics data, and its
R implementation `nichenetr` (available only on github).

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URL https://saezlab.github.io/OmnipathR/

BugReports https://github.com/saezlab/OmnipathR/issues

biocViews GraphAndNetwork, Network, Pathways, Software,
ThirdPartyClient, DataImport, DataRepresentation,
GeneSignaling, GeneRegulation, SystemsBiology, Transcriptomics,
SingleCell, Annotation, KEGG

Encoding UTF-8

VignetteBuilder knitr

Depends R(>= 4.0)

Imports checkmate, crayon, curl, digest, dplyr, httr, igraph,
jsonlite, later, logger, magrittr, progress, purrr, rappdirs,
readr(>= 2.0.0), readxl, rlang, rmarkdown, rvest, stats,
stringr, tibble, tidyr, tidyselect, tools, utils, withr, xml2,
yaml

Suggests BiocStyle, biomaRt, bookdown, dnet, ggplot2, ggraph,
gprofiler2, knitr, mlrMBO, parallelMap, ParamHelpers,
Rgraphviz, smoof, supraHex, testthat

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Author Alberto Valdeolivas [aut] (<https://orcid.org/0000-0001-5482-9023>),
Denes Turei [cre, aut] (<https://orcid.org/0000-0002-7249-9379>),
Attila Gabor [aut] (<https://orcid.org/0000-0002-0776-1182>)

Maintainer Denes Turei <turei.denes@gmail.com>

R topics documented:

`.omnipath_options_defaults`

Default values for the package options

Description

These options describe the default settings for OmnipathR so you do not need to pass these parameters at each function call. Currently the only option useful for the public web service at omnipathdb.org is "omnipath.license". If you are a for-profit user set it to "'commercial'" to make sure all the data you download from OmniPath is legally allowed for commercial use. Otherwise just leave it as it is: "'academic'". If you don't use omnipathdb.org but within your organization you deployed your own pypath server and want to share data whith a limited availability to outside users, you may want to use a password. For this you can use the "omnipath.password" option. Also if you want the R package to work from another pypath server instead of omnipathdb.org, you can change the option "omnipath.url".

Usage

```
.omnipath_options_defaults
```

Format

An object of class `list` of length 16.

`all_uniprots` *A table with all UniProt records*

Description

Retrieves a table from UniProt with all proteins for a certain organism.

Usage

```
all_uniprots(fields = "id", reviewed = TRUE, organism = 9606)
```

Arguments

`fields`	Character vector of fields as defined by UniProt. For possible values please refer to https://www.uniprot.org/help/uniprotkb%5Fcolumn%5Fnames
`reviewed`	Retrieve only reviewed ('TRUE'), only unreviewed ('FALSE') or both ('NULL').
`organism`	Character or integer: name or identifier of the organism.

Value

Data frame (tibble) with the requested UniProt entries and fields.

Examples

```
human_swissprot_entries <- all_uniprots(fields = 'entry name')
human_swissprot_entries
# # A tibble: 20,396 x 1
#    `Entry name`
#    <chr>
#  1 OR4K3_HUMAN
#  2 O52A1_HUMAN
#  3 O2AG1_HUMAN
#  4 O10S1_HUMAN
#  5 O11G2_HUMAN
# # . with 20,386 more rows
```

all_uniprot_acs *All UniProt ACs for one organism*

Description

All UniProt ACs for one organism

Usage

```
all_uniprot_acs(organism = 9606, reviewed = TRUE)
```

Arguments

organism	Character or integer: name or identifier of the organism.
reviewed	Retrieve only reviewed ('TRUE'), only unreviewed ('FALSE') or both ('NULL').

Value

Character vector of UniProt accession numbers.

Examples

```
human_swissprot_acs <- all_uniprot_acs()
human_swissprot_acs[1:5]
# [1] "P51451" "A6H8Y1" "O60885" "Q9Y3X0" "P22223"
length(human_swissprot_acs)
# [1] 20376
mouse_swissprot_acs <- all_uniprot_acs("mouse")
```

ancestors *All ancestors in the ontology tree*

Description

Starting from the selected nodes, recursively walks the ontology tree until it reaches the root. Collects all visited nodes, which are the ancestors (parents) of the starting nodes.

Usage

```
ancestors(
  terms,
  db_key = "go_basic",
  ids = TRUE,
 relations = c("is_a", "part_of", "occurs_in", "regulates", "positively_regulates",
    "negatively_regulates")
)
```

Arguments

terms	Character vector of ontology term IDs or names. A mixture of IDs and names can be provided.
db_key	Character: key to identify the ontology database. For the available keys see omnipath_show_db.
ids	Logical: whether to return IDs or term names.
relations	Character vector of ontology relation types. Only these relations will be used.

Details

Note: this function relies on the database manager, the first call might take long because of the database load process. Subsequent calls within a short period should be faster. See get_ontology_db.

Value

Character vector of ontology IDs. If the input terms are all root nodes, NULL is returned. The starting nodes won't be included in the result unless some of them are ancestors of other starting nodes.

Examples

```
ancestors('GO:0005035', ids = FALSE)
# [1] "molecular_function"
# [2] "transmembrane signaling receptor activity"
# [3] "signaling receptor activity"
# [4] "molecular transducer activity"
```

---

annotated_network *Network interactions with annotations*

Description

Annotations are often useful in a network context, e.g. one might want to label the interacting partners by their pathway membership. This function takes a network data frame and joins an annotation data frame from both the left and the right side, so both the source and target molecular entities will be labeled by their annotations. If one entity has many annotations these will yield many rows, hence the interacting pairs won't be unique across the data frame any more. Also if one entity has really many annotations the resulting data frame might be huge, we recommend to be careful with that. Finally, if you want to do the same but with intercell annotations, there is the import_intercell_network function.

Usage

```
annotated_network(
  network = NULL,
  annot = NULL,
  network_args = list(),
  annot_args = list(),
  ...
)
```

Arguments

network	Behaviour depends on type: if list, will be passed as arguments to import_omnipath_interactions to obtain a network data frame; if a data frame or tibble, it will be used as a network data frame; if a character vector, will be assumed to be a set of resource names and interactions will be queried from these resources.
annot	Either the name of an annotation resource (for a list of available resources call get_annotation_resources), or an annotation data frame. If the data frame contains more than one resources, only the first one will be used.
network_args	List: if 'network' is a resource name, pass these additional arguments to import_omnipath_interaction
annot_args	List: if 'annot' is a resource name, pass these additional arguments to import_omnipath_annotations.
...	Column names selected from the annotation data frame (passed to dplyr::select, if empty all columns will be selected.)

Value

A data frame of interactions with annotations for both interacting entities.

Examples

```
signalink_with_pathways <-
    annotated_network('SignaLink3', 'SignaLink_pathway')
```

---

annotation_categories *Annotation categories and resources*

Description

A full list of annotation resources, keys and values.

Usage

```
annotation_categories()
```

Value

A data frame with resource names, annotation key labels and for each key all possible values.

Examples

```
annot_cat <- annotation_categories()
annot_cat
# # A tibble: 46,307 x 3
#    source           label    value
#    <chr>            <chr>    <chr>
#  1 connectomeDB2020 role     ligand
#  2 connectomeDB2020 role     receptor
#  3 connectomeDB2020 location ECM
#  4 connectomeDB2020 location plasma membrane
#  5 connectomeDB2020 location secreted
#  6 KEGG-PC          pathway  Alanine, aspartate and glutamate metabolism
#  7 KEGG-PC          pathway  Amino sugar and nucleotide sugar metabolism
#  8 KEGG-PC          pathway  Aminoacyl-tRNA biosynthesis
#  9 KEGG-PC          pathway  Arachidonic acid metabolism
# 10 KEGG-PC          pathway  Arginine and proline metabolism
```

---

biomart_query *Query the Ensembl BioMart web service*

Description

Query the Ensembl BioMart web service

Usage

```
biomart_query(
  attrs = NULL,
  filters = NULL,
  transcript = FALSE,
  peptide = FALSE,
  gene = FALSE,
  dataset = "hsapiens_gene_ensembl"
)
```

Arguments

attrs	Character vector: one or more Ensembl attribute names.
filters	Character vector: one or more Ensembl filter names.
transcript	Logical: include Ensembl transcript IDs in the result.
peptide	Logical: include Ensembl peptide IDs in the result.
gene	Logical: include Ensembl gene IDs in the result.
dataset	Character: An Ensembl dataset name.

Value

Data frame with the query result

Examples

```
cel_genes <- biomart_query(
    attrs = c("external_gene_name", "start_position", "end_position"),
    gene = TRUE,
    dataset = "celegans_gene_ensembl"
)
cel_genes
# # A tibble: 46,934 × 4
#   ensembl_gene_id external_gene_name start_position end_position
#   <chr>           <chr>                      <dbl>        <dbl>
# 1 WBGene00000001  aap-1                    5107843      5110183
# 2 WBGene00000002  aat-1                    9599178      9601695
# 3 WBGene00000003  aat-2                    9244402      9246360
# 4 WBGene00000004  aat-3                    2552260      2557736
# 5 WBGene00000005  aat-4                    6272529      6275721
# # . with 46,924 more rows
```

---

bioplex1 *Downloads the BioPlex version 1.0 interaction dataset*

Description

This dataset contains ~24,000 interactions detected in HEK293T cells using 2,594 baits. More details at https://bioplex.hms.harvard.edu/interactions.php.

Usage

```
bioplex1()
```

Value

Data frame (tibble) with interactions.

See Also

- bioplex2
- bioplex3
- bioplex_hct116_1
- bioplex_all

Examples

```
bioplex_interactions <- bioplex1()
nrow(bioplex_interactions)
# [1] 23744
colnames(bioplex_interactions)
# [1] "GeneA"         "GeneB"        "UniprotA"   "UniprotB"
```

```
# [5] "SymbolA"       "SymbolB"      "p_wrong"    "p_no_interaction"
# [9] "p_interaction"
```

`bioplex2` *Downloads the BioPlex version 2.0 interaction dataset*

Description

This dataset contains ~56,000 interactions detected in HEK293T cells using 5,891 baits. More details at https://bioplex.hms.harvard.edu/interactions.php

Usage

```
bioplex2()
```

Value

Data frame (tibble) with interactions.

See Also

- `bioplex1`
- `bioplex3`
- `bioplex_hct116_1`
- `bioplex_all`

Examples

```
bioplex_interactions <- bioplex2()
nrow(bioplex_interactions)
# [1] 56553
colnames(bioplex_interactions)
# [1] "GeneA"         "GeneB"        "UniprotA"   "UniprotB"
# [5] "SymbolA"       "SymbolB"      "p_wrong"    "p_no_interaction"
# [9] "p_interaction"
```

bioplex3 *Downloads the BioPlex version 3.0 interaction dataset*

Description

This dataset contains ~120,000 interactions detected in HEK293T cells using 10,128 baits. More details at https://bioplex.hms.harvard.edu/interactions.php.

Usage

```
bioplex3()
```

Value

Data frame (tibble) with interactions.

See Also

- bioplex1
- bioplex2
- bioplex_hct116_1
- bioplex_all

Examples

```
bioplex_interactions <- bioplex3()
nrow(bioplex_interactions)
# [1] 118162
colnames(bioplex_interactions)
# [1] "GeneA"          "GeneB"         "UniprotA"    "UniprotB"
# [5] "SymbolA"        "SymbolB"       "p_wrong"     "p_no_interaction"
# [9] "p_interaction"
```

bioplex_all *Downloads all BioPlex interaction datasets*

Description

BioPlex provides four interaction datasets: version 1.0, 2.0, 3.0 and HCT116 version 1.0. This function downloads all of them, merges them to one data frame, removes the duplicates (based on unique pairs of UniProt IDs) and separates the isoform numbers from the UniProt IDs. More details at https://bioplex.hms.harvard.edu/interactions.php.

Usage

```
bioplex_all(unique = TRUE)
```

Arguments

unique	Logical. Collapse the duplicate interactions into single rows or keep them as they are. In case of merging duplicate records the maximum p value will be choosen for each record.

Value

Data frame (tibble) with interactions.

See Also

- bioplex1
- bioplex2
- bioplex3
- bioplex_hct116_1

Examples

```
bioplex_interactions <- bioplex_all()
bioplex_interactions
# # A tibble: 195,538 x 11
#    UniprotA IsoformA UniprotB IsoformB GeneA GeneB SymbolA SymbolB
#    <chr>       <int> <chr>       <int> <dbl> <dbl> <chr>   <chr>
#  1 A0AV02          2 Q5K4L6         NA 84561 11000 SLC12A8 SLC27A3
#  2 A0AV02          2 Q8N5V2         NA 84561 25791 SLC12A8 NGEF
#  3 A0AV02          2 Q9H6S3         NA 84561 64787 SLC12A8 EPS8L2
#  4 A0AV96          2 O00425          2 54502 10643 RBM47   IGF2BP3
#  5 A0AV96          2 O00443         NA 54502  5286 RBM47   PIK3C2A
#  6 A0AV96          2 O43426         NA 54502  8867 RBM47   SYNJ1
#  7 A0AV96          2 O75127         NA 54502 26024 RBM47   PTCD1
#  8 A0AV96          2 O95208          2 54502 22905 RBM47   EPN2
#  9 A0AV96          2 O95900         NA 54502 26995 RBM47   TRUB2
# 10 A0AV96          2 P07910          2 54502  3183 RBM47   HNRNPC
# # . with 195,528 more rows, and 3 more variables: p_wrong <dbl>,
# #   p_no_interaction <dbl>, p_interaction <dbl>
```

---

bioplex_hct116_1 *Downloads the BioPlex HCT116 version 1.0 interaction dataset*

Description

This dataset contains ~71,000 interactions detected in HCT116 cells using 5,522 baits. More details at https://bioplex.hms.harvard.edu/interactions.php.

Usage

```
bioplex_hct116_1()
```

Value

Data frame (tibble) with interactions.

See Also

- bioplex1
- bioplex2
- bioplex3
- bioplex_all

Examples

```
bioplex_interactions <- bioplex_hct116_1()
nrow(bioplex_interactions)
# [1] 70966
colnames(bioplex_interactions)
# [1] "GeneA"         "GeneB"        "UniprotA"   "UniprotB"
# [5] "SymbolA"       "SymbolB"      "p_wrong"    "p_no_interaction"
# [9] "p_interaction"
```

---

bma_motif_es *BMA motifs from a sequence of edges*

Description

These motifs can be added to a BMA canvas.

Usage

```
bma_motif_es(edge_seq, G, granularity = 2)
```

Arguments

edge_seq	An igraph edge sequence.
G	An igraph graph object.
granularity	Numeric: granularity value.

Value

Character: BMA motifs as a single string.

Examples

```
interactions <- import_omnipath_interactions(resources = 'ARN')
graph <- interaction_graph(interactions)
motifs <- bma_motif_es(igraph::E(graph)[1], graph)
```

`bma_motif_vs` *Prints a BMA motif to the screen from a sequence of nodes, which can be copy/pasted into the BMA canvas*

Description

Intended to parallel print_path_vs

Usage

```
bma_motif_vs(node_seq, G)
```

Arguments

`node_seq`	An igraph node sequence.
`G`	An igraph graph object.

Value

Character: BMA motifs as a single string.

Examples

```
interactions <- import_omnipath_interactions(resources = 'ARN')
graph <- interaction_graph(interactions)
bma_string <- bma_motif_vs(
    igraph::all_shortest_paths(
        graph,
        from = 'ULK1',
        to = 'ATG13'
    )$res,
    graph
)
```

common_name *Common (English) names of organisms*

Description

Common (English) names of organisms

Usage

```
common_name(name)
```

Arguments

name	Vector with any kind of organism name or identifier, can be also mixed type.

Value

Character vector with common (English) taxon names, NA if a name in the input could not be found.

See Also

- ncbi_taxid
- latin_name
- ensembl_name

Examples

```
common_name(c(10090, "cjacchus", "Vicugna pacos"))
# [1] "Mouse" "White-tufted-ear marmoset" "Alpaca"
```

consensuspathdb_download *Retrieves the ConsensusPathDB network*

Description

Compiles a table of binary interactions from ConsensusPathDB (http://cpdb.molgen.mpg.de/) and translates the UniProtKB ACs to Gene Symbols.

Usage

```
consensuspathdb_download(complex_max_size = 4, min_score = 0.9)
```

Arguments

`complex_max_size`
: Numeric: do not expand complexes with a higher number of elements than this. ConsensusPathDB does not contain conventional interactions but lists of participants, which might be members of complexes. Some records include dozens of participants and expanding them to binary interactions result thousands, sometimes hundreds of thousands of interactions from one single record. At the end, this process consumes >10GB of memory and results rather unusable data, hence it is recommended to limit the complex sizes at some low number.

`min_score`
: Numeric: each record in ConsensusPathDB comes with a confidence score, expressing the amount of evidences. The default value, a minimum score of 0.9 retains approx. the top 30 percent of the interactions.

Value

Data frame (tibble) with interactions.

Examples

```
## Not run:
cpdb_data <- consensuspathdb_download(
    complex_max_size = 1,
    min_score = .99
)
nrow(cpdb_data)
# [1] 252302
colnames(cpdb_data)
# [1] "databases"  "references" "uniprot_a"    "confidence"   "record_id"
# [6] "uniprot_b"  "in_complex" "genesymbol_a" "genesymbol_b"
cpdb_data
# # A tibble: 252,302 x 9
#    databases references uniprot_a confidence record_id uniprot_b in_com
#    <chr>     <chr>      <chr>          <dbl>     <int> <chr>     <lgl>
#  1 Reactome  NA         SUMF2_HU.      1             1 SUMF1_HU. TRUE
#  2 Reactome  NA         SUMF1_HU.      1             1 SUMF2_HU. TRUE
#  3 DIP,Reac. 22210847,. STIM1_HU.      0.998         2 TRPC1_HU. TRUE
#  4 DIP,Reac. 22210847,. TRPC1_HU.      0.998         2 STIM1_HU. TRUE
# # . with 252,292 more rows, and 2 more variables: genesymbol_a <chr>,
# #   genesymbol_b <chr

## End(Not run)
```

---

consensuspathdb_raw_table

Downloads interaction data from ConsensusPathDB

Description

Downloads interaction data from ConsensusPathDB

Usage

```
consensuspathdb_raw_table()
```

Value

Data frame (tibble) with interactions.

Examples

```
cpdb_raw <- consensuspathdb_raw_table()
```

---

curated_ligand_receptor_interactions

Curated ligand-receptor interactions

Description

The OmniPath *intercell* database annotates individual proteins and complexes, and we combine these annotations with network interactions on the client side, using `import_intercell_network`. The architecture of this database is complex, aiming to cover a broad range of knowledge on various levels of details and confidence. We can use the `intercell_consensus_filter` and `filter_intercell_network` functions for automated, data driven quality filtering, in order to enrich the cell-cell communication network in higher confidence interactions. However, for many users, a simple combination of the most established, expert curated ligand-receptor resources, provided by this function, fits better their purpose.

Usage

```
curated_ligand_receptor_interactions(
 curated_resources = c("Guide2Pharma", "HPMR", "ICELLNET", "Kirouac2010", "CellTalkDB",
    "CellChatDB", "connectomeDB2020"),
  cellphonedb = TRUE,
  cellinker = TRUE,
  talklr = TRUE,
  signalink = TRUE,
  ...
)
```

Arguments

curated_resources	Character vector of the resource names which are considered to be expert curated. You can include any post-translational network resource here, but if you include non ligand-receptor or non curated resources, the result will not fulfill the original intention of this function.
cellphonedb	Logical: include the curated interactions from *CellPhoneDB* (not the whole *CellPhoneDB* but a subset of it).
cellinker	Logical: include the curated interactions from *Cellinker* (not the whole *Cellinker* but a subset of it).
talklr	Logical: include the curated interactions from *talklr* (not the whole *talklr* but a subset of it).
signalink	Logical: include the ligand-receptor interactions from *SignaLink*. These are all expert curated.
...	Passed to import_post_translational_interactions: further parameters for the interaction data. Should not contain 'resources' argument as that would interfere with the downstream calls.

Details

Some resources are a mixture of curated and bulk imported interactions, and sometimes it's not trivial to separate these, we take care of these here. This function does not use the *intercell* database of OmniPath, but retrieves and filters a handful of network resources. The returned data frame has the layout of *interactions* (network) data frames, and the *source* and *target* partners implicitly correspond to *ligand* and *receptor*. The data frame shows all resources and references for all interactions, but each interaction is supported by at least one ligand-receptor resource which is supposed to based on expert curation in a ligand-receptor context.

Value

A data frame similar to *interactions* (network) data frames, the *source* and *target* partners being ligand and receptor, respectively.

See Also

- import_intercell_network
- filter_intercell_network
- annotated_network
- import_post_translational_interactions
- import_ligrecextra_interactions
- curated_ligrec_stats

Examples

```
lr <- curated_ligand_receptor_interactions()
lr
```

`curated_ligrec_stats` *Statistics about literature curated ligand-receptor interactions*

Description

Statistics about literature curated ligand-receptor interactions

Usage

```
curated_ligrec_stats(...)
```

Arguments

`...`	Passed to `curated_ligand_receptor_interactions`, determines the set of all curated L-R interactions which will be compared against each of the individual resources.

Details

The data frame contains the total number of interactions, the number of interactions which overlap with the set of curated interactions *(curated_overlap),* the number of interactions with literature references from the given resource *(literature)* and the number of interactions which are curated by the given resource *(curated_self)*. This latter we defined according to our best knowledge, in many cases it's not possible to distinguish curated interactions). All these numbers are also presented as a percent of the total. Importantly, here we consider interactions curated only if they've been curated in a cell-cell communication context.

Value

A data frame with estimated counts of curated ligand-receptor interactions for each L-R resource.

See Also

`curated_ligand_receptor_interactions`

Examples

```
clr <- curated_ligrec_stats()
clr
```

descendants *All descendants in the ontology tree*

Description

Starting from the selected nodes, recursively walks the ontology tree until it reaches the leaf nodes. Collects all visited nodes, which are the descendants (children) of the starting nodes.

Usage

```
descendants(
  terms,
  db_key = "go_basic",
  ids = TRUE,
 relations = c("is_a", "part_of", "occurs_in", "regulates", "positively_regulates",
    "negatively_regulates")
)
```

Arguments

terms	Character vector of ontology term IDs or names. A mixture of IDs and names can be provided.
db_key	Character: key to identify the ontology database. For the available keys see omnipath_show_db.
ids	Logical: whether to return IDs or term names.
relations	Character vector of ontology relation types. Only these relations will be used.

Details

Note: this function relies on the database manager, the first call might take long because of the database load process. Subsequent calls within a short period should be faster. See get_ontology_db.

Value

Character vector of ontology IDs. If the input terms are all leaves NULL is returned. The starting nodes won't be included in the result unless some of them are descendants of other starting nodes.

Examples

```
descendants('GO:0005035', ids = FALSE)
# [1] "tumor necrosis factor-activated receptor activity"
# [2] "TRAIL receptor activity"
# [3] "TNFSF11 receptor activity"
```

ensembl_dataset *Ensembl dataset name from organism*

Description

Ensembl dataset name from organism

Usage

```
ensembl_dataset(organism)
```

Arguments

organism	Character or integer: an organism (taxon) name or identifier. If an Ensembl dataset name is provided

Value

Character: name of an ensembl dataset.

Examples

```
ensembl_dataset(10090)
# [1] "mmusculus_gene_ensembl"
```

ensembl_id_mapping_table *Identifier translation table from Ensembl*

Description

Identifier translation table from Ensembl

Usage

```
ensembl_id_mapping_table(to, from = "uniprot", organism = 9606)
```

Arguments

to	Character or symbol: target ID type. See Details for possible values.
from	Character or symbol: source ID type. See Details for possible values.
organism	Character or integer: NCBI Taxonomy ID or name of the organism (by default 9606 for human).

Details

The arguments `to` and `from` can be provided either as character or as symbol (NSE). Their possible values are either Ensembl attribute names or synonyms listed at `translate_ids`.

Value

A data frame (tibble) with columns ‘From‘ and ‘To‘.

See Also

- `translate_ids`
- `uniprot_full_id_mapping_table`
- `uniprot_id_mapping_table`

Examples

```
ensp_up <- ensembl_id_mapping_table("ensp")
ensp_up
# # A tibble: 119,129 × 2
#    From   To
#    <chr>  <chr>
#  1 P03886 ENSP00000354687
#  2 P03891 ENSP00000355046
#  3 P00395 ENSP00000354499
#  4 P00403 ENSP00000354876
#  5 P03928 ENSP00000355265
# # . with 119,124 more rows
```

`ensembl_id_type` *Ensembl identifier type label*

Description

Ensembl identifier type label

Usage

```
ensembl_id_type(label)
```

Arguments

`label`	Character: an ID type label, as shown in the table at `translate_ids`

Value

Character: the Ensembl specific ID type label, or the input unchanged if it could not be translated (still might be a valid identifier name). These labels should be valid Ensembl attribute names, directly usable in Ensembl queries.

See Also

- uniprot_id_type
- uploadlists_id_type

Examples

```
ensembl_id_type("uniprot")
# [1] "uniprotswissprot"
```

`ensembl_name` *Ensembl identifiers of organisms*

Description

Ensembl identifiers of organisms

Usage

```
ensembl_name(name)
```

Arguments

`name`	Vector with any kind of organism name or identifier, can be also mixed type.

Value

Character vector with Ensembl taxon names, NA if a name in the input could not be found.

See Also

- ncbi_taxid
- common_name
- latin_name

Examples

```
ensembl_name(c(9606, "cat", "dog"))
# [1] "hsapiens" "fcatus" "clfamiliaris"
ensembl_name(c("human", "kitten", "cow"))
# [1] "hsapiens" NA  "btaurus"
```

ensembl_organisms *Organism names and identifiers from Ensembl*

Description

A table with various taxon names and identifiers: English common names, latin (scientific) names, Ensembl organism IDs and NCBI taxonomy IDs.

Usage

```
ensembl_organisms()
```

Value

A data frame with the above mentioned columns.

Examples

```
ens_org <- ensembl_organisms()
ens_org
```

ensembl_organisms_raw *Table of Ensembl organisms*

Description

A table with various taxon IDs and metadata about related Ensembl database contents, as shown at https://www.ensembl.org/info/about/species.html. The "Taxon ID" column contains the NCBI Taxonomy identifiers.

Usage

```
ensembl_organisms_raw()
```

Value

The table described above as a data frame.

Examples

```
ens_org <- ensembl_organisms_raw()
ens_org
```

`ensembl_orthology` *Orthologous gene pairs from Ensembl*

Description

Orthologous gene pairs from Ensembl

Usage

```
ensembl_orthology(
  organism_a = 9606,
  organism_b = 10090,
  attrs_a = NULL,
  attrs_b = NULL,
  colrename = TRUE
)
```

Arguments

`organism_a`	Character or integer: organism name or identifier for the left side organism. We query the Ensembl dataset of this organism and add the orthologues of the other organism to it. Ideally this is the organism you translate from.
`organism_b`	Character or integer: organism name or identifier for the right side organism. We add orthology information of this organism to the gene records of the left side organism.
`attrs_a`	Further attributes about organism_a genes. Will be simply added to the attributes list.
`attrs_b`	Further attributes about organism_b genes (orthologues). The available attributes are: "associated_gene_name", "chromosome", "chrom_start", "chrom_end", "wga_coverage", "goc_score", "perc_id_r1", "perc_id", "subtype". Attributes included by default: "ensembl_gene", "ensembl_peptide", "canonical_transcript_protein", "orthology_confidence" and "orthology_type".
`colrename`	Logical: replace prefixes from organism_b attribute column names, so the returned table always have the same column names, no matter the organism. E.g. for mouse these columns all have the prefix "mmusculus_homolog_", which this option changes to "b_".

Details

Only the records with orthology information are returned. The order of columns is the following: defaults of organism_a, extra attributes of organism_b, defaults of organism_b, extra attributes of organism_b.

Examples

```
## Not run:
sffish <- ensembl_orthology(
    organism_b = 'Siamese fighting fish',
    attrs_a = 'external_gene_name',
    attrs_b = 'associated_gene_name'
)
sffish
# # A tibble: 175,608 × 10
#    ensembl_gene_id ensembl_transcript_id ensembl_peptide. external_gene_n.
#    <chr>           <chr>                 <chr>            <chr>
#  1 ENSG00000277196 ENST00000621424       ENSP00000481127  NA
#  2 ENSG00000277196 ENST00000615165       ENSP00000482462  NA
#  3 ENSG00000278817 ENST00000613204       ENSP00000482514  NA
#  4 ENSG00000274847 ENST00000400754       ENSP00000478910  MAFIP
#  5 ENSG00000273748 ENST00000612919       ENSP00000479921  NA
# # . with 175,603 more rows, and 6 more variables:
# #   b_ensembl_peptide <chr>, b_ensembl_gene <chr>,
# #   b_orthology_type <chr>, b_orthology_confidence <dbl>,
# #   b_canonical_transcript_protein <chr>, b_associated_gene_name <chr>
#

## End(Not run)
```

---

enzsub_graph *Enzyme-substrate graph*

Description

Transforms the a data frame with enzyme-substrate relationships (obtained by `import_omnipath_enzsub`) to an igraph graph object.

Usage

```
enzsub_graph(enzsub)
```

Arguments

enzsub	Data frame created by `import_omnipath_enzsub`

Value

An igraph directed graph object.

See Also

- `import_omnipath_enzsub`
- `giant_component`
- `find_all_paths`

Examples

```
enzsub <- import_omnipath_enzsub(resources = c('PhosphoSite', 'SIGNOR'))
enzsub_g <- enzsub_graph(enzsub = enzsub)
```

`evex_download` *Interactions from the EVEX database*

Description

Downloads interactions from EVEX, a versatile text mining resource (http://evexdb.org). Translates the Entrez Gene IDs to Gene Symbols and combines the interactions and references into a single data frame.

Usage

```
evex_download(
    min_confidence = NULL,
    remove_negatives = TRUE,
    top_confidence = NULL
)
```

Arguments

`min_confidence`	Numeric: a threshold for confidence scores. EVEX confidence scores span roughly from -3 to 3. By providing a numeric value in this range the lower confidence interactions can be removed. If NULL no filtering performed.
`remove_negatives`	Logical: remove the records with the "negation" attribute set.
`top_confidence`	Confidence cutoff as quantile (a number between 0 and 1). If NULL no filtering performed.

Value

Data frame (tibble) with interactions.

Examples

```
evex_interactions <- evex_download()
evex_interactions
# # A tibble: 368,297 x 13
#   general_event_id source_entrezge. target_entrezge. confidence negation
#              <dbl> <chr>            <chr>                 <dbl>    <dbl>
# 1               98 8651             6774                -1.45         0
# 2              100 8431             6774                -1.45         0
# 3              205 6261             6263                 0.370        0
# 4              435 1044             1045                -1.09         0
# . with 368,287 more rows, and 8 more variables: speculation <dbl>,
```

```
#   coarse_type <chr>, coarse_polarity <chr>, refined_type <chr>,
#   refined_polarity <chr>, source_genesymbol <chr>,
#   target_genesymbol <chr>, references <chr>
```

`extra_attrs` *Extra attribute names in an interaction data frame*

Description

Interaction data frames might have an ‘extra_attrs‘ column if this field has been requested in the query by passing the ‘fields = ’extra_attrs’ argument. This column contains resource specific attributes for the interactions. The names of the attributes consist of the name of the resource and the name of the attribute, separated by an underscore. This function returns the names of the extra attributes available in the provided data frame.

Usage

```
extra_attrs(data)
```

Arguments

`data`	An interaction data frame, as provided by any of the `import...interactions` functions.

Value

Character: the names of the extra attributes in the data frame.

See Also

- `extra_attrs_to_cols`
- `has_extra_attrs`
- `with_extra_attrs`
- `filter_extra_attrs`
- `extra_attr_values`

Examples

```
i <- import_omnipath_interactions(fields = 'extra_attrs')
extra_attrs(i)
```

`extra_attrs_to_cols` *New columns from extra attributes*

Description

New columns from extra attributes

Usage

```
extra_attrs_to_cols(data, ..., flatten = FALSE, keep_empty = TRUE)
```

Arguments

`data`	An interaction data frame.
`...`	The names of the extra attributes; NSE is supported. Custom column names can be provided as argument names.
`flatten`	Logical: unnest the list column even if some records have multiple values for the attributes; these will yield multiple records in the resulted data frame.
`keep_empty`	Logical: if 'flatten' is 'TRUE', shall we keep the records which do not have the attribute?

Value

Data frame with the new column created; the new column is list type if one interaction might have multiple values of the attribute, or character type if

See Also

- `extra_attrs`
- `has_extra_attrs`
- `with_extra_attrs`
- `filter_extra_attrs`
- `extra_attr_values`

Examples

```
i <- import_omnipath_interactions(fields = 'extra_attrs')
extra_attrs_to_cols(i, Cellinker_type, Macrophage_type)
extra_attrs_to_cols(
    i,
    Cellinker_type,
    Macrophage_type,
    flatten = TRUE,
    keep_empty = FALSE
)
```

`extra_attr_values` *Possible values of an extra attribute*

Description

Extracts all unique values of an extra attribute occuring in this data frame.

Usage

```
extra_attr_values(data, key)
```

Arguments

`data`	An interaction data frame with *extra_attrs* column.
`key`	The name of an extra attribute.

Details

Note, at the end we unlist the result, which means it works well for attributes which are atomic vectors but gives not so useful result if the attribute values are more complex objects. As the time of writing this, no such complex extra attribute exist in OmniPath.

Value

A vector, most likely character, with the unique values of the extra attribute occuring in the data frame.

See Also

- `extra_attrs_to_cols`
- `has_extra_attrs`
- `with_extra_attrs`
- `filter_extra_attrs`
- `extra_attrs`

Examples

```
op <- import_omnipath_interactions(fields = 'extra_attrs')
extra_attr_values(op, SIGNOR_mechanism)
```

filter_by_resource *Filters OmniPath data by resources*

Description

Keeps only those records which are supported by any of the resources of interest.

Usage

```
filter_by_resource(data, resources = NULL)
```

Arguments

data	A data frame downloaded from the OmniPath web service (interactions, enzyme-substrate or complexes).
resources	Character vector with resource names to keep.

Value

The data frame filtered.

Examples

```
interactions <- import_omnipath_interactions()
signor <- filter_by_resource(interactions, resources = 'SIGNOR')
```

filter_extra_attrs *Filter interactions by extra attribute values*

Description

Filter interactions by extra attribute values

Usage

```
filter_extra_attrs(data, ..., na_ok = TRUE)
```

Arguments

data	An interaction data frame with *extra_attrs* column.
...	Extra attribute names and values. The contents of the extra attribute *name* for each record will be checked against the values provided. The check by default is a set intersection: if any element is common between the user provided values and the values of the extra attribute for the record, the record will be matched. Alternatively, any value can be a custom function which accepts the value of the extra attribute and returns a single logical value. Finally, if the extra attribute name starts with a dot, the result of the check will be negated.
na_ok	Logical: keep the records which do not have the extra attribute. Typically these are the records which are not from the resource providing the extra attribute.

Value

The input data frame with records removed according to the filtering criteria.

See Also

- extra_attrs
- has_extra_attrs
- extra_attrs_to_cols
- with_extra_attrs
- extra_attr_values

Examples

```
cl <- import_post_translational_interactions(
    resources = 'Cellinker',
    fields = 'extra_attrs'
)
# Only cell adhesion interactions from Cellinker
filter_extra_attrs(cl, Cellinker_type = 'Cell adhesion')

op <- import_omnipath_interactions(fields = 'extra_attrs')
# Any mechanism except phosphorylation
filter_extra_attrs(op, .SIGNOR_mechanism = 'phosphorylation')
```

---

filter_intercell *Filter intercell annotations*

Description

Filters a data frame retrieved by import_omnipath_intercell.

Usage

```
filter_intercell(
  data,
  categories = NULL,
  resources = NULL,
  parent = NULL,
  scope = NULL,
  aspect = NULL,
  source = NULL,
  transmitter = NULL,
  receiver = NULL,
  secreted = NULL,
  plasma_membrane_peripheral = NULL,
  plasma_membrane_transmembrane = NULL,
  proteins = NULL,
  causality = NULL,
  topology = NULL,
  ...
)
```

Arguments

data	An intercell annotation data frame as provided by import_omnipath_intercell.
categories	Character: allow only these values in the category column.
resources	Character: allow records only from these resources.
parent	Character: filter for records with these parent categories.
scope	Character: filter for records with these annotation scopes. Possible values are generic and specific.
aspect	Character: filter for records with these annotation aspects. Possible values are functional and locational.
source	Character: filter for records with these annotation sources. Possible values are composite and resource_specific.
transmitter	Logical: if TRUE only transmitters, if FALSE only non-transmitters will be selected, if NULL it has no effect.
receiver	Logical: works the same way as transmitters.
secreted	Logical: works the same way as transmitters.
plasma_membrane_peripheral	Logical: works the same way as transmitters.
plasma_membrane_transmembrane	Logical: works the same way as transmitters.
proteins	Character: filter for annotations of these proteins. Gene symbols or UniProt IDs can be used.
causality	Character: filter for records with these causal roles. Possible values are transmitter and receiver. The filter applied simultaneously to the transmitter and receiver arguments, it's just a different notation for the same thing.

topology	Character: filter for records with these localization topologies. Possible values are secreced, plasma_membrane_peripheral and plasma_membrane_transmembrane; the shorter notations sec, pmp and pmtm can be used. Has the same effect as the logical type arguments, just uses a different notation.
...	Ignored.

Value

The intercell annotation data frame filtered according to the specified conditions.

Examples

```
ic <- import_omnipath_intercell()
ic <- filter_intercell(
    ic,
    transmitter = TRUE,
    secreted = TRUE,
    scope = "specific"
)
```

---

filter_intercell_network

Quality filter an intercell network

Description

The intercell database of OmniPath covers a very broad range of possible ways of cell to cell communication, and the pieces of information, such as localization, topology, function and interaction, are combined from many, often independent sources. This unavoidably result some weird and unexpected combinations which are false positives in the context of intercellular communication. import_intercell_network provides a shortcut (high_confidence) to do basic quality filtering. For custom filtering or experimentation with the parameters we offer this function.

Usage

```
filter_intercell_network(
  network,
  transmitter_topology = c("secreted", "plasma_membrane_transmembrane",
    "plasma_membrane_peripheral"),
  receiver_topology = "plasma_membrane_transmembrane",
  min_curation_effort = 2,
  min_resources = 1,
  min_references = 0,
  min_provenances = 1,
  consensus_percentile = 50,
  loc_consensus_percentile = 30,
  ligand_receptor = FALSE,
```

```
  simplify = FALSE,
  unique_pairs = FALSE,
  omnipath = TRUE,
  ligrecextra = TRUE,
  kinaseextra = FALSE,
  pathwayextra = FALSE,
  ...
)
```

Arguments

`network`	An intercell network data frame, as provided by `import_intercell_network`, without `simplify`.
`transmitter_topology`	Character vector: topologies allowed for the entities in transmitter role. Abbreviations allowed: "sec", "pmtm" and "pmp".
`receiver_topology`	Same as `transmitter_topology` for the entities in the receiver role.
`min_curation_effort`	Numeric: a minimum value of curation effort (resource-reference pairs) for network interactions. Use zero to disable filtering.
`min_resources`	Numeric: minimum number of resources for interactions. The value 1 means no filtering.
`min_references`	Numeric: minimum number of references for interactions. Use zero to disable filtering.
`min_provenances`	Numeric: minimum number of provenances (either resources or references) for interactions. Use zero or one to disable filtering.
`consensus_percentile`	Numeric: percentile threshold for the consensus score of generic categories in intercell annotations. The consensus score is the number of resources supporting the classification of an entity into a category based on combined information of many resources. Here you can apply a cut-off, keeping only the annotations supported by a higher number of resources than a certain percentile of each category. If `NULL` no filtering will be performed. The value is either in the 0-1 range, or will be divided by 100 if greater than 1. The percentiles will be calculated against the generic composite categories and then will be applied to their resource specific annotations and specific child categories.
`loc_consensus_percentile`	Numeric: similar to `consensus_percentile` for major localizations. For example, with a value of 50, the secreted, plasma membrane transmembrane or peripheral attributes will be `TRUE` only where at least 50 percent of the resources support these.
`ligand_receptor`	Logical. If `TRUE`, only *ligand* and *receptor* annotations will be used instead of the more generic *transmitter* and *receiver* categories.

simplify	Logical: keep only the most often used columns. This function combines a network data frame with two copies of the intercell annotation data frames, all of them already having quite some columns. With this option we keep only the names of the interacting pair, their intercellular communication roles, and the minimal information of the origin of both the interaction and the annotations.
unique_pairs	Logical: instead of having separate rows for each pair of annotations, drop the annotations and reduce the data frame to unique interacting pairs. See unique_intercell_network for details.
omnipath	Logical: shortcut to include the *omnipath* dataset in the interactions query.
ligrecextra	Logical: shortcut to include the *ligrecextra* dataset in the interactions query.
kinaseextra	Logical: shortcut to include the *kinaseextra* dataset in the interactions query.
pathwayextra	Logical: shortcut to include the *pathwayextra* dataset in the interactions query.
...	If simplify or unique_pairs is TRUE, additional column names can be passed here to dplyr::select on the final data frame. Otherwise ignored.

Value

An intercell network data frame filtered.

See Also

- import_intercell_network
- unique_intercell_network
- simplify_intercell_network

Examples

```
icn <- import_intercell_network()
icn_f <- filter_intercell_network(
    icn,
    consensus_percentile = 75,
    min_provenances = 3,
    simplify = TRUE
)
```

---

find_all_paths *All paths between two groups of vertices*

Description

Finds all paths up to length ‘maxlen‘ between specified groups of vertices. This function is needed only becaues igraph‘s ‘all_shortest_paths‘ finds only the shortest, not any path up to a defined length.

Usage

```
find_all_paths(
    graph,
    start,
    end,
    attr = NULL,
    mode = 'OUT',
    maxlen = 2,
    progress = TRUE
)
```

Arguments

graph	An igraph graph object.
start	Integer or character vector with the indices or names of one or more start vertices.
end	Integer or character vector with the indices or names of one or more end vertices.
attr	Character: name of the vertex attribute to identify the vertices by. Necessary if ‘start‘ and ‘end‘ are not igraph vertex ids but for example vertex names or labels.
mode	Character: IN, OUT or ALL. Default is OUT.
maxlen	Integer: maximum length of paths in steps, i.e. if maxlen = 3, then the longest path may consist of 3 edges and 4 nodes.
progress	Logical: show a progress bar.

Value

List of vertex paths, each path is a character or integer vector.

See Also

- interaction_graph
- enzsub_graph
- giant_component

Examples

```
interactions <- import_omnipath_interactions()
graph <- interaction_graph(interactions)
paths <- find_all_paths(
    graph = graph,
    start = c('EGFR', 'STAT3'),
    end = c('AKT1', 'ULK1'),
    attr = 'name'
)
```

get_annotation_resources

Retrieves a list of available resources in the annotations database of OmniPath

Description

Get the names of the resources from https://omnipath.org/annotations.

Usage

```
get_annotation_resources(dataset = NULL, ...)
```

Arguments

dataset	ignored for this query type
...	optional additional arguments

Value

character vector with the names of the annotation resources

See Also

- get_resources
- import_omnipath_annotations

Examples

```
get_annotation_resources()
```

get_complex_genes

Get all the molecular complexes for a given gene(s)

Description

This function returns all the molecular complexes where an input set of genes participate. User can choose to retrieve every complex where any of the input genes participate or just retrieve these complexes where all the genes in input set participate together.

Usage

```
get_complex_genes(
  complexes = import_omnipath_complexes(),
  select_genes,
  total_match = FALSE
)
```

Arguments

complexes	complexes data frame (obtained using import_omnipath_complexes)
select_genes	vector containing the genes for whom complexes will be retrieved (hgnc format).
total_match	[default=FALSE] logical indicating if the user wants to get all the complexes where any of the input genes participate (FALSE) or to get only the complexes where all the input genes participate together (TRUE).

Value

Data frame of complexes

See Also

import_omnipath_complexes

Examples

```
complexes <- import_omnipath_complexes(
    filter_databases = c("CORUM", "hu.MAP")
)
query_genes <- c("LMNA", "BANF1")
complexes_query_genes <- get_complex_genes(complexes, query_genes)
```

---

get_complex_resources *Retrieve a list of complex resources available in Omnipath*

Description

Get the names of the resources from https://omnipath.org/complexes

Usage

```
get_complex_resources(dataset = NULL)
```

Arguments

dataset	ignored for this query type

Value

character vector with the names of the databases

See Also

- get_resources
- import_omnipath_complexes

Examples

```
get_complex_resources()
```

`get_db` *Access a built in database*

Description

Databases are resources which might be costly to load but can be used many times by functions which usually automatically load and retrieve them from the database manager. Each database has a lifetime and will be unloaded automatically upon expiry.

Usage

```
get_db(key, param = NULL, reload = FALSE, ...)
```

Arguments

`key`	Character: the key of the database to load. For a list of available keys see `omnipath_show_db`.
`param`	List: override the defaults or pass further parameters to the database loader function. See the loader functions and their default parameters in `omnipath_show_db`. If the database is already loaded with different parameters it will be reloaded with the new parameters only if the `reload` option is `TRUE`.
`reload`	Reload the database if `param` passed here is different from the parameters used the last time the database was loaded. If different functions with different parameters access the database repeatedly and request reload the frequent reloads might cost substantial time and resource use.
`...`	Arguments for the loader function of the database. These override the default arguments.

Value

An object with the database contents. The exact format depends on the database, most often it is a data frame or a list.

See Also

`omnipath_show_db`.

Examples

```
goslim <- get_db('go_slim')
```

get_enzsub_resources *Retrieves a list of enzyme-substrate resources available in OmniPath*

Description

Get the names of the enzyme-substrate relationship resources available in https://omnipath.org/enzsub

Usage

```
get_enzsub_resources(dataset = NULL)
```

Arguments

dataset	ignored for this query type

Value

character vector with the names of the enzyme-substrate resources

See Also

- get_resources
- import_omnipath_enzsub

Examples

```
get_enzsub_resources()
```

get_interaction_resources *Retrieve a list of interaction resources available in Omnipath*

Description

Gets the names of the resources from https://omnipath.org/interactions.

Usage

```
get_interaction_resources(dataset = NULL)
```

Arguments

dataset	a dataset within the interactions query type. Currently available datasets are 'omnipath', 'kinaseextra', 'pathwayextra', 'ligrecextra', 'dorothea', 'tf_target', 'tf_mirna', 'mirnatarget' and 'lncrna_mrna'

Value

character vector with the names of the interaction databases

See Also

- get_resources
- import_all_interactions
- import_omnipath_interactions
- import_pathwayextra_interactions
- import_kinaseextra_interactions
- import_ligrecextra_interactions
- import_mirnatarget_interactions
- import_dorothea_interactions

Examples

```
get_interaction_resources()
```

---

get_intercell_categories

Categories in the intercell database of OmniPath

Description

Retrieves a list of categories from https://omnipath.org/intercell.

Usage

```
get_intercell_categories()
```

Value

character vector with the different intercell categories

See Also

- import_omnipath_intercell
- get_intercell_generic_categories

Examples

```
get_intercell_categories()
```

get_intercell_generic_categories

Retrieves a list of the generic categories in the intercell database of OmniPath

Description

Retrieves a list of the generic categories from https://omnipath.org/intercell.

Usage

```
get_intercell_generic_categories()
```

Value

character vector with the different intercell main classes

See Also

- import_omnipath_intercell
- get_intercell_categories

Examples

```
get_intercell_generic_categories()
```

get_intercell_resources

Retrieves a list of intercellular communication resources available in OmniPath

Description

Retrieves a list of the databases from https://omnipath.org/intercell.

Usage

```
get_intercell_resources(dataset = NULL)
```

Arguments

dataset	ignored at this query type

Value

character vector with the names of the databases

See Also

- get_resources
- import_omnipath_intercell

Examples

```
get_intercell_resources()
```

`get_ontology_db` *Access an ontology database*

Description

Retrieves an ontology database with relations in the desired data structure. The database is automatically loaded and the requested data structure is constructed if necessary. The databases stay loaded up to a certain time period (see the option `omnipath.db_lifetime`). Hence the first one of repeated calls to this function might take long and the subsequent ones should be really quick.

Usage

```
get_ontology_db(key, rel_fmt = "tbl", child_parents = TRUE)
```

Arguments

`key`	Character: key of the ontology database. For the available keys see omnipath_show_db.
`rel_fmt`	Character: the data structure of the ontology relations. Posible values are 1) "tbl" a data frame, 2) "lst" a list or 3) "gra" a graph.
`child_parents`	Logical: whether the ontology relations should point from child to parents (`TRUE`) or from parent to children (`FALSE`).

Value

A list with the following elements: 1) "names" a table with term IDs and names; 2) "namespaces" a table to connect term IDs and namespaces they belong to; 3) "relations" a table with relations between terms and their parent terms; 4) "subsets" a table with terms and the subsets they are part of; 5) "obsolete" character vector with all the terms labeled as obsolete.

See Also

- omnipath_show_db
- get_db

Examples

```
go <- get_ontology_db('go_basic', child_parents = FALSE)
```

get_resources *Retrieve the available resources for a given query type*

Description

Collects the names of the resources available in OmniPath for a certain query type and optionally for a dataset within that.

Usage

```
get_resources(query_type, datasets = NULL, generic_categories = NULL)
```

Arguments

query_type	one of the query types ‘interactions‘, ‘enz_sub‘, ‘complexes‘, ‘annotations‘ or ‘intercell‘
datasets	currently within the ‘interactions‘ query type only, multiple datasets are available: ‘omnipath‘, ‘kinaseextra‘, ‘pathwayextra‘, ‘ligrecextra‘, ‘dorothea‘, ‘tf_target‘, ‘tf_mirna‘, ‘mirnatarget‘ and ‘lncrna_mrna‘.
generic_categories	for the ‘intercell‘ query type, restrict the search for some generic categories e.g. ‘ligand‘ or ‘receptor‘.

Value

a character vector with resource names

Examples

```
get_resources(query_type = 'interactions')
```

get_signed_ptms *Signs for enzyme-substrate interactions*

Description

Enzyme-substrate data does not contain sign (activation/inhibition), we generate this information based on the interaction network.

Usage

```
get_signed_ptms(
  enzsub = import_omnipath_enzsub(),
  interactions = import_omnipath_interactions()
)
```

Arguments

enzsub	Enzyme-substrate data frame generated by import_omnipath_enzsub
interactions	interaction data frame generated by import_omnipath_interactions

Value

Data frame of enzyme-substrate relationships with is_inhibition and is_stimulation columns.

See Also

- import_omnipath_enzsub
- import_omnipath_interactions

Examples

```
enzsub <- import_omnipath_enzsub(resources = c('PhosphoSite', 'SIGNOR'))
interactions <- import_omnipath_interactions()
enzsub <- get_signed_ptms(enzsub, interactions)
```

---

giant_component *Giant component of a graph*

Description

For an igraph graph object returns its giant component.

Usage

```
giant_component(graph)
```

Arguments

graph	An igraph graph object.

Value

An igraph graph object containing only the giant component.

Examples

```
interactions <- import_post_translational_interactions()
graph <- interaction_graph(interactions)
graph_gc <- giant_component(graph)
```

`go_annot_download` *Gene annotations from Gene Ontology*

Description

Gene Ontology is an ontology of gene subcellular localizations, molecular functions and involvement in biological processes. Gene products across many organisms are annotated with the ontology terms. This function downloads the gene-ontology term associations for certain model organisms or all organisms. For a description of the columns see http://geneontology.org/docs/go-annotation-file-gaf-format-2.2/.

Usage

```
go_annot_download(organism = "human", aspects = c("C", "F", "P"), slim = NULL)
```

Arguments

`organism`	Character: either "chicken", "cow", "dog", "human", "pig" or "uniprot_all".
`aspects`	Character vector with some of the following elements: "C" (cellular component), "F" (molecular function) and "P" (biological process). Gene Ontology is three separate ontologies called as three aspects. By this parameter you can control which aspects to include in the output.
`slim`	Character: if not `NULL`, the name of a GOsubset (slim). instead of the full GO annotation, the slim annotation will be returned. See details at `go_annot_slim`. If `TRUE`, the "generic" slim will be used.

Value

A tibble (data frame) of annotations as it is provided by the database

Examples

```
goa_data <- go_annot_download()
goa_data
# # A tibble: 606,840 x 17
#    db       db_object_id db_object_symbol qualifier go_id   db_ref
#    <fct>    <chr>        <chr>            <fct>     <chr>   <chr>
#  1 UniProt. A0A024RBG1   NUDT4B           NA        GO:000. GO_REF:00.
#  2 UniProt. A0A024RBG1   NUDT4B           NA        GO:000. GO_REF:00.
#  3 UniProt. A0A024RBG1   NUDT4B           NA        GO:004. GO_REF:00.
#  4 UniProt. A0A024RBG1   NUDT4B           NA        GO:005. GO_REF:00.
#  5 UniProt. A0A024RBG1   NUDT4B           NA        GO:005. GO_REF:00.
# # . with 606,830 more rows, and 11 more variables:
# #   evidence_code <fct>, with_or_from <chr>, aspect <fct>,
# #   db_object_name <chr>, db_object_synonym <chr>,
# #   db_object_type <fct>, taxon <fct>, date <date>,
# #   assigned_by <fct>, annotation_extension <chr>,
# #   gene_product_from_id <chr>
```

`go_annot_slim` *GO slim gene annotations*

Description

GO slims are subsets of the full GO which "give a broad overview of the ontology content without the detail of the specific fine grained terms". In order to annotate genes with GO slim terms, we take the annotations and search all ancestors of the terms up to the root of the ontology tree. From the ancestors we select the terms which are part of the slim subset.

Usage

```
go_annot_slim(
  organism = "human",
  slim = "generic",
  aspects = c("C", "F", "P"),
  cache = TRUE
)
```

Arguments

`organism`	Character: either "chicken", "cow", "dog", "human", "pig" or "uniprot_all".
`slim`	Character: the GO subset (GO slim) name. Available GO slims are: "agr" (Alliance for Genomics Resources), "generic", "aspergillus", "candida", "drosophila", "chembl", "metagenomic", "mouse", "plant", "pir" (Protein Information Resource), "pombe" and "yeast".
`aspects`	Character vector with some of the following elements: "C" (cellular component), "F" (molecular function) and "P" (biological process). Gene Ontology is three separate ontologies called as three aspects. By this parameter you can control which aspects to include in the output.
`cache`	Logical: Load the result from cache if available.

Details

Building the GO slim is resource intensive in its current implementation. For human annotation and generic GO slim it might take around 20 minutes. The result is saved into the cache so next time loading the data from there is really quick. If the `cache` option is `FALSE` the data will be built fresh (the annotation and ontology files still might come from cache), and the newly build GO slim will overwrite the cache instance.

Value

A tibble (data frame) of genes annotated with ontology terms in in the GO slim (subset).

See Also

- go_annot_download
- go_ontology_download
- get_db

Examples

```
## Not run:
goslim <- go_annot_slim(organism = 'human', slim = 'generic')
goslim
# # A tibble: 276,371 x 8
#    db     db_object_id db_object_symbol go_id aspect db_object_name
#    <fct>  <chr>        <chr>            <chr> <fct>  <chr>
#  1 UniPr. A0A024RBG1   NUDT4B           GO:0. F      Diphosphoinosito.
#  2 UniPr. A0A024RBG1   NUDT4B           GO:0. F      Diphosphoinosito.
#  3 UniPr. A0A024RBG1   NUDT4B           GO:0. C      Diphosphoinosito.
#  4 UniPr. A0A024RBG1   NUDT4B           GO:0. C      Diphosphoinosito.
#  5 UniPr. A0A024RBG1   NUDT4B           GO:0. C      Diphosphoinosito.
# # . with 276,366 more rows, and 2 more variables:
# #   db_object_synonym <chr>, db_object_type <fct>

## End(Not run)
```

---

go_ontology_download *The Gene Ontology tree*

Description

The Gene Ontology tree

Usage

```
go_ontology_download(
  basic = TRUE,
  tables = TRUE,
  subset = NULL,
 relations = c("is_a", "part_of", "occurs_in", "regulates", "positively_regulates",
    "negatively_regulates")
)
```

Arguments

basic	Logical: use the basic or the full version of GO. As written on the GO home page: "the basic version of the GO is filtered such that the graph is guaranteed to be acyclic and annotations can be propagated up the graph. The relations included are is a, part of, regulates, negatively regulates and positively regulates. This version excludes relationships that cross the 3 GO hierarchies. This version should be used with most GO-based annotation tools."

tables	In the result return data frames or nested lists. These later can be converted to each other if necessary. However converting from table to list is faster.
subset	Character: the GO subset (GO slim) name. GO slims are subsets of the full GO which "give a broad overview of the ontology content without the detail of the specific fine grained terms". This option, if not NULL, overrides the basic parameter. Available GO slims are: "agr" (Alliance for Genomics Resources), "generic", "aspergillus", "candida", "drosophila", "chembl", "metagenomic", "mouse", "plant", "pir" (Protein Information Resource), "pombe" and "yeast".
relations	Character vector: the relations to include in the processed data.

Value

A list with the following elements: 1) "names" a list with terms as names and names as values; 2) "namespaces" a list with terms as names and namespaces as values; 3) "relations" a list with relations between terms: terms are keys, values are lists with relations as names and character vectors of related terms as values; 4) "subsets" a list with terms as keys and character vectors of subset names as values (or NULL if the term does not belong to any subset); 5) "obsolete" character vector with all the terms labeled as obsolete. If the tables parameter is TRUE, "names", "namespaces", "relations" and "subsets" will be data frames (tibbles).

Examples

```
# retrieve the generic GO slim, a small subset of the full ontology
go <- go_ontology_download(subset = 'generic')
```

---

guide2pharma_download *Downloads interactions from the Guide to Pharmacology database*

Description

Downloads ligand-receptor interactions from the Guide to Pharmacology (IUPHAR/BPS) database (https://www.guidetopharmacology.org/).

Usage

```
guide2pharma_download()
```

Value

A tibble (data frame) of interactions as it is provided by the database

Examples

```
g2p_data <- guide2pharma_download()
g2p_data
# # A tibble: 21,586 x 38
#    target target_id target_gene_sym. target_uniprot target_ensembl_.
#    <chr>      <dbl> <chr>            <chr>          <chr>
#  1 12S-L.      1387 ALOX12           P18054         ENSG00000108839
#  2 15-LO.      1388 ALOX15           P16050         ENSG00000161905
#  3 15-LO.      1388 ALOX15           P16050         ENSG00000161905
#  4 15-LO.      1388 ALOX15           P16050         ENSG00000161905
# # . with 21,576 more rows, and 33 more variables: target_ligand <chr>,
# #   target_ligand_id <chr>, target_ligand_gene_symbol <chr>,
# ... (truncated)
```

`harmonizome_download` *Downloads a Harmonizome network dataset*

Description

Downloads a single network dataset from Harmonizome https://maayanlab.cloud/Harmonizome.

Usage

```
harmonizome_download(dataset)
```

Arguments

`dataset`	The dataset part of the URL. Please refer to the download section of the Harmonizome webpage.

Value

Data frame (tibble) with interactions.

Examples

```
harmonizome_data <- harmonizome_download('phosphositeplus')
harmonizome_data
# # A tibble: 6,013 x 7
#    source   source_desc source_id target target_desc target_id weight
#    <chr>    <chr>           <dbl> <chr>  <chr>           <dbl>  <dbl>
#  1 TP53     na               7157 STK17A na               9263      1
#  2 TP53     na               7157 TP53RK na             112858      1
#  3 TP53     na               7157 SMG1   na              23049      1
#  4 UPF1     na               5976 SMG1   na              23049      1
# # . with 6,003 more rows
```

has_extra_attrs *Tells if an interaction data frame has an extra_attrs column*

Description

Tells if an interaction data frame has an extra_attrs column

Usage

```
has_extra_attrs(data)
```

Arguments

data	An interaction data frame.

Value

Logical: TRUE if the data frame has the "extra_attrs" column.

See Also

- extra_attrs
- extra_attrs_to_cols
- with_extra_attrs
- filter_extra_attrs
- extra_attr_values

Examples

```
i <- import_omnipath_interactions(fields = 'extra_attrs')
has_extra_attrs(i)
```

homologene_download *Orthology table for a pair of organisms*

Description

Orthologous pairs of genes for a pair of organisms from NCBI HomoloGene, using one identifier type.

Usage

```
homologene_download(
  target = 10090L,
  source = 9606L,
  id_type = "genesymbol",
  hgroup_size = FALSE
)
```

Arguments

target	Character or integer: name or ID of the target organism.
source	Character or integer: name or ID of the source organism.
id_type	Symbol or character: identifier type, possible values are "genesymbol", "entrez", "refseqp" or "gi".
hgroup_size	Logical: include a column with the size of the homology groups. This column distinguishes one-to-one and one-to-many or many-to-many mappings.

Details

The operation of this function is symmetric, *source* and *target* are interchangeable but determine the column layout of the output. The column "hgroup" is a numberic identifier of the homology groups. Most of the groups consist of one pair of orthologous genes (one-to-one mapping), and a few of them multiple ones (one-to-many or many-to-many mappings).

Value

A data frame with orthologous identifiers between the two organisms.

See Also

- homologene_raw
- homologene_uniprot_orthology

Examples

```
chimp_human <- homologene_download(chimpanzee, human, refseqp)
chimp_human
# # A tibble: 17,737 × 3
#    hgroup refseqp_source refseqp_target
#     <int> <chr>          <chr>
#  1      3 NP_000007.1    NP_001104286.1
#  2      5 NP_000009.1    XP_003315394.1
#  3      6 NP_000010.1    XP_508738.2
#  4      7 NP_001096.1    XP_001145316.1
#  5      9 NP_000014.1    XP_523792.2
# # . with 17,732 more rows
```

homologene_raw *Orthology data from NCBI HomoloGene*

Description

Retrieves NCBI HomoloGene data without any processing. Processed tables are more useful for most purposes, see below other functions that provide those. Genes of various organisms are grouped into homology groups ("hgroup" column). Organisms are identified by NCBI Taxonomy IDs, genes are identified by four different identifier types.

Usage

```
homologene_raw()
```

Value

A data frame as provided by NCBI HomoloGene.

See Also

- homologene_download

Examples

```
hg <- homologene_raw()
hg
# # A tibble: 275,237 × 6
#    hgroup ncbi_taxid entrez  genesymbol  gi        refseqp
#     <int>      <int> <chr>   <chr>       <chr>     <chr>
#  1      3       9606 34      ACADM       4557231   NP_000007.1
#  2      3       9598 469356  ACADM       160961497 NP_001104286.1
#  3      3       9544 705168  ACADM       109008502 XP_001101274.1
#  4      3       9615 490207  ACADM       545503811 XP_005622188.1
#  5      3       9913 505968  ACADM       115497690 NP_001068703.1
# # . with 275,232 more rows

# which organisms are available?
common_name(unique(hg$ncbi_taxid))
#  [1] "Human" "Chimpanzee" "Macaque" "Dog" "Cow" "Mouse" "Rat" "Zebrafish"
#  [9] "D. melanogaster" "Caenorhabditis elegans (PRJNA13758)"
# [11] "Tropical clawed frog" "Chicken"
# ...and 9 more organisms with missing English names.
```

homologene_uniprot_orthology
Orthology table with UniProt IDs

Description

Orthologous pairs of UniProt IDs for a pair of organisms, based on NCBI HomoloGene data.

Usage

```
homologene_uniprot_orthology(target = 10090L, source = 9606L, by = entrez, ...)
```

Arguments

target	Character or integer: name or ID of the target organism.
source	Character or integer: name or ID of the source organism.
by	Symbol or character: the identifier type in NCBI HomoloGene to use. Possible values are "refseqp", "entrez", "genesymbol", "gi".
...	Further arguments passed to translate_ids.

Value

A data frame with orthologous pairs of UniProt IDs.

Examples

```
homologene_uniprot_orthology(by = genesymbol)
# # A tibble: 14,235 × 2
#    source target
#    <chr>  <chr>
#  1 P11310 P45952
#  2 P49748 P50544
#  3 P24752 Q8QZT1
#  4 Q04771 P37172
#  5 Q16586 P82350
# # . with 14,230 more rows
```

homology_translate *Homology translation*

Description

Translates identifiers between organisms using orthology data from Ensembl.

Usage

```
homology_translate(
  d,
  ...,
  target = 10090,
  source = 9606,
  ensembl_orthology_types = c("one2one", "one2many"),
  ensembl_min_orthology_confidence = 1L
)
```

Arguments

d	Data frame or character vector.
...	Column specification: from zero to up to three arguments, with or without names. NSE is supported. Arguments beyond the third one will be ignored. • The name of the arguments should be column names, the values identifier types, either as character or as symbols. • Arguments without names assumed to be both column names and identifier types, e.g. a column called "uniprot" containing UniProt IDs. • The first column spefication describes the source column, with identifiers of the source organism. This column must exist in the data and this will be the input of the homology translation. This column will be removed from the returned data frame. • In case of "uniprot", the source column name can be anything, if it contains only UniProt IDs it will be handled accordingly. • In case of "genesymbol", is enough if the source column name contains the word "genesymbol", e.g. "ligand_genesymbol". • The second column spefication describes the target column, with its name and identifier type. If not provided, both the column name and type will be the same as the source • Optionally a third column can be specified with another identifier type. This is convenient if you want, for example also Gene Symbols along with UniProt IDs. • If no specification provided, the input assumed to have a column named either "uniprot" or "genesymbol", or be a character vector of UniProt IDs or Gene Symbols.

`target` Character or integer: name or identifier of the target organism (the one we translate to). The default target organism is mouse.

`source` Character or integer: name of identifier of the source organism (the one the IDs in the input data belong to). The default source organism is human.

`ensembl_orthology_types`
Character vector: use only this orthology relationship types. Possible values are "one2one", "one2many" and "many2many".

`ensembl_min_orthology_confidence`
Integer: use only orthology relations with at least this level of confidence. In Ensembl the confidence can be either 0 or 1, so only these values make sense. If 0, all the orthology records will be used, if 1, only the ones with higher confidence.

Value

Data frame with the translated columns or character vector with translated identifiers.

Examples

```
## Not run:
# these proteins are ULK1, IFNG, EGFR, TGFB1, IL1R1
human_uniprots <- c("O75385", "P01579", "P00533", "P01137", "P14778")
homology_translate(human_uniprots)

## End(Not run)
```

---

hpo_download *Downloads protein annotations from Human Phenotype Ontology*

Description

Human Phenotype Ontology (HPO) provides a standardized vocabulary of phenotypic abnormalities encountered in human disease. Each term in the HPO describes a phenotypic abnormality. HPO currently contains over 13,000 terms and over 156,000 annotations to hereditary diseases. See more at https://hpo.jax.org/app/.

Usage

```
hpo_download()
```

Value

A tibble (data frame) of annotations as it is provided by the database

Examples

```
hpo_data <- hpo_download()
hpo_data
# # A tibble: 231,738 x 9
#    entrez_gene_id entrez_gene_symb. hpo_term_id hpo_term_name
#             <dbl> <chr>             <chr>       <chr>
#  1           8192 CLPP              HP:0000013  Hypoplasia of the ute.
#  2           8192 CLPP              HP:0004322  Short stature
#  3           8192 CLPP              HP:0000786  Primary amenorrhea
#  4           8192 CLPP              HP:0000007  Autosomal recessive i.
#  5           8192 CLPP              HP:0000815  Hypergonadotropic hyp.
# # . with 231,733 more rows, and 5 more variables:
# #   frequency_raw <chr>, frequency_hpo <chr>, info_gd_source <chr>,
# #   gd_source <chr>, disease_id <chr>
```

`htridb_download` *Downloads TF-target interactions from HTRIdb*

Description

HTRIdb (https://www.lbbc.ibb.unesp.br/htri/) is a database of literature curated human TF-target interactions. As the database is recently offline, the data is distributed by the OmniPath rescued data repository (https://rescued.omnipathdb.org/).

Usage

```
htridb_download()
```

Value

Data frame (tibble) with interactions.

Examples

```
htridb_data <- htridb_download()
htridb_data
# # A tibble: 18,630 x 7
#      OID GENEID_TF SYMBOL_TF GENEID_TG SYMBOL_TG TECHNIQUE
#    <dbl>     <dbl> <chr>         <dbl> <chr>     <chr>
#  1 32399       142 PARP1           675 BRCA2     Electrophoretic Mobi.
#  2 32399       142 PARP1           675 BRCA2     Chromatin Immunoprec.
#  3 28907       196 AHR            1543 CYP1A1    Chromatin Immunoprec.
#  4 29466       196 AHR            1543 CYP1A1    Electrophoretic Mobi.
#  5 28911       196 AHR            1543 CYP1A1    Chromatin Immunoprec.
# # . with 18,620 more rows, and 1 more variable: PUBMED_ID <chr>
```

`import_all_interactions` *Imports all interaction datasets available in OmniPath*

Description

The interaction datasets currently available in OmniPath:

Usage

```
import_all_interactions(
  resources = NULL,
  organism = 9606,
  dorothea_levels = c("A", "B"),
  exclude = NULL,
  fields = NULL,
  default_fields = TRUE,
  references_by_resource = TRUE,
  ...
)

import_AllInteractions(...)
```

Arguments

`resources`	interactions not reported in these databases are removed. See `get_interaction_resources` for more information.
`organism`	Interactions are available for human, mouse and rat. Choose among: 9606 human (default), 10116 rat and 10090 Mouse
`dorothea_levels`	The confidence levels of the dorothea interactions (TF-target) which range from A to D. Set to A and B by default.
`exclude`	Character: datasets or resources to exclude.
`fields`	The user can define here the fields to be added. If used, set the next argument, 'default_fields', to FALSE.
`default_fields`	whether to include the default fields (columns) for the query type. If FALSE, only the fields defined by the user in the 'fields' argument will be added.
`references_by_resource`	if FALSE, removes the resource name prefixes from the references (PubMed IDs); this way the information which reference comes from which resource will be lost and the PubMed IDs will be unique.
`...`	Passed to `import_all_interactions`.

Details

omnipath: the OmniPath data as defined in the paper, an arbitrary optimum between coverage and quality pathwayextra: activity flow interactions without literature reference kinaseextra: enzyme-substrate interactions without literature reference ligrecextra: ligand-receptor interactions without literature reference dorothea: transcription factor (TF)-target interactions from DoRothEA tf_target: transcription factor (TF)-target interactions from other resources mirnatarget: miRNA-mRNA interactions tf_mirna: TF-miRNA interactions lncrna_mrna: lncRNA-mRNA interactions

Value

A dataframe containing all the datasets in the interactions query

See Also

- get_interaction_resources
- interaction_graph
- print_interactions

Examples

```
interactions <- import_all_interactions(
    resources = c('HPRD', 'BioGRID'),
    organism = 9606
)
```

---

import_dorothea_interactions

From the OmniPath webservice imports interactions from the DoRothEA dataset

---

Description

Imports the dataset from: https://omnipathdb.org/interactions?datasets=dorothea which contains transcription factor (TF)-target interactions from DoRothEA https://github.com/saezlab/DoRothEA

Usage

```
import_dorothea_interactions(
  resources = NULL,
  organism = 9606,
  dorothea_levels = c("A", "B"),
  fields = NULL,
  default_fields = TRUE,
  references_by_resource = TRUE,
  exclude = NULL,
  ...
)
```

Arguments

resources	interactions not reported in these databases are removed. See get_interaction_resources for more information.
organism	Interactions are available for human, mouse and rat. Choose among: 9606 human (default), 10116 rat and 10090 Mouse
dorothea_levels	Vector detailing the confidence levels of the interactions to be downloaded. In dorothea, every TF-target interaction has a confidence score ranging from A to E, being A the most reliable interactions. By default we take A and B level interactions (c(A, B)). It is to note that E interactions are not available in OmnipathR.
fields	The user can define here the fields to be added. If used, set the next argument, 'default_fields', to FALSE.
default_fields	whether to include the default fields (columns) for the query type. If FALSE, only the fields defined by the user in the 'fields' argument will be added.
references_by_resource	if FALSE, removes the resource name prefixes from the references (PubMed IDs); this way the information which reference comes from which resource will be lost and the PubMed IDs will be unique.
exclude	Character: datasets or resources to exclude.
...	optional additional arguments

Value

A dataframe containing TF-target interactions from DoRothEA

See Also

- get_interaction_resources
- import_all_interactions
- interaction_graph
- print_interactions

Examples

```
interactions <- import_dorothea_interactions(
    resources = c('DoRothEA', 'ARACNe-GTEx_DoRothEA'),
    organism = 9606,
    dorothea_levels = c('A', 'B', 'C')
)
```

`import_intercell_network` *Intercellular communication network*

Description

Imports an intercellular network by combining intercellular annotations and protein interactions. First imports a network of protein-protein interactions. Then, it retrieves annotations about the proteins intercellular communication roles, once for the transmitter (delivering information from the expressing cell) and second, the receiver (receiving signal and relaying it towards the expressing cell) side. These 3 queries can be customized by providing parameters in lists which will be passed to the respective methods (`import_omnipath_interactions` for the network and `import_omnipath_intercell` for the annotations). Finally the 3 data frames combined in a way that the source proteins in each interaction annotated by the transmitter, and the target proteins by the receiver categories. If undirected interactions present (these are disabled by default) they will be duplicated, i.e. both partners can be both receiver and transmitter.

Usage

```
import_intercell_network(
  interactions_param = list(),
  transmitter_param = list(),
  receiver_param = list(),
  resources = NULL,
  entity_types = NULL,
  ligand_receptor = FALSE,
  high_confidence = FALSE,
  simplify = FALSE,
  unique_pairs = FALSE,
  consensus_percentile = NULL,
  loc_consensus_percentile = NULL,
  omnipath = TRUE,
  ligrecextra = TRUE,
  kinaseextra = !high_confidence,
  pathwayextra = !high_confidence,
  ...
)
```

Arguments

`interactions_param`
: a list with arguments for an interactions query: `import_omnipath_interactions`, `import_pathwayextra_interactions`, `import_kinaseextra_interactions`, `import_ligrecextra_interactions`

`transmitter_param`
: a list with arguments for `import_omnipath_intercell`, to define the transmitter side of intercellular connections

`receiver_param` a list with arguments for `import_omnipath_intercell`, to define the receiver side of intercellular connections

`resources` A character vector of resources to be applied to both the interactions and the annotations. For example, `resources = 'CellChatDB'` will download the transmitters and receivers defined by CellChatDB, connected by connections from CellChatDB.

`entity_types` Character, possible values are "protein", "complex" or both.

`ligand_receptor`
Logical. If `TRUE`, only *ligand* and *receptor* annotations will be used instead of the more generic *transmitter* and *receiver* categories.

`high_confidence`
Logical: shortcut to do some filtering in order to include only higher confidence interactions. The intercell database of OmniPath covers a very broad range of possible ways of cell to cell communication, and the pieces of information, such as localization, topology, function and interaction, are combined from many, often independent sources. This unavoidably result some weird and unexpected combinations which are false positives in the context of intercellular communication. This option sets some minimum criteria to remove most (but definitely not all!) of the wrong connections. These criteria are the followings: 1) the receiver must be plasma membrane transmembrane; 2) the curation effort for interactions must be larger than one; 3) the consensus score for annotations must be larger than the 50 percentile within the generic category (you can override this by `consensus_percentile`). 4) the transmitter must be secreted or exposed on the plasma membrane. 5) The major localizations have to be supported by at least 30 percent of the relevant resources (you can override this by `loc_consensus_percentile`). 6) The datasets with lower level of curation (*kinaseextra* and *pathwayextra*) will be disabled. These criteria are of medium stringency, you can always tune them to be more relaxed or stringent by filtering manually, using `filter_intercell_network`.

`simplify` Logical: keep only the most often used columns. This function combines a network data frame with two copies of the intercell annotation data frames, all of them already having quite some columns. With this option we keep only the names of the interacting pair, their intercellular communication roles, and the minimal information of the origin of both the interaction and the annotations.

`unique_pairs` Logical: instead of having separate rows for each pair of annotations, drop the annotations and reduce the data frame to unique interacting pairs. See `unique_intercell_network` for details.

`consensus_percentile`
Numeric: a percentile cut off for the consensus score of generic categories in intercell annotations. The consensus score is the number of resources supporting the classification of an entity into a category based on combined information of many resources. Here you can apply a cut-off, keeping only the annotations supported by a higher number of resources than a certain percentile of each category. If `NULL` no filtering will be performed. The value is either in the 0-1 range, or will be divided by 100 if greater than 1. The percentiles will be calculated against the generic composite categories and then will be applied to their resource specific annotations and specific child categories.

loc_consensus_percentile
: Numeric: similar to consensus_percentile for major localizations. For example, with a value of 50, the secreted, plasma membrane transmembrane or peripheral attributes will be TRUE only where at least 50 percent of the resources support these.

omnipath
: Logical: shortcut to include the *omnipath* dataset in the interactions query.

ligrecextra
: Logical: shortcut to include the *ligrecextra* dataset in the interactions query.

kinaseextra
: Logical: shortcut to include the *kinaseextra* dataset in the interactions query.

pathwayextra
: Logical: shortcut to include the *pathwayextra* dataset in the interactions query.

...
: If simplify or unique_pairs is TRUE, additional column names can be passed here to dplyr::select on the final data frame. Otherwise ignored.

Details

By default this function creates almost the largest possible network of intercellular interactions. However, this might contain a large number of false positives. Please refer to the documentation of the arguments, especially high_confidence, and the filter_intercell_network function. Note: if you restrict the query to certain intercell annotation resources or small categories, it's not recommended to use the consensus_percentile or high_confidence options, instead filter the network with filter_intercell_network for more consistent results.

Value

A dataframe containing information about protein-protein interactions and the inter-cellular roles of the protiens involved in those interactions.

See Also

- get_intercell_categories
- get_intercell_generic_categories
- import_omnipath_intercell
- import_omnipath_interactions
- import_pathwayextra_interactions
- import_kinaseextra_interactions
- import_ligrecextra_interactions
- unique_intercell_network
- simplify_intercell_network
- filter_intercell_network

Examples

```
intercell_network <- import_intercell_network(
    interactions_param = list(datasets = 'ligrecextra'),
    receiver_param = list(categories = c('receptor', 'transporter')),
    transmitter_param = list(categories = c('ligand', 'secreted_enzyme'))
)
```

`import_kinaseextra_interactions` *Imports interactions from the 'kinase extra' dataset of OmniPath*

Description

Imports the dataset from: https://omnipathdb.org/interactions?datasets=kinaseextra, which contains enzyme-substrate interactions without literature reference. The enzyme-substrate interactions supported by literature references are part of the 'omnipath' dataset.

Usage

```
import_kinaseextra_interactions(
  resources = NULL,
  organism = 9606,
  fields = NULL,
  default_fields = TRUE,
  references_by_resource = TRUE,
  exclude = NULL,
  ...
)
```

Arguments

`resources`	interactions not reported in these databases are removed. See `get_interaction_resources` for more information.
`organism`	Interactions are available for human, mouse and rat. Choose among: 9606 human (default), 10116 rat and 10090 Mouse
`fields`	The user can define here the fields to be added. If used, set the next argument, 'default_fields', to FALSE.
`default_fields`	whether to include the default fields (columns) for the query type. If FALSE, only the fields defined by the user in the 'fields' argument will be added.
`references_by_resource`	if FALSE, removes the resource name prefixes from the references (PubMed IDs); this way the information which reference comes from which resource will be lost and the PubMed IDs will be unique.
`exclude`	Character: datasets or resources to exclude.
`...`	Optional additional arguments.

Value

A dataframe containing enzyme-substrate interactions without literature reference

See Also

- get_interaction_resources
- import_all_interactions
- interaction_graph
- print_interactions

Examples

```
interactions <-
   import_kinaseextra_interactions(
       resources = c('PhosphoPoint', 'PhosphoSite'),
       organism = 9606
   )
```

`import_ligrecextra_interactions` *Imports interactions from the 'ligrec extra' dataset of OmniPath*

Description

Imports the dataset from: https://omnipathdb.org/interactions?datasets=ligrecextra, which contains ligand-receptor interactions without literature reference. The ligand-receptor interactions supported by literature references are part of the 'omnipath' dataset.

Usage

```
import_ligrecextra_interactions(
  resources = NULL,
  organism = 9606,
  fields = NULL,
  default_fields = TRUE,
  references_by_resource = TRUE,
  exclude = NULL,
  ...
)
```

Arguments

resources	interactions not reported in these databases are removed. See get_interaction_resources for more information.
organism	Interactions are available for human, mouse and rat. Choose among: 9606 human (default), 10116 rat and 10090 Mouse
fields	The user can define here the fields to be added. If used, set the next argument, 'default_fields', to FALSE.

default_fields whether to include the default fields (columns) for the query type. If FALSE, only the fields defined by the user in the 'fields' argument will be added.

references_by_resource if FALSE, removes the resource name prefixes from the references (PubMed IDs); this way the information which reference comes from which resource will be lost and the PubMed IDs will be unique.

exclude Character: datasets or resources to exclude.

... optional additional arguments

Value

A dataframe containing ligand-receptor interactions including the ones without literature references

See Also

- get_interaction_resources
- import_all_interactions
- interaction_graph
- print_interactions

Examples

```
interactions <- import_ligrecextra_interactions(
    resources = c('HPRD', 'Guide2Pharma'),
    organism = 9606
)
```

---

import_lncrna_mrna_interactions

Imports interactions from the lncRNA-mRNA dataset of OmniPath

---

Description

Imports the dataset from: https://omnipathdb.org/interactions?datasets=lncrna_mrna, which contains lncRNA-mRNA interactions

Usage

```
import_lncrna_mrna_interactions(
  resources = NULL,
  organism = 9606,
  fields = NULL,
  default_fields = TRUE,
  references_by_resource = TRUE,
  exclude = NULL,
  ...
)
```

Arguments

resources	interactions not reported in these databases are removed. See get_interaction_resources for more information.
organism	Interactions are available for human, mouse and rat. Choose among: 9606 human (default), 10116 rat and 10090 Mouse
fields	The user can define here the fields to be added. If used, set the next argument, 'default_fields', to FALSE.
default_fields	whether to include the default fields (columns) for the query type. If FALSE, only the fields defined by the user in the 'fields' argument will be added.
references_by_resource	if FALSE, removes the resource name prefixes from the references (PubMed IDs); this way the information which reference comes from which resource will be lost and the PubMed IDs will be unique.
exclude	Character: datasets or resources to exclude.
...	optional additional arguments

Value

A dataframe containing lncRNA-mRNA interactions

See Also

- get_interaction_resources
- import_all_interactions
- interaction_graph
- print_interactions

Examples

```
interactions <-
    import_lncrna_mrna_interactions(
        resources = c('ncRDeathDB')
    )
```

---

import_mirnatarget_interactions

Imports interactions from the miRNA-target dataset of OmniPath

---

Description

Imports the dataset from: https://omnipathdb.org/interactions?datasets=mirnatarget, which contains miRNA-mRNA interactions.

Usage

```
import_mirnatarget_interactions(
  resources = NULL,
  organism = 9606,
  fields = NULL,
  default_fields = TRUE,
  references_by_resource = TRUE,
  exclude = NULL,
  ...
)
```

Arguments

resources	interactions not reported in these databases are removed. See get_interaction_resources for more information.
organism	Interactions are available for human, mouse and rat. Choose among: 9606 human (default), 10116 rat and 10090 Mouse
fields	The user can define here the fields to be added. If used, set the next argument, 'default_fields', to FALSE.
default_fields	whether to include the default fields (columns) for the query type. If FALSE, only the fields defined by the user in the 'fields' argument will be added.
references_by_resource	if FALSE, removes the resource name prefixes from the references (PubMed IDs); this way the information which reference comes from which resource will be lost and the PubMed IDs will be unique.
exclude	Character: datasets or resources to exclude.
...	optional additional arguments

Value

A dataframe containing miRNA-mRNA interactions

See Also

- get_interaction_resources
- import_all_interactions
- interaction_graph
- print_interactions

Examples

```
interactions <-
    import_mirnatarget_interactions(
        resources = c('miRTarBase', 'miRecords')
    )
```

`import_omnipath_annotations` *Imports annotations from OmniPath*

Description

Imports protein annotations about function, localization, expression, structure and other properties of proteins from OmniPath https://omnipathdb.org/annotations. Note: there might be also a few miRNAs annotated; a vast majority of protein complex annotations are inferred from the annotations of the members: if all members carry the same annotation the complex inherits.

Usage

```
import_omnipath_annotations(
  proteins = NULL,
  resources = NULL,
  wide = FALSE,
  ...
)
```

Arguments

`proteins`	Vector containing the genes or proteins for whom annotations will be retrieved (UniProt IDs or HGNC Gene Symbols or miRBase IDs). It is also possible to donwload annotations for protein complexes. To do so, write 'COMPLEX:' right before the genesymbols of the genes integrating the complex. Check the vignette for examples.
`resources`	Load the annotations only from these databases. See `get_annotation_resources` for possible values.
`wide`	Convert the annotation table to wide format, which corresponds more or less to the original resource. If the data comes from more than one resource a list of wide tables will be returned. See examples at `pivot_annotations`.
`...`	Additional arguments.

Details

Downloading the full annotations dataset is disabled by default because the size of this data is around 1GB. We recommend to retrieve the annotations for a set of proteins or only from a few resources, depending on your interest. You can always download the full database from https://archive.omnipathdb.org/omnipath_webservice_annotations__recent.tsv using any standard R or `readr` method.

Value

A data frame containing different gene and complex annotations.

See Also

- get_annotation_databases
- pivot_annotations

Examples

```
annotations <- import_omnipath_annotations(
    proteins = c('TP53', 'LMNA'),
    resources = c('HPA_subcellular')
)
```

`import_omnipath_complexes` *Imports protein complexes from OmniPath*

Description

Imports the complexes stored in Omnipath database from https://omnipathdb.org/complexes.

Usage

```
import_omnipath_complexes(resources = NULL, ...)
```

Arguments

resources	complexes not reported in these databases are removed. See get_complexes_databases for more information.
...	optional additional arguments

Value

A dataframe containing information about complexes

See Also

- get_complexes_databases

Examples

```
complexes = import_omnipath_complexes(
    resources = c('CORUM', 'hu.MAP')
)
```

`import_omnipath_enzsub` *Imports enzyme-substrate relationships from OmniPath*

Description

Imports the enzyme-substrate (more exactly, enzyme-PTM) relationship database from https://omnipathdb.org/enzsub

Usage

```
import_omnipath_enzsub(
  resources = NULL,
  organism = 9606,
  fields = NULL,
  default_fields = TRUE,
  references_by_resource = TRUE,
  exclude = NULL,
  ...
)
```

Arguments

`resources`	PTMs not reported in these databases are removed. See `get_ptms_databases` for more information.
`organism`	PTMs are available for human, mouse and rat. Choose among: 9606 human (default), 10116 rat and 10090 Mouse
`fields`	You can define here additional fields to be added to the result. If used, set the next argument, `default_fields`, to `FALSE`.
`default_fields`	Whether to include the default fields (columns) for the query type. If `FALSE`, only the fields defined by the user in the `fields` argument will be added.
`references_by_resource`	if FALSE, removes the resource name prefixes from the references (PubMed IDs); this way the information which reference comes from which resource will be lost and the PubMed IDs will be unique.
`exclude`	Character: datasets or resources to exclude.
`...`	Optional additional arguments.

Value

A data frame containing the information about ptms

See Also

- get_enzsub_resources
- import_omnipath_interactions
- enzsub_graph
- print_interactions

Examples

```
enzsub <- import_omnipath_enzsub(
    resources = c('PhosphoSite', 'SIGNOR'),
    organism = 9606
)
```

`import_omnipath_interactions` *Imports interactions from the 'omnipath' dataset of Omnipath*

Description

Imports the database from https://omnipathdb.org/interactions, which contains only interactions supported by literature references. This part of the interaction database compiled a similar way as it has been presented in the first paper describing OmniPath (Turei et al. 2016).

Usage

```
import_omnipath_interactions(
  resources = NULL,
  organism = 9606,
  datasets = "omnipath",
  fields = NULL,
  default_fields = TRUE,
  references_by_resource = TRUE,
  exclude = NULL,
  ...
)
```

Arguments

resources	interactions not reported in these databases are removed. See get_interaction_resources for more information.
organism	Interactions are available for human, mouse and rat. Choose among: 9606 human (default), 10116 rat and 10090 Mouse
datasets	Names of the interaction datasets to download: omnipath (by default). Other possiblites are: pathwayextra, kinaseextra, ligrecextra, dorothea,tf_target, mirnatarget, tf_mirna, lncrna_mrna. The user can select multiple datasets as for example: c('omnipath', 'pathwayextra', 'kinaseextra')

`fields`	The user can define here the fields to be added. If used, set the next argument, 'default_fields', to FALSE.
`default_fields`	whether to include the default fields (columns) for the query type. If FALSE, only the fields defined by the user in the 'fields' argument will be added.
`references_by_resource`	if FALSE, removes the resource name prefixes from the references (PubMed IDs); this way the information which reference comes from which resource will be lost and the PubMed IDs will be unique.
`exclude`	Character: datasets or resources to exclude.
`...`	optional additional arguments

Value

A dataframe of protein-protein interactions

See Also

- `get_interaction_resources`
- `import_all_interactions`
- `interaction_graph`
- `print_interactions`

Examples

```
interactions = import_omnipath_interactions(
    resources = c('SignaLink3'),
    organism = 9606
)
```

---

import_omnipath_intercell

Imports OmniPath intercell annotations

Description

Imports the OmniPath intercellular communication role annotation database from https://omnipathdb.org/intercell. It provides information on the roles in inter-cellular signaling. E.g. if a protein is a ligand, a receptor, an extracellular matrix (ECM) component, etc.

Usage

```
import_omnipath_intercell(
  categories = NULL,
  resources = NULL,
  parent = NULL,
  scope = NULL,
  aspect = NULL,
  source = NULL,
  transmitter = NULL,
  receiver = NULL,
  secreted = NULL,
  plasma_membrane_peripheral = NULL,
  plasma_membrane_transmembrane = NULL,
  proteins = NULL,
  topology = NULL,
  causality = NULL,
  consensus_percentile = NULL,
  loc_consensus_percentile = NULL,
  ...
)
```

Arguments

categories	vector containing the categories to be retrieved. All the genes belonging to those categories will be returned. For further information about the categories see codeget_intercell_categories.
resources	limit the query to certain resources; see the available resources by get_intercell_resources.
parent	vector containing the parent classes to be retrieved. All the genes belonging to those classes will be returned. For furter information about the main classes see get_intercell_categories.
scope	either 'specific' or 'generic'
aspect	either 'locational' or 'functional'
source	either 'resource_specific' or 'composite'
transmitter	logical, include only transmitters i.e. proteins delivering signal from a cell to its environment.
receiver	logical, include only receivers i.e. proteins delivering signal to the cell from its environment.
secreted	logical, include only secreted proteins
plasma_membrane_peripheral	logical, include only plasma membrane peripheral membrane proteins.
plasma_membrane_transmembrane	logical, include only plasma membrane transmembrane proteins.
proteins	limit the query to certain proteins
topology	topology categories: one or more of 'secreted' (sec), 'plasma_membrane_peripheral' (pmp), 'plasma_membrane_transmembrane' (pmtm) (both short or long notation can be used).

causality ‘transmitter‘ (trans), ‘receiver‘ (rec) or ‘both‘ (both short or long notation can be used).

consensus_percentile
Numeric: a percentile cut off for the consensus score of generic categories. The consensus score is the number of resources supporting the classification of an entity into a category based on combined information of many resources. Here you can apply a cut-off, keeping only the annotations supported by a higher number of resources than a certain percentile of each category. If NULL no filtering will be performed. The value is either in the 0-1 range, or will be divided by 100 if greater than 1. The percentiles will be calculated against the generic composite categories and then will be applied to their resource specific annotations and specific child categories.

loc_consensus_percentile
Numeric: similar to codeconsensus_percentile for major localizations. For example, with a value of 50, the secreted, plasma membrane transmembrane or peripheral attributes will be true only where at least 50 percent of the resources support these.

... Additional optional arguments, ignored.

Value

A dataframe cotaining information about roles in intercellular signaling.

See Also

- get_intercell_categories
- get_intercell_generic_categories
- import_intercell_network
- intercell_consensus_filter

Examples

```
intercell <- import_omnipath_intercell(categories = 'ecm')
```

---

import_pathwayextra_interactions

Imports interactions from the ‘pathway extra‘ dataset of Omnipath

---

Description

Imports the dataset from: https://omnipathdb.org/interactions?datasets=pathwayextra, which contains activity flow interactions without literature reference. The activity flow interactions supported by literature references are part of the ‘omnipath‘ dataset.

Usage

```
import_pathwayextra_interactions(
  resources = NULL,
  organism = 9606,
  fields = NULL,
  default_fields = TRUE,
  references_by_resource = TRUE,
  exclude = NULL,
  ...
)
```

Arguments

resources	interactions not reported in these databases are removed. See get_interaction_resources for more information.
organism	Interactions are available for human, mouse and rat. Choose one of those: 9606 human (default), 10116 rat or 10090 Mouse.
fields	The user can define here the fields to be added. If used, set the next argument, 'default_fields', to FALSE.
default_fields	whether to include the default fields (columns) for the query type. If FALSE, only the fields defined by the user in the 'fields' argument will be added.
references_by_resource	if FALSE, removes the resource name prefixes from the references (PubMed IDs); this way the information which reference comes from which resource will be lost and the PubMed IDs will be unique.
exclude	Character: datasets or resources to exclude.
...	optional additional arguments

Value

A dataframe containing activity flow interactions between proteins without literature reference

See Also

- get_interaction_resources
- import_all_interactions
- interaction_graph
- print_interactions

Examples

```
interactions <-
    import_pathwayextra_interactions(
        resources = c('BioGRID', 'IntAct'),
        organism = 9606
    )
```

`import_post_translational_interactions` *All post-translational interactions from OmniPath*

Description

Imports interactions from all post-translational datasets of OmniPath. The datasets are "omnipath", "kinaseextra", "pathwayextra" and "ligrecextra".

Usage

```
import_post_translational_interactions(
  resources = NULL,
  organism = 9606,
  exclude = NULL,
  references_by_resource = TRUE,
  ...
)
```

Arguments

`resources`	interactions not reported in these databases are removed. See `get_interaction_resources` for more information.
`organism`	Interactions are available for human, mouse and rat. Choose among: 9606 human (default), 10116 rat and 10090 Mouse
`exclude`	Character: datasets or resources to exclude.
`references_by_resource`	if FALSE, removes the resource name prefixes from the references (PubMed IDs); this way the information which reference comes from which resource will be lost and the PubMed IDs will be unique.
`...`	optional additional arguments

Value

A dataframe containing post-translational interactions

See Also

- `get_interaction_resources`
- `import_all_interactions`
- `interaction_graph`
- `print_interactions`

Examples

```
interactions <-
    import_post_translational_interactions(
        resources = c('BioGRID')
    )
```

`import_small_molecule_protein_interactions`
Interactions from the small molecule-protein dataset of OmniPath

---

Description

Imports the dataset from: https://omnipathdb.org/interactions?datasets=small_molecule, which contains small molecule-protein interactions. Small molecules can be metabolites, intrinsic ligands or drug compounds.

Usage

```
import_small_molecule_protein_interactions(
  resources = NULL,
  organism = 9606,
  fields = NULL,
  default_fields = TRUE,
  references_by_resource = TRUE,
  exclude = NULL,
  ...
)
```

Arguments

`resources`	interactions not reported in these databases are removed. See `get_interaction_resources` for more information.
`organism`	Interactions are available for human, mouse and rat. Choose among: 9606 human (default), 10116 rat and 10090 Mouse.
`fields`	Optional fields to be added.
`default_fields`	whether to include the default fields (columns) for the query type. If FALSE, only the fields defined by the user in the ‘fields‘ argument will be added.
`references_by_resource`	If `FALSE`, removes the resource name prefixes from the references (PubMed IDs); this way the information which reference comes from which resource will be lost and the PubMed IDs will be unique.
`exclude`	Character: datasets or resources to exclude.
`...`	optional additional arguments

Value

A dataframe of small molecule-protein interactions

See Also

- get_interaction_resources
- import_all_interactions
- interaction_graph
- print_interactions

Examples

```
# What are the targets of aspirin?
interactions <-
    import_small_molecule_protein_interactions(
        sources = 'ASPIRIN'
    )
# The prostaglandin synthases:
interactions
```

---

import_tf_mirna_interactions

Imports interactions from the TF-miRNA dataset of OmniPath

Description

Imports the dataset from: https://omnipathdb.org/interactions?datasets=tf_mirna, which contains transcription factor-miRNA gene interactions

Usage

```
import_tf_mirna_interactions(
  resources = NULL,
  organism = 9606,
  fields = NULL,
  default_fields = TRUE,
  references_by_resource = TRUE,
  exclude = NULL,
  ...
)
```

Arguments

resources	interactions not reported in these databases are removed. See get_interaction_resources for more information.
organism	Interactions are available for human, mouse and rat. Choose among: 9606 human (default), 10116 rat and 10090 Mouse
fields	The user can define here the fields to be added. If used, set the next argument, 'default_fields', to FALSE.
default_fields	whether to include the default fields (columns) for the query type. If FALSE, only the fields defined by the user in the 'fields' argument will be added.
references_by_resource	if FALSE, removes the resource name prefixes from the references (PubMed IDs); this way the information which reference comes from which resource will be lost and the PubMed IDs will be unique.
exclude	Character: datasets or resources to exclude.
...	optional additional arguments

Value

A dataframe containing TF-miRNA interactions

See Also

- get_interaction_resources
- import_all_interactions
- interaction_graph
- print_interactions

Examples

```
interactions <-
    import_tf_mirna_interactions(
        resources = c('TransmiR')
    )
```

---

import_tf_target_interactions

Imports interactions from the TF-target dataset of OmniPath

Description

Imports the dataset from: https://omnipathdb.org/interactions?datasets=tf_target, which contains transcription factor-target protein coding gene interactions. Note: this is not the only TF-target dataset in OmniPath, 'dorothea' is the other one and the 'tf_mirna' dataset provides TF-miRNA gene interactions.

Usage

```
import_tf_target_interactions(
  resources = NULL,
  organism = 9606,
  fields = NULL,
  default_fields = TRUE,
  references_by_resource = TRUE,
  exclude = NULL,
  ...
)
```

Arguments

resources	interactions not reported in these databases are removed. See get_interaction_resources for more information.
organism	Interactions are available for human, mouse and rat. Choose among: 9606 human (default), 10116 rat and 10090 Mouse
fields	The user can define here the fields to be added. If used, set the next argument, 'default_fields', to FALSE.
default_fields	whether to include the default fields (columns) for the query type. If FALSE, only the fields defined by the user in the 'fields' argument will be added.
references_by_resource	if FALSE, removes the resource name prefixes from the references (PubMed IDs); this way the information which reference comes from which resource will be lost and the PubMed IDs will be unique.
exclude	Character: datasets or resources to exclude.
...	Optional additional arguments

Value

A dataframe containing TF-target interactions

See Also

- get_interaction_resources
- import_all_interactions
- interaction_graph
- print_interactions

Examples

```
interactions <-
    import_tf_target_interactions(
        resources = c('DoRothEA', 'SIGNOR')
    )
```

`import_transcriptional_interactions` *Imports all TF-target interactions from OmniPath*

Description

Imports the dataset from: https://omnipathdb.org/interactions?datasets=tf_target,dorothea, which contains transcription factor-target protein coding gene interactions.

Usage

```
import_transcriptional_interactions(
  resources = NULL,
  organism = 9606,
  dorothea_levels = c("A", "B"),
  references_by_resource = TRUE,
  exclude = NULL,
  ...
)
```

Arguments

`resources`	interactions not reported in these databases are removed. See `get_interaction_resources` for more information.
`organism`	Interactions are available for human, mouse and rat. Choose among: 9606 human (default), 10116 rat and 10090 Mouse
`dorothea_levels`	Vector detailing the confidence levels of the interactions to be downloaded. In dorothea, every TF-target interaction has a confidence score ranging from A to E, being A the most reliable interactions. By default we take A and B level interactions (`c(A, B)`). It is to note that E interactions are not available in OmnipathR.
`references_by_resource`	if FALSE, removes the resource name prefixes from the references (PubMed IDs); this way the information which reference comes from which resource will be lost and the PubMed IDs will be unique.
`exclude`	Character: datasets or resources to exclude.
`...`	Optional additional arguments.

Value

A dataframe containing TF-target interactions.

See Also

- get_interaction_resources
- import_all_interactions
- interaction_graph
- print_interactions

Examples

```
interactions <-
    import_transcriptional_interactions(
        resources = c('PAZAR', 'ORegAnno', 'DoRothEA')
    )
```

---

inbiomap_download *Downloads and preprocesses network data from InWeb InBioMap*

Description

Downloads the data by inbiomap_raw, extracts the UniProt IDs, Gene Symbols and scores and removes the irrelevant columns.

Usage

```
inbiomap_download(...)
```

Arguments

...	Passed to inbiomap_raw.

Value

A data frame (tibble) of interactions.

See Also

inbiomap_raw

Examples

```
## Not run:
inbiomap_interactions <- inbiomap_download()
inbiomap_interactions

## End(Not run)
# # A tibble: 625,641 x 7
#    uniprot_a uniprot_b genesymbol_a genesymbol_b inferred score1 score2
```

```
#    <chr>     <chr>     <chr>        <chr>        <lgl>     <dbl>  <dbl>
#  1 A0A5B9    P01892    TRBC2        HLA-A        FALSE     0.417 0.458
#  2 A0AUZ9    Q96CV9    KANSL1L      OPTN         FALSE     0.155 0.0761
#  3 A0AV02    P24941    SLC12A8      CDK2         TRUE      0.156 0.0783
#  4 A0AV02    Q00526    SLC12A8      CDK3         TRUE      0.157 0.0821
#  5 A0AV96    P0CG48    RBM47        UBC          FALSE     0.144 0.0494
# # . with 625,631 more rows
```

`inbiomap_raw` *Downloads network data from InWeb InBioMap*

Description

Downloads the data from https://inbio-discover.com/map.html#downloads in tar.gz format, extracts the PSI MITAB table and returns it as a data frame.

Usage

```
inbiomap_raw(curl_verbose = FALSE)
```

Arguments

`curl_verbose`	Logical. Perform CURL requests in verbose mode for debugging purposes.

Value

A data frame (tibble) with the extracted interaction table.

See Also

`inbiomap_download`

Examples

```
## Not run:
inbiomap_psimitab <- inbiomap_raw()

## End(Not run)
```

`interaction_graph` *Build Omnipath interaction graph*

Description

Transforms the interactions data frame to an igraph graph object.

Usage

```
interaction_graph(interactions = interactions)
```

Arguments

`interactions`	data.frame created by • `import_omnipath_enzsub` • `import_omnipath_interactions` • `import_pathwayextra_interactions` • `import_kinaseextra_interactions` • `import_ligrecextra_interactions` • `import_post_translational_interactions` • `import_dorothea_interactions` • `import_tf_target_interactions` • `import_transcriptional_interactions` • `import_mirnatarget_interactions` • `import_all_interactions`

Value

An igraph graph object.

See Also

- `import_omnipath_interactions`
- `import_pathwayextra_interactions`
- `import_kinaseextra_interactions`
- `import_ligrecextra_interactions`
- `import_dorothea_interactions`
- `import_mirnatarget_interactions`
- `import_all_interactions`
- `giant_component`
- `find_all_paths`

Examples

```
interactions <- import_omnipath_interactions(resources = c('SignaLink3'))
g <- interaction_graph(interactions)
```

`intercell_categories` *Full list of intercell categories and resources*

Description

Full list of intercell categories and resources

Usage

```
intercell_categories()
```

Value

A data frame of categories and resources.

Examples

```
ic_cat <- intercell_categories()
ic_cat
# # A tibble: 1,125 x 3
#    category               parent                 database
#    <chr>                  <chr>                  <chr>
#  1 transmembrane          transmembrane          UniProt_location
#  2 transmembrane          transmembrane          UniProt_topology
#  3 transmembrane          transmembrane          UniProt_keyword
#  4 transmembrane          transmembrane_predicted Phobius
#  5 transmembrane_phobius  transmembrane_predicted Almen2009
# # . with 1,120 more rows
```

`intercell_consensus_filter`
Quality filter for intercell annotations

Description

Quality filter for intercell annotations

Usage

```
intercell_consensus_filter(
  data,
  percentile = NULL,
  loc_percentile = NULL,
  topology = NULL
)
```

Arguments

data	A data frame with intercell annotations, as provided by import_omnipath_intercell.
percentile	Numeric: a percentile cut off for the consensus score of composite categories. The consensus score is the number of resources supporting the classification of an entity into a category based on combined information of many resources. Here you can apply a cut-off, keeping only the annotations supported by a higher number of resources than a certain percentile of each category. If NULL no filtering will be performed. The value is either in the 0-1 range, or will be divided by 100 if greater than 1. The percentiles will be calculated against the generic composite categories and then will be applied to their resource specific annotations and specific child categories.
loc_percentile	Numeric: similar to percentile for major localizations. For example, with a value of 50, the secreted, plasma membrane transmembrane or peripheral attributes will be TRUE only where at least 50 percent of the resources support these.
topology	Character vector: list of allowed topologies, possible values are *"secreted"*, *"plasma_membrane_peripheral"* and *"plasma_membrane_transmembrane"*.

Value

The data frame in data filtered by the consensus scores.

Examples

```
intercell <- import_omnipath_intercell(parent = c('ligand', 'receptor'))
nrow(intercell)
# [1] 50174
intercell_q50 <- intercell_consensus_filter(intercell, 50)
nrow(intercell_q50)
# [1] 42863
```

---

is_ontology_id *Looks like an ontology ID*

Description

Tells if the input has the typical format of ontology IDs, i.e. a code of capital letters, a colon, followed by a numeric code.

Usage

```
is_ontology_id(terms)
```

Arguments

terms	Character vector with strings to check.

Value

A logical vector with the same length as the input.

Examples

```
is_ontology_id(c('GO:0000001', 'reproduction'))
# [1]  TRUE FALSE
```

---

is_swissprot *Check for SwissProt IDs*

Description

Check for SwissProt IDs

Usage

```
is_swissprot(uniprots, organism = 9606)
```

Arguments

uniprots	Character vector of UniProt IDs.
organism	Character or integer: name or identifier of the organism.

Value

Logical vector TRUE for SwissProt IDs and FALSE for any other element.

Examples

```
is_swissprot(c("Q05BL1", "A0A654IBU3", "P00533"))
# [1] FALSE FALSE TRUE
```

is_trembl *Check for TrEMBL IDs*

Description

Check for TrEMBL IDs

Usage

```
is_trembl(uniprots, organism = 9606)
```

Arguments

uniprots	Character vector of UniProt IDs.
organism	Character or integer: name or identifier of the organism.

Value

Logical vector TRUE for TrEMBL IDs and FALSE for any other element.

Examples

```
is_trembl(c("Q05BL1", "A0A654IBU3", "P00533"))
# [1] TRUE TRUE FALSE
```

is_uniprot *Looks like a UniProt ID?*

Description

This function checks only the format of the IDs, no guarantee that these IDs exist in UniProt.

Usage

```
is_uniprot(identifiers)
```

Arguments

identifiers	Character: one or more identifiers (typically a single string, a vector or a data frame column).

Value

Logical: true if all elements in the input (except NAs) looks like valid UniProt IDs. If the input is not a character vector, 'FALSE' is returned.

Examples

```
is_uniprot(all_uniprot_acs())
# [1] TRUE
is_uniprot("P00533")
# [1] TRUE
is_uniprot("pizza")
# [1] FALSE
```

---

kegg_info *Information about a KEGG Pathway*

Description

Information about a KEGG Pathway

Usage

```
kegg_info(pathway_id)
```

Arguments

pathway_id	Character: a KEGG Pathway identifier, e.g. "hsa04710". For a complete list of IDs see kegg_pathway_list.

Value

List with the pathway information.

See Also

- kegg_pathway_list
- kegg_picture
- kegg_open

Examples

```
kegg_info('map00563')
```

kegg_open *Open a KEGG Pathway diagram in the browser*

Description

Open a KEGG Pathway diagram in the browser

Usage

```
kegg_open(pathway_id)
```

Arguments

pathway_id	Character: a KEGG Pathway identifier, e.g. "hsa04710". For a complete list of IDs see kegg_pathway_list.

Details

To open URLs in the web browser the "browser" option must to be set to a a valid executable. You can check the value of this option by `getOption("browser")`. If your browser is firefox and the executable is located in the system path, you can set the option to point to it: `options(browser = "firefox")`. To make it a permanent setting, you can also include this in your `.Rprofile` file.

Value

Returns `NULL`.

See Also

- kegg_pathway_list
- kegg_picture
- kegg_info

Examples

```
if(any(getOption('browser') != '')) kegg_open('hsa04710')
```

kegg_pathways_download *Download the KEGG Pathways database*

Description

Downloads all pathway diagrams in the KEGG Pathways database in KGML format and processes the XML to extract the interactions.

Usage

```
kegg_pathways_download(max_expansion = NULL, simplify = FALSE)
```

Arguments

max_expansion	Numeric: the maximum number of relations derived from a single relation record. As one entry might represent more than one molecular entities, one relation might yield a large number of relations in the processing. This happens in a combinatorial way, e.g. if the two entries represent 3 and 4 entities, that results 12 relations. If NULL, all relations will be expanded.
simplify	Logical: remove KEGG's internal identifiers and the pathway annotations, keep only unique interactions with direction and effect sign.

Value

A data frame (tibble) of interactions.

See Also

- kegg_pathway_list
- kegg_process
- kegg_pathway_download

Examples

```
## Not run:
kegg_pw <- kegg_pathways_download(simplify = TRUE)
kegg_pw
# # A tibble: 6,765 x 6
#    uniprot_source uniprot_target type  effect genesymbol_source
#    <chr>          <chr>          <chr> <chr>  <chr>
#  1 Q03113         Q15283         PPrel activ. GNA12
#  2 Q9Y4G8         P62070         PPrel activ. RAPGEF2
#  3 Q13972         P62070         PPrel activ. RASGRF1
#  4 O95267         P62070         PPrel activ. RASGRP1
#  5 P62834         P15056         PPrel activ. RAP1A
# # . with 6,760 more rows, and 1 more variable: genesymbol_target <chr>
```

```
## End(Not run)
```

---

kegg_pathway_annotations *Protein pathway annotations*

Description

Downloads all KEGG pathways and creates a table of protein-pathway annotations.

Usage

```
kegg_pathway_annotations(pathways = NULL)
```

Arguments

pathways	A table of KEGG pathways as produced by kegg_pathways_download.

Value

A data frame (tibble) with UniProt IDs and pathway names.

See Also

kegg_pathways_download

Examples

```
## Not run:
kegg_pw_annot <- kegg_pathway_annotations()
kegg_pw_annot
# # A tibble: 7,341 x 4
#    uniprot genesymbol pathway                pathway_id
#    <chr>   <chr>      <chr>                  <chr>
#  1 Q03113  GNA12      MAPK signaling pathway hsa04010
#  2 Q9Y4G8  RAPGEF2    MAPK signaling pathway hsa04010
#  3 Q13972  RASGRF1    MAPK signaling pathway hsa04010
#  4 O95267  RASGRP1    MAPK signaling pathway hsa04010
#  5 P62834  RAP1A      MAPK signaling pathway hsa04010
# # . with 7,336 more rows

## End(Not run)
```

`kegg_pathway_download` *Download one KEGG pathway*

Description

Downloads one pathway diagram from the KEGG Pathways database in KGML format and processes the XML to extract the interactions.

Usage

```
kegg_pathway_download(
  pathway_id,
  process = TRUE,
  max_expansion = NULL,
  simplify = FALSE
)
```

Arguments

`pathway_id`	Character: a KEGG pathway identifier, for example "hsa04350".
`process`	Logical: process the data or return it in raw format. processing means joining the entries and relations into a single data frame and adding UniProt IDs.
`max_expansion`	Numeric: the maximum number of relations derived from a single relation record. As one entry might represent more than one molecular entities, one relation might yield a large number of relations in the processing. This happens in a combinatorial way, e.g. if the two entries represent 3 and 4 entities, that results 12 relations. If `NULL`, all relations will be expanded.
`simplify`	Logical: remove KEGG's internal identifiers and the pathway annotations, keep only unique interactions with direction and effect sign.

Value

A data frame (tibble) of interactions if `process` is `TRUE`, otherwise a list with two data frames: "entries" is a raw table of the entries while "relations" is a table of relations extracted from the KGML file.

See Also

- `kegg_process`
- `kegg_pathways_download`
- `kegg_pathway_list`

Examples

```
tgf_pathway <- kegg_pathway_download('hsa04350')
tgf_pathway
# # A tibble: 50 x 12
#    source target type  effect arrow relation_id kegg_id_source
#    <chr>  <chr>  <chr> <chr>  <chr> <chr>       <chr>
#  1 51     49     PPrel activ. -->   hsa04350:1  hsa:7040 hsa:.
#  2 57     55     PPrel activ. -->   hsa04350:2  hsa:151449 hs.
#  3 34     32     PPrel activ. -->   hsa04350:3  hsa:3624 hsa:.
#  4 20     17     PPrel activ. -->   hsa04350:4  hsa:4838
#  5 60     46     PPrel activ. -->   hsa04350:5  hsa:4086 hsa:.
# # . with 45 more rows, and 5 more variables: genesymbol_source <chr>,
# #   uniprot_source <chr>, kegg_id_target <chr>,
# #   genesymbol_target <chr>, uniprot_target <chr>
```

---

kegg_pathway_list *List of KEGG pathways*

Description

Retrieves a list of available KEGG pathways.

Usage

```
kegg_pathway_list()
```

Value

Data frame of pathway names and identifiers.

See Also

- kegg_process
- kegg_pathway_download
- kegg_pathways_download
- kegg_open
- kegg_picture
- kegg_info

Examples

```
kegg_pws <- kegg_pathway_list()
kegg_pws
# # A tibble: 521 x 2
#    id       name
#    <chr>    <chr>
```

```
#  1 map01100 Metabolic pathways
#  2 map01110 Biosynthesis of secondary metabolites
#  3 map01120 Microbial metabolism in diverse environments
#  4 map01200 Carbon metabolism
#  5 map01210 2-Oxocarboxylic acid metabolism
#  6 map01212 Fatty acid metabolism
#  7 map01230 Biosynthesis of amino acids
# # . with 514 more rows
```

---

kegg_picture *Download a pathway diagram as a picture*

Description

Downloads a KEGG Pathway diagram as a PNG image.

Usage

```
kegg_picture(pathway_id, path = NULL)
```

Arguments

pathway_id	Character: a KEGG Pathway identifier, e.g. "hsa04710". For a complete list of IDs see kegg_pathway_list.
path	Character: save the image to this path. If NULL, the image will be saved in the current directory under the name <pathway_id>.png.

Value

Invisibly returns the path to the downloaded file.

See Also

kegg_pathway_list

- kegg_pathway_list
- kegg_open
- kegg_info

Examples

```
kegg_picture('hsa04710')
kegg_picture('hsa04710', path = 'foo/bar')
kegg_picture('hsa04710', path = 'foo/bar/circadian.png')
```

kegg_process *Interactions from KGML*

Description

Processes KEGG Pathways data extracted from a KGML file. Joins the entries and relations into a single data frame and translates the Gene Symbols to UniProt IDs.

Usage

```
kegg_process(entries, relations, max_expansion = NULL, simplify = FALSE)
```

Arguments

entries	A data frames with entries extracted from a KGML file by kegg_pathway_download.
relations	A data frames with relations extracted from a KGML file by kegg_pathway_download.
max_expansion	Numeric: the maximum number of relations derived from a single relation record. As one entry might represent more than one molecular entities, one relation might yield a large number of relations in the processing. This happens in a combinatorial way, e.g. if the two entries represent 3 and 4 entities, that results 12 relations. If NULL, all relations will be expanded.
simplify	Logical: remove KEGG's internal identifiers and the pathway annotations, keep only unique interactions with direction and effect sign.

Value

A data frame (tibble) of interactions. In rare cases when a pathway doesn't contain any relation, returns NULL.

See Also

- kegg_pathway_download
- kegg_pathways_download
- kegg_pathway_list

Examples

```
hsa04350 <- kegg_pathway_download('hsa04350', process = FALSE)
tgf_pathway <- kegg_process(hsa04350$entries, hsa04350$relations)
tgf_pathway
# # A tibble: 50 x 12
#    source target type  effect arrow relation_id kegg_id_source
#    <chr>  <chr>  <chr> <chr>  <chr> <chr>       <chr>
#  1 51     49     PPrel activ. -->   hsa04350:1  hsa:7040 hsa:.
#  2 57     55     PPrel activ. -->   hsa04350:2  hsa:151449 hs.
#  3 34     32     PPrel activ. -->   hsa04350:3  hsa:3624 hsa:.
#  4 20     17     PPrel activ. -->   hsa04350:4  hsa:4838
```

```
#  5 60     46     PPrel activ. -->   hsa04350:5  hsa:4086 hsa:.
# # . with 45 more rows, and 5 more variables: genesymbol_source <chr>,
# #   uniprot_source <chr>, kegg_id_target <chr>,
# #   genesymbol_target <chr>, uniprot_target <chr>
```

`latin_name` *Latin (scientific) names of organisms*

Description

Latin (scientific) names of organisms

Usage

```
latin_name(name)
```

Arguments

`name`	Vector with any kind of organism name or identifier, can be also mixed type.

Value

Character vector with latin (scientific) names, NA if a name in the input could not be found.

See Also

- `ncbi_taxid`
- `common_name`
- `ensembl_name`

Examples

```
latin_name(c(9606, "cat", "dog"))
# [1] "Homo sapiens" "Felis catus" "Canis lupus familiaris"
latin_name(c(9606, "cat", "doggy"))
# [1] "Homo sapiens" "Felis catus"  NA
```

load_db *Load a built in database*

Description

Load a built in database

Usage

```
load_db(key, param = list())
```

Arguments

key	Character: the key of the database to load. For a list of available keys see omnipath_show_db.
param	List: override the defaults or pass further parameters to the database loader function. See the loader functions and their default parameters in omnipath_show_db.

Details

This function loads a database which is stored within the package namespace until its expiry. The loaded database is accessible by get_db and the loading process is typically initiated by get_db, not by the users directly.

Value

Returns NULL.

See Also

omnipath_show_db, get_db

Examples

```
load_db('go_slim')
omnipath_show_db()
```

ncbi_taxid *NCBI Taxonomy IDs of organisms*

Description

NCBI Taxonomy IDs of organisms

Usage

```
ncbi_taxid(name)
```

Arguments

name	Vector with any kind of organism name or identifier, can be also mixed type.

Value

Integer vector with NCBI Taxonomy IDs, NA if a name in the input could not be found.

See Also

- latin_name
- common_name
- ensembl_name

Examples

```
ncbi_taxid(c("Homo sapiens", "cat", "dog"))
# [1] 9606 9685 9615
ncbi_taxid(c(9606, "cat", "doggy"))
# [1] 9606 9685   NA
```

nichenet_build_model *Construct a NicheNet ligand-target model*

Description

Construct a NicheNet ligand-target model

Usage

```
nichenet_build_model(optimization_results, networks, use_weights = TRUE)
```

Arguments

optimization_results	The outcome of NicheNet parameter optimization as produced by nichenet_optimization.
networks	A list with NicheNet format signaling, ligand-receptor and gene regulatory networks as produced by nichenet_networks.
use_weights	Logical: whether to use the optimized weights.

Value

A named list with two elements: ‘weighted_networks‘ and ‘optimized_parameters‘.

Examples

```
## Not run:
expression <- nichenet_expression_data()
networks <- nichenet_networks()
optimization_results <- nichenet_optimization(networks, expression)
nichenet_model <- nichenet_build_model(optimization_results, networks)

## End(Not run)
```

---

nichenet_expression_data

Expression data from ligand-receptor perturbation experiments used by NicheNet

---

Description

NicheNet uses expression data from a collection of published ligand or receptor KO or perturbation experiments to build its model. This function retrieves the original expression data, deposited in Zenodo (https://zenodo.org/record/3260758).

Usage

```
nichenet_expression_data()
```

Value

Nested list, each element contains a data frame of processed expression data and key variables about the experiment.

Examples

```
## Not run:
exp_data <- nichenet_expression_data()
head(names(exp_data))
# [1] "bmp4_tgfb"     "tgfb_bmp4"     "nodal_Nodal"   "spectrum_Il4"
# [5] "spectrum_Tnf"  "spectrum_Ifng"
purrr::map_chr(head(exp_data), 'from')
#     bmp4_tgfb     tgfb_bmp4   nodal_Nodal  spectrum_Il4  spectrum_Tnf
#       "BMP4"       "TGFB1"       "NODAL"          "IL4"         "TNF"
# spectrum_Ifng
#       "IFNG"

## End(Not run)
```

---

nichenet_gr_network *Builds a NicheNet gene regulatory network*

Description

Builds gene regulatory network prior knowledge for NicheNet using multiple resources.

Usage

```
nichenet_gr_network(
  omnipath = list(),
  harmonizome = list(),
  regnetwork = list(),
  htridb = list(),
  remap = list(),
  evex = list(),
  pathwaycommons = list(),
  trrust = list(),
  only_omnipath = FALSE
)
```

Arguments

omnipath	List with paramaters to be passed to nichenet_gr_network_omnipath.
harmonizome	List with paramaters to be passed to nichenet_gr_network_harmonizome.
regnetwork	List with paramaters to be passed to nichenet_gr_network_regnetwork.
htridb	List with paramaters to be passed to nichenet_gr_network_htridb.
remap	List with paramaters to be passed to nichenet_gr_network_remap.
evex	List with paramaters to be passed to nichenet_gr_network_evex.
pathwaycommons	List with paramaters to be passed to nichenet_gr_network_pathwaycommons.
trrust	List with paramaters to be passed to nichenet_gr_network_trrust.
only_omnipath	Logical: a shortcut to use only OmniPath as network resource.

Value

A network data frame (tibble) with gene regulatory interactions suitable for use with NicheNet.

See Also

- nichenet_gr_network_evex
- nichenet_gr_network_harmonizome
- nichenet_gr_network_htridb
- nichenet_gr_network_omnipath
- nichenet_gr_network_pathwaycommons
- nichenet_gr_network_regnetwork
- nichenet_gr_network_remap
- nichenet_gr_network_trrust

Examples

```
## Not run:
# load everything with the default parameters:
gr_network <- nichenet_gr_network()

# less targets from ReMap, not using RegNetwork:
gr_network <- nichenet_gr_network(
    # I needed to disable ReMap here due to some issues
    # of one of the Bioconductor build servers
    # remap = list(top_targets = 200),
    remap = NULL,
    regnetwork = NULL,
)

## End(Not run)

# use only OmniPath:
gr_network_omnipath <- nichenet_gr_network(only_omnipath = TRUE)
```

---

nichenet_gr_network_evex

NicheNet gene regulatory network from EVEX

Description

Builds a gene regulatory network using data from the EVEX database and converts it to a format suitable for NicheNet.

Usage

```
nichenet_gr_network_evex(
  top_confidence = 0.75,
  indirect = FALSE,
  regulation_of_expression = FALSE
)
```

Arguments

top_confidence	Double, between 0 and 1. Threshold based on the quantile of the confidence score.
indirect	Logical: whether to include indirect interactions.
regulation_of_expression	Logical: whether to include also the "regulation of expression" type interactions.

Value

Data frame of interactions in NicheNet format.

Data frame with gene regulatory interactions in NicheNet format.

See Also

- nichenet_gr_network
- evex_download

Examples

```
# use only the 10% with the highest confidence:
evex_gr_network <- nichenet_gr_network_evex(top_confidence = .9)
```

---

nichenet_gr_network_harmonizome

NicheNet gene regulatory network from Harmonizome

Description

Builds gene regulatory network prior knowledge for NicheNet using Harmonizome

Usage

```
nichenet_gr_network_harmonizome(
  datasets = c("cheappi", "encodetfppi", "jasparpwm", "transfac", "transfacpwm",
    "motifmap", "geotf", "geokinase", "geogene"),
  ...
)
```

Arguments

datasets	The datasets to use. For possible values please refer to default value and the Harmonizome webpage.
...	Ignored.

Value

Data frame with gene regulatory interactions in NicheNet format.

See Also

- nichenet_gr_network
- harmonizome_download

Examples

```
# use only JASPAR and TRANSFAC:
hz_gr_network <- nichenet_gr_network_harmonizome(
    datasets = c('jasparpwm', 'transfac', 'transfacpwm')
)
```

---

nichenet_gr_network_htridb

NicheNet gene regulatory network from HTRIdb

Description

Builds a gene regulatory network using data from the HTRIdb database and converts it to a format suitable for NicheNet.

Usage

```
nichenet_gr_network_htridb()
```

Value

Data frame with gene regulatory interactions in NicheNet format.

See Also

htridb_download, nichenet_gr_network

Examples

```
htri_gr_network <- nichenet_gr_network_htridb()
```

nichenet_gr_network_omnipath *Builds gene regulatory network for NicheNet using OmniPath*

Description

Retrieves network prior knowledge from OmniPath and provides it in a format suitable for NicheNet. This method never downloads the 'ligrecextra' dataset because the ligand-receptor interactions are supposed to come from nichenet_lr_network_omnipath.

Usage

```
nichenet_gr_network_omnipath(min_curation_effort = 0, ...)
```

Arguments

min_curation_effort	Lower threshold for curation effort
...	Passed to import_transcriptional_interactions

Value

A network data frame (tibble) with gene regulatory interactions suitable for use with NicheNet.

See Also

- nichenet_gr_network_evex
- nichenet_gr_network_harmonizome
- nichenet_gr_network_htridb
- nichenet_gr_network_omnipath
- nichenet_gr_network_pathwaycommons
- nichenet_gr_network_regnetwork
- nichenet_gr_network_remap
- nichenet_gr_network_trrust

Examples

```
# use interactions up to confidence level "C" from DoRothEA:
op_gr_network <- nichenet_gr_network_omnipath(
    dorothea_levels = c('A', 'B', 'C')
)
```

nichenet_gr_network_pathwaycommons

NicheNet gene regulatory network from PathwayCommons

Description

Builds gene regulation prior knowledge for NicheNet using PathwayCommons.

Usage

```
nichenet_gr_network_pathwaycommons(
  interaction_types = "controls-expression-of",
  ...
)
```

Arguments

interaction_types	Character vector with PathwayCommons interaction types. Please refer to the default value and the PathwayCommons webpage.
...	Ignored.

Value

Data frame with gene regulatory interactions in NicheNet format.

See Also

- nichenet_gr_network
- pathwaycommons_download

Examples

```
pc_gr_network <- nichenet_gr_network_pathwaycommons()
```

nichenet_gr_network_regnetwork

NicheNet gene regulatory network from RegNetwork

Description

Builds a gene regulatory network using data from the RegNetwork database and converts it to a format suitable for NicheNet.

Usage

```
nichenet_gr_network_regnetwork()
```

Value

Data frame with gene regulatory interactions in NicheNet format.

See Also

- regnetwork_download
- nichenet_gr_network

Examples

```
regn_gr_network <- nichenet_gr_network_regnetwork()
```

---

nichenet_gr_network_remap

NicheNet gene regulatory network from ReMap

Description

Builds a gene regulatory network using data from the ReMap database and converts it to a format suitable for NicheNet.

Usage

```
nichenet_gr_network_remap(
  score = 100,
  top_targets = 500,
  only_known_tfs = TRUE
)
```

Arguments

score	Numeric: a minimum score between 0 and 1000, records with lower scores will be excluded. If NULL no filtering performed.
top_targets	Numeric: the number of top scoring targets for each TF. Essentially the maximum number of targets per TF. If NULL the number of targets is not restricted.
only_known_tfs	Logical: whether to exclude TFs which are not in TF census.

Value

Data frame with gene regulatory interactions in NicheNet format.

See Also

- remap_filtered
- nichenet_gr_network

Examples

```
## Not run:
# use only max. top 100 targets for each TF:
remap_gr_network <- nichenet_gr_network_remap(top_targets = 100)

## End(Not run)
```

---

nichenet_gr_network_trrust

NicheNet gene regulatory network from TRRUST

Description

Builds a gene regulatory network using data from the TRRUST database and converts it to a format suitable for NicheNet.

Usage

```
nichenet_gr_network_trrust()
```

Value

Data frame with gene regulatory interactions in NicheNet format.

See Also

- trrust_download
- nichenet_gr_network

Examples

```
trrust_gr_network <- nichenet_gr_network_trrust()
```

nichenet_ligand_activities *Calls the NicheNet ligand activity analysis*

Description

Calls the NicheNet ligand activity analysis

Usage

```
nichenet_ligand_activities(
  ligand_target_matrix,
  lr_network,
  expressed_genes_transmitter,
  expressed_genes_receiver,
  genes_of_interest,
  background_genes = NULL,
  n_top_ligands = 42,
  n_top_targets = 250
)
```

Arguments

ligand_target_matrix	A matrix with rows and columns corresponding to ligands and targets, respectively. Produced by nichenet_ligand_target_matrix or nichenetr::construct_ligand_target_ma
lr_network	A data frame with ligand-receptor interactions, as produced by nichenet_lr_network.
expressed_genes_transmitter	Character vector with the gene symbols of the genes expressed in the cells transmitting the signal.
expressed_genes_receiver	Character vector with the gene symbols of the genes expressed in the cells receiving the signal.
genes_of_interest	Character vector with the gene symbols of the genes of interest. These are the genes in the receiver cell population that are potentially affected by ligands expressed by interacting cells (e.g. genes differentially expressed upon cell-cell interaction).
background_genes	Character vector with the gene symbols of the genes to be used as background.
n_top_ligands	How many of the top ligands to include in the ligand-target table.
n_top_targets	For each ligand, how many of the top targets to include in the ligand-target table.

Value

A named list with 'ligand_activities' (a tibble giving several ligand activity scores; following columns in the tibble: $test_ligand, $auroc, $aupr and $pearson) and 'ligand_target_links' (a tibble with columns ligand, target and weight (i.e. regulatory potential score)).

Examples

```
## Not run:
networks <- nichenet_networks()
expression <- nichenet_expression_data()
optimization_results <- nichenet_optimization(networks, expression)
nichenet_model <- nichenet_build_model(optimization_results, networks)
lt_matrix <- nichenet_ligand_target_matrix(
    nichenet_model$weighted_networks,
    networks$lr_network,
    nichenet_model$optimized_parameters
)
ligand_activities <- nichenet_ligand_activities(
    ligand_target_matrix = lt_matrix,
    lr_network = networks$lr_network,
    # the rest of the parameters should come
    # from your transcriptomics data:
    expressed_genes_transmitter = expressed_genes_transmitter,
    expressed_genes_receiver = expressed_genes_receiver,
    genes_of_interest = genes_of_interest
)

## End(Not run)
```

---

nichenet_ligand_target_links

Compiles a table with weighted ligand-target links

Description

A wrapper around `nichenetr::get_weighted_ligand_target_links` to compile a data frame with weighted links from the top ligands to their top targets.

Usage

```
nichenet_ligand_target_links(
  ligand_activities,
  ligand_target_matrix,
  genes_of_interest,
  n_top_ligands = 42,
  n_top_targets = 250
)
```

Arguments

ligand_activities	Ligand activity table as produced by nichenetr::predict_ligand_activities.
ligand_target_matrix	Ligand-target matrix as produced by nichenetr::construct_ligand_target_matrix or the wrapper around it in the current package: nichenet_ligand_target_matrix.
genes_of_interest	Character vector with the gene symbols of the genes of interest. These are the genes in the receiver cell population that are potentially affected by ligands expressed by interacting cells (e.g. genes differentially expressed upon cell-cell interaction).
n_top_ligands	How many of the top ligands to include in the ligand-target table.
n_top_targets	For each ligand, how many of the top targets to include in the ligand-target table.

Value

A tibble with columns ligand, target and weight (i.e. regulatory potential score).

Examples

```
## Not run:
networks <- nichenet_networks()
expression <- nichenet_expression_data()
optimization_results <- nichenet_optimization(networks, expression)
nichenet_model <- nichenet_build_model(optimization_results, networks)
lt_matrix <- nichenet_ligand_target_matrix(
    nichenet_model$weighted_networks,
    networks$lr_network,
    nichenet_model$optimized_parameters
)
ligand_activities <- nichenet_ligand_activities(
    ligand_target_matrix = lt_matrix,
    lr_network = networks$lr_network,
    # the rest of the parameters should come
    # from your transcriptomics data:
    expressed_genes_transmitter = expressed_genes_transmitter,
    expressed_genes_receiver = expressed_genes_receiver,
    genes_of_interest = genes_of_interest
)
lt_links <- nichenet_ligand_target_links(
    ligand_activities = ligand_activities,
    ligand_target_matrix = lt_matrix,
    genes_of_interest = genes_of_interest,
    n_top_ligands = 20,
    n_top_targets = 100
)

## End(Not run)
```

nichenet_ligand_target_matrix *Creates a NicheNet ligand-target matrix*

Description

Creates a NicheNet ligand-target matrix

Usage

```
nichenet_ligand_target_matrix(
  weighted_networks,
  lr_network,
  optimized_parameters,
  use_weights = TRUE,
  construct_ligand_target_matrix_param = list()
)
```

Arguments

weighted_networks	Weighted networks as provided by nichenet_build_model.
lr_network	A data frame with ligand-receptor interactions, as produced by nichenet_lr_network.
optimized_parameters	The outcome of NicheNet parameter optimization as produced by nichenet_build_model.
use_weights	Logical: wether the network sources are weighted. In this function it only affects the output file name.
construct_ligand_target_matrix_param	Override parameters for nichenetr::construct_ligand_target_matrix.

Value

A matrix containing ligand-target probability scores.

Examples

```
## Not run:
networks <- nichenet_networks()
expression <- nichenet_expression_data()
optimization_results <- nichenet_optimization(networks, expression)
nichenet_model <- nichenet_build_model(optimization_results, networks)
lt_matrix <- nichenet_ligand_target_matrix(
    nichenet_model$weighted_networks,
    networks$lr_network,
    nichenet_model$optimized_parameters
)
```

```
## End(Not run)
```

`nichenet_lr_network` *Builds a NicheNet ligand-receptor network*

Description

Builds ligand-receptor network prior knowledge for NicheNet using multiple resources.

Usage

```
nichenet_lr_network(
  omnipath = list(),
  guide2pharma = list(),
  ramilowski = list(),
  only_omnipath = FALSE,
  quality_filter_param = list()
)
```

Arguments

`omnipath`	List with paramaters to be passed to `nichenet_lr_network_omnipath`.
`guide2pharma`	List with paramaters to be passed to `nichenet_lr_network_guide2pharma`.
`ramilowski`	List with paramaters to be passed to `nichenet_lr_network_ramilowski`.
`only_omnipath`	Logical: a shortcut to use only OmniPath as network resource.
`quality_filter_param`	Arguments for `filter_intercell_network` (quality filtering of the OmniPath ligand-receptor network). It is recommended to check these parameters and apply some quality filtering. The defaults already ensure certain filtering, but you might want more relaxed or stringent options.

Value

A network data frame (tibble) with ligand-receptor interactions suitable for use with NicheNet.

See Also

- `nichenet_lr_network_omnipath`
- `nichenet_lr_network_guide2pharma`
- `nichenet_lr_network_ramilowski`
- `filter_intercell_network`

Examples

```
# load everything with the default parameters:
lr_network <- nichenet_lr_network()

# don't use Ramilowski:
lr_network <- nichenet_lr_network(ramilowski = NULL)

# use only OmniPath:
lr_network_omnipath <- nichenet_lr_network(only_omnipath = TRUE)
```

---

nichenet_lr_network_guide2pharma

Ligand-receptor network from Guide to Pharmacology

Description

Downloads ligand-receptor interactions from the Guide to Pharmacology database and converts it to a format suitable for NicheNet.

Usage

```
nichenet_lr_network_guide2pharma()
```

Value

Data frame with ligand-receptor interactions in NicheNet format.

See Also

nichenet_lr_network, guide2pharma_download

Examples

```
g2p_lr_network <- nichenet_lr_network_guide2pharma()
```

nichenet_lr_network_omnipath *Builds ligand-receptor network for NicheNet using OmniPath*

Description

Retrieves network prior knowledge from OmniPath and provides it in a format suitable for NicheNet. This method never downloads the 'ligrecextra' dataset because the ligand-receptor interactions are supposed to come from nichenet_lr_network_omnipath.

Usage

```
nichenet_lr_network_omnipath(quality_filter_param = list(), ...)
```

Arguments

quality_filter_param	List with arguments for filter_intercell_network. It is recommended to check these parameters and apply some quality filtering. The defaults already ensure certain filtering, but you might want more relaxed or stringent options.
...	Passed to import_intercell_network

Value

A network data frame (tibble) with ligand-receptor interactions suitable for use with NicheNet.

See Also

- nichenet_lr_network
- import_intercell_network

Examples

```
# use only ligand-receptor interactions (not for example ECM-adhesion):
op_lr_network <- nichenet_lr_network_omnipath(ligand_receptor = TRUE)

# use only CellPhoneDB and Guide to Pharmacology:
op_lr_network <- nichenet_lr_network_omnipath(
    resources = c('CellPhoneDB', 'Guide2Pharma')
)

# only interactions where the receiver is a transporter:
op_lr_network <- nichenet_lr_network_omnipath(
    receiver_param = list(parent = 'transporter')
)
```

nichenet_lr_network_ramilowski *Ligand-receptor network from Ramilowski 2015*

Description

Downloads ligand-receptor interactions from Supplementary Table 2 of the paper 'A draft network of ligand–receptor-mediated multicellular signalling in human' (Ramilowski et al. 2015, https://www.nature.com/articles/ncomms8866). It converts the downloaded table to a format suitable for NicheNet.

Usage

```
nichenet_lr_network_ramilowski(
  evidences = c("literature supported", "putative")
)
```

Arguments

evidences	Character: evidence types, "literature supported", "putative" or both.

Value

Data frame with ligand-receptor interactions in NicheNet format.

See Also

- nichenet_lr_network
- ramilowski_download

Examples

```
# use only the literature supported data:
rami_lr_network <- nichenet_lr_network_ramilowski(
    evidences = 'literature supported'
)
```

nichenet_main *Executes the full NicheNet pipeline*

Description

Builds all prior knowledge data required by NicheNet. For this it calls a multitude of methods to download and combine data from various databases according to the settings. The content of the prior knowledge data is highly customizable, see the documentation of the related functions. After the prior knowledge is ready, it performs parameter optimization to build a NicheNet model. This results a weighted ligand- target matrix. Then, considering the expressed genes from user provided data, a gene set of interest and background genes, it executes the NicheNet ligand activity analysis.

Usage

```
nichenet_main(
  only_omnipath = FALSE,
  expressed_genes_transmitter = NULL,
  expressed_genes_receiver = NULL,
  genes_of_interest = NULL,
  background_genes = NULL,
  use_weights = TRUE,
  n_top_ligands = 42,
  n_top_targets = 250,
  signaling_network = list(),
  lr_network = list(),
  gr_network = list(),
  small = FALSE,
  tiny = FALSE,
  make_multi_objective_function_param = list(),
  objective_function_param = list(),
  mlrmbo_optimization_param = list(),
  construct_ligand_target_matrix_param = list(),
  results_dir = NULL,
  quality_filter_param = list()
)
```

Arguments

only_omnipath	Logical: use only OmniPath for network knowledge. This is a simple switch for convenience, further options are available by the other arguments. By default we use all available resources. The networks can be customized on a resource by resource basis, as well as providing custom parameters for individual resources, using the parameters 'signaling_network', 'lr_network' and 'gr_network'.
expressed_genes_transmitter	Character vector with the gene symbols of the genes expressed in the cells transmitting the signal.

`expressed_genes_receiver`
: Character vector with the gene symbols of the genes expressed in the cells receiving the signal.

`genes_of_interest`
: Character vector with the gene symbols of the genes of interest. These are the genes in the receiver cell population that are potentially affected by ligands expressed by interacting cells (e.g. genes differentially expressed upon cell-cell interaction).

`background_genes`
: Character vector with the gene symbols of the genes to be used as background.

`use_weights`
: Logical: calculate and use optimized weights for resources (i.e. one resource seems to be better than another, hence the former is considered with a higher weight).

`n_top_ligands`
: How many of the top ligands to include in the ligand-target table.

`n_top_targets`
: How many of the top targets (for each of the top ligands) to consider in the ligand-target table.

`signaling_network`
: A list of parameters for building the signaling network, passed to `nichenet_signaling_network`.

`lr_network`
: A list of parameters for building the ligand-receptor network, passed to `nichenet_lr_network`.

`gr_network`
: A list of parameters for building the gene regulatory network, passed to `nichenet_gr_network`.

`small`
: Logical: build a small network for testing purposes, using only OmniPath data. It is also a high quality network, it is reasonable to try the analysis with this small network.

`tiny`
: Logical: build an even smaller network for testing purposes. As this involves random subsetting, it's not recommended to use this network for analysis.

`make_multi_objective_function_param`
: Override parameters for `smoof::makeMultiObjectiveFunction`.

`objective_function_param`
: Override additional arguments passed to the objective function.

`mlrmbo_optimization_param`
: Override arguments for `nichenetr::mlrmbo_optimization`.

`construct_ligand_target_matrix_param`
: Override parameters for `nichenetr::construct_ligand_target_matrix`.

`results_dir`
: Character: path to the directory to save intermediate and final outputs from NicheNet methods.

`quality_filter_param`
: Arguments for `filter_intercell_network` (quality filtering of the OmniPath ligand-receptor network). It is recommended to check these parameters and apply some quality filtering. The defaults already ensure certain filtering, but you might want more relaxed or stringent options.

Details

About *small* and *tiny* networks: Building a NicheNet model is computationally demanding, taking several hours to run. As this is related to the enormous size of the networks, to speed up testing we

can use smaller networks, around 1,000 times smaller, with few thousands of interactions instead of few millions. Random subsetting of the whole network would result disjunct fragments, instead we load only a few resources. To run the whole pipeline with tiny networks use nichenet_test.

Value

A named list with the intermediate and final outputs of the pipeline: 'networks', 'expression', 'optimized_parameters', 'weighted_networks' and 'ligand_target_matrix'.

See Also

- nichenet_networks
- nichenet_signaling_network
- nichenet_lr_network
- nichenet_gr_network
- nichenet_test
- nichenet_workarounds
- nichenet_results_dir

Examples

```
## Not run:
nichenet_results <- nichenet_main(
    # altering some network resource parameters, the rest
    # of the resources will be loaded according to the defaults
    signaling_network = list(
        cpdb = NULL, # this resource will be excluded
        inbiomap = NULL,
        evex = list(min_confidence = 1.0) # override some parameters
    ),
    gr_network = list(only_omnipath = TRUE),
    n_top_ligands = 20,
    # override the default number of CPU cores to use
    mlrmbo_optimization_param = list(ncores = 4)
)

## End(Not run)
```

---

nichenet_networks *Builds NicheNet network prior knowledge*

Description

Builds network knowledge required by NicheNet. For this it calls a multitude of methods to download and combine data from various databases according to the settings. The content of the prior knowledge data is highly customizable, see the documentation of the related functions.

Usage

```
nichenet_networks(
  signaling_network = list(),
  lr_network = list(),
  gr_network = list(),
  only_omnipath = FALSE,
  small = FALSE,
  tiny = FALSE,
  quality_filter_param = list()
)
```

Arguments

signaling_network	A list of parameters for building the signaling network, passed to nichenet_signaling_network
lr_network	A list of parameters for building the ligand-receptor network, passed to nichenet_lr_network
gr_network	A list of parameters for building the gene regulatory network, passed to nichenet_gr_network
only_omnipath	Logical: a shortcut to use only OmniPath as network resource.
small	Logical: build a small network for testing purposes, using only OmniPath data. It is also a high quality network, it is reasonable to try the analysis with this small network.
tiny	Logical: build an even smaller network for testing purposes. As this involves random subsetting, it's not recommended to use this network for analysis.
quality_filter_param	Arguments for filter_intercell_network (quality filtering of the OmniPath ligand-receptor network). It is recommended to check these parameters and apply some quality filtering. The defaults already ensure certain filtering, but you might want more relaxed or stringent options.

Value

A named list with three network data frames (tibbles): the signaling, the ligand-receptor (lr) and the gene regulatory (gr) networks.

See Also

- nichenet_signaling_network
- nichenet_lr_network
- nichenet_gr_network

Examples

```
## Not run:
networks <- nichenet_networks()
dplyr::sample_n(networks$gr_network, 10)
# # A tibble: 10 x 4
#    from    to       source              database
```

```
#    <chr>   <chr>    <chr>                <chr>
#  1 MAX     ALG3     harmonizome_ENCODE   harmonizome
#  2 MAX     IMPDH1   harmonizome_ENCODE   harmonizome
#  3 SMAD5   LCP1     Remap_5              Remap
#  4 HNF4A   TNFRSF19 harmonizome_CHEA     harmonizome
#  5 SMC3    FAP      harmonizome_ENCODE   harmonizome
#  6 E2F6    HIST1H1B harmonizome_ENCODE   harmonizome
#  7 TFAP2C  MAT2B    harmonizome_ENCODE   harmonizome
#  8 USF1    TBX4     harmonizome_TRANSFAC harmonizome
#  9 MIR133B FETUB    harmonizome_TRANSFAC harmonizome
# 10 SP4     HNRNPH2  harmonizome_ENCODE   harmonizome

## End(Not run)

# use only OmniPath:
omnipath_networks <- nichenet_networks(only_omnipath = TRUE)
```

`nichenet_optimization` *Optimizes NicheNet model parameters*

Description

Optimize NicheNet method parameters, i.e. PageRank parameters and source weights, basedon a collection of experiments where the effect of a ligand on gene expression was measured.

Usage

```
nichenet_optimization(
  networks,
  expression,
  make_multi_objective_function_param = list(),
  objective_function_param = list(),
  mlrmbo_optimization_param = list()
)
```

Arguments

`networks`	A list with NicheNet format signaling, ligand-receptor and gene regulatory networks as produced by `nichenet_networks`.
`expression`	A list with expression data from ligand perturbation experiments, as produced by `nichenet_expression_data`.
`make_multi_objective_function_param`	Override parameters for `smoof::makeMultiObjectiveFunction`.
`objective_function_param`	Override additional arguments passed to the objective function.
`mlrmbo_optimization_param`	Override arguments for `nichenetr::mlrmbo_optimization`.

Value

A result object from the function 'mlrMBO::mbo'. Among other things, this contains the optimal parameter settings, the output corresponding to every input etc.

Examples

```
## Not run:
networks <- nichenet_networks()
expression <- nichenet_expression_data()
optimization_results <- nichenet_optimization(networks, expression)

## End(Not run)
```

---

nichenet_remove_orphan_ligands

Removes experiments with orphan ligands

Description

Removes from the expression data the perturbation experiments involving ligands without connections.

Usage

```
nichenet_remove_orphan_ligands(expression, lr_network)
```

Arguments

expression	Expression data as returned by nichenet_expression_data.
lr_network	A NicheNet format ligand-recptor network data frame as produced by nichenet_lr_network.

Value

The same list as 'expression' with certain elements removed.

Examples

```
## Not run:
lr_network <- nichenet_lr_network()
expression <- nichenet_expression_data()
expression <- nichenet_remove_orphan_ligands(expression, lr_network)

## End(Not run)
```

nichenet_results_dir *Path to the current NicheNet results directory*

Description

Path to the directory to save intermediate and final outputs from NicheNet methods.

Usage

```
nichenet_results_dir()
```

Value

Character: path to the NicheNet results directory.

Examples

```
nichenet_results_dir()
# [1] "nichenet_results"
```

nichenet_signaling_network *Builds a NicheNet signaling network*

Description

Builds signaling network prior knowledge for NicheNet using multiple resources.

Usage

```
nichenet_signaling_network(
  omnipath = list(),
  pathwaycommons = list(),
  harmonizome = list(),
  vinayagam = list(),
  cpdb = list(),
  evex = list(),
  inbiomap = list(),
  only_omnipath = FALSE
)
```

Arguments

omnipath	List with paramaters to be passed to nichenet_signaling_network_omnipath.
pathwaycommons	List with paramaters to be passed to nichenet_signaling_network_pathwaycommons.
harmonizome	List with paramaters to be passed to nichenet_signaling_network_harmonizome.
vinayagam	List with paramaters to be passed to nichenet_signaling_network_vinayagam.
cpdb	List with paramaters to be passed to nichenet_signaling_network_cpdb.
evex	List with paramaters to be passed to nichenet_signaling_network_evex.
inbiomap	List with paramaters to be passed to nichenet_signaling_network_inbiomap.
only_omnipath	Logical: a shortcut to use only OmniPath as network resource.

Value

A network data frame (tibble) with signaling interactions suitable for use with NicheNet.

See Also

- nichenet_signaling_network_omnipath
- nichenet_signaling_network_pathwaycommons
- nichenet_signaling_network_harmonizome
- nichenet_signaling_network_vinayagam
- nichenet_signaling_network_cpdb
- nichenet_signaling_network_evex
- nichenet_signaling_network_inbiomap

Examples

```
# load everything with the default parameters:
# we don't load inBio Map due to the - hopefully
# temporary - issues of their server
sig_network <- nichenet_signaling_network(inbiomap = NULL, cpdb = NULL)

# override parameters for some resources:
sig_network <- nichenet_signaling_network(
    omnipath = list(resources = c('SIGNOR', 'SignaLink3', 'SPIKE')),
    pathwaycommons = NULL,
    harmonizome = list(datasets = c('phosphositeplus', 'depod')),
    # we can not include this in everyday tests as it takes too long:
    # cpdb = list(complex_max_size = 1, min_score = .98),
    cpdb = NULL,
    evex = list(min_confidence = 1.5),
    inbiomap = NULL
)

# use only OmniPath:
sig_network_omnipath <- nichenet_signaling_network(only_omnipath = TRUE)
```

nichenet_signaling_network_cpdb *Builds signaling network for NicheNet using ConsensusPathDB*

Description

Builds signaling network prior knowledge using ConsensusPathDB (CPDB) data. Note, the interactions from CPDB are not directed and many of them comes from complex expansion. Find out more at http://cpdb.molgen.mpg.de/.

Usage

```
nichenet_signaling_network_cpdb(...)
```

Arguments

...	Passed to consensuspathdb_download.

Value

A network data frame (tibble) with signaling interactions suitable for use with NicheNet.

See Also

- nichenet_signaling_network
- consensuspathdb_download

Examples

```
## Not run:
# use some parameters stricter than default:
cpdb_signaling_network <- nichenet_signaling_network_cpdb(
    complex_max_size = 2,
    min_score = .99
)

## End(Not run)
```

nichenet_signaling_network_evex *NicheNet signaling network from EVEX*

Description

Builds signaling network prior knowledge for NicheNet from the EVEX database.

Usage

```
nichenet_signaling_network_evex(top_confidence = 0.75, indirect = FALSE, ...)
```

Arguments

top_confidence	Double, between 0 and 1. Threshold based on the quantile of the confidence score.
indirect	Logical: whether to include indirect interactions.
...	Ignored.

Value

A network data frame (tibble) with signaling interactions suitable for use with NicheNet.

See Also

- evex_download
- nichenet_signaling_network

Examples

```
ev_signaling_network <- nichenet_signaling_network_evex(
    top_confidence = .9
)
```

nichenet_signaling_network_harmonizome *NicheNet signaling network from Harmonizome*

Description

Builds signaling network prior knowledge for NicheNet using Harmonizome

Usage

```
nichenet_signaling_network_harmonizome(
  datasets = c("phosphositeplus", "kea", "depod"),
  ...
)
```

Arguments

datasets	The datasets to use. For possible values please refer to default value and the Harmonizome webpage.
...	Ignored.

Value

A network data frame (tibble) with signaling interactions suitable for use with NicheNet.

Examples

```
# use only KEA and PhosphoSite:
hz_signaling_network <- nichenet_signaling_network_harmonizome(
    datasets = c('kea', 'phosphositeplus')
)
```

---

nichenet_signaling_network_inbiomap

NicheNet signaling network from InWeb InBioMap

Description

Builds signaling network prior knowledge for NicheNet from the InWeb InBioMap database.

Usage

```
nichenet_signaling_network_inbiomap(...)
```

Arguments

...	Ignored.

Value

A network data frame (tibble) with signaling interactions suitable for use with NicheNet.

See Also

nichenet_signaling_network, inbiomap_download

Examples

```
## Not run:
ib_signaling_network <- nichenet_signaling_network_inbiomap()

## End(Not run)
```

`nichenet_signaling_network_omnipath` *Builds signaling network for NicheNet using OmniPath*

Description

Retrieves network prior knowledge from OmniPath and provides it in a format suitable for NicheNet. This method never downloads the 'ligrecextra' dataset because the ligand-receptor interactions are supposed to come from `nichenet_lr_network_omnipath`.

Usage

```
nichenet_signaling_network_omnipath(min_curation_effort = 0, ...)
```

Arguments

`min_curation_effort`	Lower threshold for curation effort
`...`	Passed to `import_post_translational_interactions`

Value

A network data frame (tibble) with signaling interactions suitable for use with NicheNet.

See Also

- `nichenet_signaling_network`

Examples

```
# use interactions with at least 2 evidences (reference or database)
op_signaling_network <- nichenet_signaling_network_omnipath(
    min_curation_effort = 2
)
```

nichenet_signaling_network_pathwaycommons

NicheNet signaling network from PathwayCommons

Description

Builds signaling network prior knowledge for NicheNet using PathwayCommons.

Usage

```
nichenet_signaling_network_pathwaycommons(
  interaction_types = c("catalysis-precedes", "controls-phosphorylation-of",
    "controls-state-change-of", "controls-transport-of", "in-complex-with",
    "interacts-with"),
  ...
)
```

Arguments

interaction_types	Character vector with PathwayCommons interaction types. Please refer to the default value and the PathwayCommons webpage.
...	Ignored.

Value

A network data frame (tibble) with signaling interactions suitable for use with NicheNet.

Examples

```
# use only the "controls-transport-of" interactions:
pc_signaling_network <- nichenet_signaling_network_pathwaycommons(
    interaction_types = 'controls-transport-of'
)
```

nichenet_signaling_network_vinayagam

NicheNet signaling network from Vinayagam

Description

Builds signaling network prior knowledge for NicheNet using Vinayagam 2011 Supplementary Table S6. Find out more at https://doi.org/10.1126/scisignal.2001699.

Usage

```
nichenet_signaling_network_vinayagam(...)
```

Arguments

...	Ignored.

Value

A network data frame (tibble) with signaling interactions suitable for use with NicheNet.

Examples

```
vi_signaling_network <- nichenet_signaling_network_vinayagam()
```

---

nichenet_test *Run the NicheNet pipeline with a little dummy network*

Description

Loads a tiny network and runs the NicheNet pipeline with low number of iterations in the optimization process. This way the pipeline runs in a reasonable time in order to test the code. Due to the random subsampling disconnected networks might be produced sometimes. If you see an error like "Error in if (sd(prediction_vector) == 0) ... missing value where TRUE/FALSE needed", the random subsampled input is not appropriate. In this case just interrupt and call again. This test ensures the computational integrity of the pipeline. If it fails during the optimization process, try to start it over several times, even restarting R. The unpredictability is related to codemlrMBO and `nichenetr` not being prepared to handle certain conditions, and it's also difficult to find out which conditions lead to which errors. At least 3 different errors appear time to time, depending on the input. It also seems like restarting R sometimes helps, suggesting that the entire system might be somehow stateful. You can ignore the `Parallelization was not stopped` warnings on repeated runs.

Usage

```
nichenet_test(...)
```

Arguments

...	Passed to `nichenet_main`.

Value

A named list with the intermediate and final outputs of the pipeline: 'networks', 'expression', 'optimized_parameters', 'weighted_networks' and 'ligand_target_matrix'.

Examples

```
## Not run:
nnt <- nichenet_test()

## End(Not run)
```

`nichenet_workarounds` *Workarounds using NicheNet without attaching the package*

Description

NicheNet requires the availability of some lazy loaded external data which are not available if the package is not loaded and attached. Also, the `BBmisc::convertToShortString` used for error reporting in `mlrMBO::evalTargetFun.OptState` is patched here to print longer error messages. Maybe it's a better solution to attach `nichenetr` before running the NicheNet pipeline. Alternatively you can try to call this function in the beginning. Why we don't call this automatically is just because we don't want to load datasets from another package without the user knowing about it.

Usage

```
nichenet_workarounds()
```

Value

Returns `NULL`.

Examples

```
## Not run:
nichenet_workarounds()

## End(Not run)
```

`obo_parser` *Generic OBO parser*

Description

Reads the contents of an OBO file and processes it into data frames or a list based data structure.

Usage

```
obo_parser(
  path,
 relations = c("is_a", "part_of", "occurs_in", "regulates", "positively_regulates",
    "negatively_regulates"),
  shorten_namespace = TRUE,
  tables = TRUE
)
```

Arguments

path	Path to the OBO file.
relations	Character vector: process only these relations.
shorten_namespace	Logical: shorten the namespace to a single letter code (as usual for Gene Ontology, e.g. cellular_component = "C").
tables	Logical: return data frames (tibbles) instead of nested lists.

Value

A list with the following elements: 1) "names" a list with terms as names and names as values; 2) "namespaces" a list with terms as names and namespaces as values; 3) "relations" a list with relations between terms: terms are keys, values are lists with relations as names and character vectors of related terms as values; 4) "subsets" a list with terms as keys and character vectors of subset names as values (or NULL if the term does not belong to any subset); 5) "obsolete" character vector with all the terms labeled as obsolete. If the tables parameter is TRUE, "names", "namespaces", "relations" and "subsets" will be data frames (tibbles).

See Also

- relations_list_to_table
- relations_table_to_list
- swap_relations

Examples

```
goslim_url <-
    "http://current.geneontology.org/ontology/subsets/goslim_generic.obo"
path <- tempfile()
httr::GET(goslim_url, httr::write_disk(path, overwrite = TRUE))
obo <- obo_parser(path, tables = FALSE)
unlink(path)
names(obo)
# [1] "names"      "namespaces" "relations"  "subsets"    "obsolete"
head(obo$relations, n = 2)
# $`GO:0000001`
# $`GO:0000001`$is_a
# [1] "GO:0048308" "GO:0048311"
```

```
#
# $`GO:0000002`
# $`GO:0000002`$is_a
# [1] "GO:0007005"
```

`OmnipathR` *The OmnipathR package*

Description

OmnipathR is an R package built to provide easy access to the data stored in the OmniPath web service:

https://omnipathdb.org/

And a number of other resources, such as BioPlex, ConsensusPathDB, EVEX, Guide to Pharmacology (IUPHAR/BPS), Harmonizome, HTRIdb, InWeb InBioMap, KEGG Pathway, Pathway Commons, Ramilowski et al. 2015, RegNetwork, ReMap, TF census, TRRUST and Vinayagam et al. 2011.

The OmniPath web service implements a very simple REST style API. This package make requests by the HTTP protocol to retreive the data. Hence, fast Internet access is required for a propser use of OmnipathR.

The package also provides some utility functions to filter, analyse and visualize the data. Furthermore, OmnipathR features a close integration with the NicheNet method for ligand activity prediction from transcriptomics data, and its R implementation nichenetr (available in CRAN).

Author(s)

Alberto Valdeolivas <<alvaldeolivas@gmail>> and Denes Turei <<turei.denes@gmail.com>> and Attila Gabor <<gaborattila87@gmail.com>>

Examples

```
## Not run:
# Download post-translational modifications:
enzsub <- import_omnipath_enzsub(resources = c("PhosphoSite", "SIGNOR"))

# Download protein-protein interactions
interactions <- import_omnipath_interactions(resources = c("SignaLink3"))

# Convert to igraph objects:
enzsub_g <- enzsub_graph(enzsub = enzsub)
OPI_g <- interaction_graph(interactions = interactions )

# Print some interactions:
print_interactions(head(ptms))

# interactions with references:
```

```
print_interactions(tail(ptms),writeRefs=TRUE)

# find interactions between kinase and substrate:
print_interactions(dplyr::filter(ptms,enzyme_genesymbol=="MAP2K1",
   substrate_genesymbol=="MAPK3"))

# find shortest paths on the directed network between proteins
print_path_es(shortest_paths(OPI_g, from = "TYRO3", to = "STAT3",
   output = 'epath')$epath[[1]], OPI_g)

# find all shortest paths between proteins
print_path_vs(
    all_shortest_paths(
        enzsub_g,
        from = "SRC",
        to = "STAT1"
    )$res,
    enzsub_g
)

## End(Not run)
```

`omnipath_cache_autoclean` *Keeps only the latest versions of complete downloads*

Description

Removes the old versions, the failed downloads and the files in the cache directory which are missing from the database. For more flexible operations use `omnipath_cache_remove` and `omnipath_cache_clean`.

Usage

```
omnipath_cache_autoclean()
```

Value

Invisibl returns the cache database (list of cache records).

Examples

```
## Not run:
omnipath_cache_autoclean()

## End(Not run)
```

omnipath_cache_clean *Removes the items from the cache directory which are unknown by the cache database*

Description

Removes the items from the cache directory which are unknown by the cache database

Usage

```
omnipath_cache_clean()
```

Value

Returns 'NULL'.

Examples

```
omnipath_cache_clean()
```

omnipath_cache_clean_db *Removes the cache database entries without existing files*

Description

Removes the cache database entries without existing files

Usage

```
omnipath_cache_clean_db(...)
```

Arguments

...	Ignored.

Value

Returns 'NULL'.

Examples

```
omnipath_cache_clean_db()
```

omnipath_cache_download_ready *Sets the download status to ready for a cache item*

Description

Sets the download status to ready for a cache item

Usage

```
omnipath_cache_download_ready(version, key = NULL)
```

Arguments

version	Version of the cache item. If does not exist a new version item will be created
key	Key of the cache item

Value

Character: invisibly returns the version number of the cache version item.

Examples

```
bioc_url <- 'https://bioconductor.org/'
# request a new version item (or retrieve the latest)
new_version <- omnipath_cache_latest_or_new(url = bioc_url)
# check if the version item is not a finished download
new_version$status
# [1] "unknown"
# download the file
httr::GET(bioc_url, httr::write_disk(new_version$path, overwrite = TRUE))
# report to the cache database that the download is ready
omnipath_cache_download_ready(new_version)
# now the status is ready:
version <- omnipath_cache_latest_or_new(url = bioc_url)
version$status
# "ready"
version$dl_finished
# [1] "2021-03-09 16:48:38 CET"
omnipath_cache_remove(url = bioc_url) # cleaning up
```

`omnipath_cache_filter_versions` *Filters the versions from one cache record*

Description

Filters the versions based on multiple conditions: their age and status

Usage

```
omnipath_cache_filter_versions(
  record,
  latest = FALSE,
  max_age = NULL,
  min_age = NULL,
  status = CACHE_STATUS$READY
)
```

Arguments

`record`	A cache record
`latest`	Return the most recent version
`max_age`	The maximum age in days (e.g. 5: 5 days old or more recent)
`min_age`	The minimum age in days (e.g. 5: 5 days old or older)
`status`	Character vector with status codes. By default only the versions with 'ready' (completed download) status are selected

Value

Character vector with version IDs, NA if no version satisfies the conditions.

Examples

```
# creating an example cache record
bioc_url <- 'https://bioconductor.org/'
version <- omnipath_cache_latest_or_new(url = bioc_url)
httr::GET(bioc_url, httr::write_disk(version$path, overwrite = TRUE))
omnipath_cache_download_ready(version)
record <- dplyr::first(omnipath_cache_search('biocond'))

# only the versions with status "ready"
version_numbers <- omnipath_cache_filter_versions(record, status = 'ready')
omnipath_cache_remove(url = bioc_url) # cleaning up
```

omnipath_cache_get *Retrieves one item from the cache directory*

Description

Retrieves one item from the cache directory

Usage

```
omnipath_cache_get(
  key = NULL,
  url = NULL,
  post = NULL,
  payload = NULL,
  create = TRUE,
  ...
)
```

Arguments

key	The key of the cache record
url	URL pointing to the resource
post	HTTP POST parameters as a list
payload	HTTP data payload
create	Create a new entry if doesn't exist yet
...	Passed to omnipath_cache_record (internal function)

Value

Cache record: an existing record if the entry already exists, otherwise a newly created and inserted record

Examples

```
# create an example cache record
bioc_url <- 'https://bioconductor.org/'
version <- omnipath_cache_latest_or_new(url = bioc_url)
omnipath_cache_remove(url = bioc_url) # cleaning up

# retrieve the cache record
record <- omnipath_cache_get(url = bioc_url)
record$key
# [1] "41346a00fb20d2a9df03aa70cf4d50bf88ab154a"
record$url
# [1] "https://bioconductor.org/"
```

omnipath_cache_key *Generates a hash which identifies an element in the cache database*

Description

Generates a hash which identifies an element in the cache database

Usage

```
omnipath_cache_key(url, post = NULL, payload = NULL)
```

Arguments

url	Character vector with URLs
post	List with the HTTP POST parameters or a list of lists if the url vector is longer than 1. NULL for queries without POST parameters.
payload	HTTP data payload. List with multiple items if the url vector is longer than 1. NULL for queries without data.

Value

Character vector of cache record keys.

Examples

```
bioc_url <- 'https://bioconductor.org/'
omnipath_cache_key(bioc_url)
# [1] "41346a00fb20d2a9df03aa70cf4d50bf88ab154a"
```

omnipath_cache_latest_or_new *The latest or a new version of a cache record*

Description

Looks up a record in the cache and returns its latest valid version. If the record doesn't exist or no valid version available, creates a new one.

Usage

```
omnipath_cache_latest_or_new(
  key = NULL,
  url = NULL,
  post = NULL,
  payload = NULL,
  create = TRUE,
  ...
)
```

Arguments

key	The key of the cache record
url	URL pointing to the resource
post	HTTP POST parameters as a list
payload	HTTP data payload
create	Logical: whether to create and return a new version. If FALSE only the latest existing valid version is returned, if available.
...	Passed to omnipath_cache_get

Value

A cache version item.

Examples

```
## Not run:
# retrieve the latest version of the first cache record
# found by the search keyword "bioplex"
latest_bioplex <-
    omnipath_cache_latest_or_new(
        names(omnipath_cache_search('bioplex'))[1]
    )

latest_bioplex$dl_finished
# [1] "2021-03-09 14:28:50 CET"
latest_bioplex$path
# [1] "/home/denes/.cache/OmnipathR/378e0def2ac97985f629-1.rds"

## End(Not run)

# create an example cache record
bioc_url <- 'https://bioconductor.org/'
version <- omnipath_cache_latest_or_new(url = bioc_url)
omnipath_cache_remove(url = bioc_url) # cleaning up
```

omnipath_cache_latest_version *Finds the most recent version in a cache record*

Description

Finds the most recent version in a cache record

Usage

```
omnipath_cache_latest_version(record)
```

Arguments

record	A cache record

Value

Character: the version ID with the most recent download finished time

omnipath_cache_load *Loads an R object from the cache*

Description

Loads the object from RDS format.

Usage

```
omnipath_cache_load(
  key = NULL,
  version = NULL,
  url = NULL,
  post = NULL,
  payload = NULL
)
```

Arguments

key	Key of the cache item
version	Version of the cache item. If does not exist or NULL, the latest version will be retrieved
url	URL of the downloaded resource
post	HTTP POST parameters as a list
payload	HTTP data payload

Value

Object loaded from the cache RDS file.

See Also

omnipath_cache_save

Examples

```
url <- paste0(
    'https://omnipathdb.org/intercell?resources=Adhesome,Almen2009,',
    'Baccin2019,CSPA,CellChatDB&license=academic'
)
result <- read.delim(url, sep = '\t')
omnipath_cache_save(result, url = url)
# works only if you have already this item in the cache
intercell_data <- omnipath_cache_load(url = url)
class(intercell_data)
# [1] "data.frame"
nrow(intercell_data)
# [1] 16622
attr(intercell_data, 'origin')
# [1] "cache"

# basic example of saving and loading to and from the cache:
bioc_url <- 'https://bioconductor.org/'
bioc_html <- readChar(url(bioc_url), nchars = 99999)
omnipath_cache_save(bioc_html, url = bioc_url)
bioc_html <- omnipath_cache_load(url = bioc_url)
```

---

omnipath_cache_move_in

Moves an existing file into the cache

Description

Either the key or the URL (with POST and payload) must be provided.

Usage

```
omnipath_cache_move_in(
  path,
  key = NULL,
  version = NULL,
  url = NULL,
  post = NULL,
  payload = NULL,
  keep_original = FALSE
)
```

Arguments

path	Path to the source file
key	Key of the cache item
version	Version of the cache item. If does not exist a new version item will be created
url	URL of the downloaded resource
post	HTTP POST parameters as a list
payload	HTTP data payload
keep_original	Whether to keep or remove the original file

Value

Character: invisibly returns the version number of the cache version item.

See Also

omnipath_cache_save

Examples

```
path <- tempfile()
saveRDS(rnorm(100), file = path)
omnipath_cache_move_in(path, url = 'the_download_address')

# basic example of moving a file to the cache:

bioc_url <- 'https://bioconductor.org/'
html_file <- tempfile(fileext = '.html')
httr::GET(bioc_url, httr::write_disk(html_file, overwrite = TRUE))
omnipath_cache_move_in(path = html_file, url = bioc_url)
omnipath_cache_remove(url = bioc_url) # cleaning up
```

---

omnipath_cache_remove *Removes contents from the cache directory*

Description

According to the parameters, it can remove contents older than a certain age, or contents having a more recent version, one specific item, or wipe the entire cache.

Usage

```
omnipath_cache_remove(key = NULL, url = NULL, post = NULL,
    payload = NULL, max_age = NULL, min_age = NULL, status = NULL,
    only_latest = FALSE, wipe = FALSE, autoclean = TRUE)
```

Arguments

key	The key of the cache record
url	URL pointing to the resource
post	HTTP POST parameters as a list
payload	HTTP data payload
max_age	Age of cache items in days. Remove everything that is older than this age
min_age	Age of cache items in days. Remove everything more recent than this age
status	Remove items having any of the states listed here
only_latest	Keep only the latest version
wipe	Logical: if TRUE, removes all files from the cache and the cache database. Same as calling omnipath_cache_wipe.
autoclean	Remove the entries about failed downloads, the files in the cache directory which are missing from the cache database, and the entries without existing files in the cache directory

Value

Invisibly returns the cache database (list of cache records).

See Also

- omnipath_cache_wipe
- omnipath_cache_clean
- omnipath_cache_autoclean

Examples

```
## Not run:
# remove all cache data from the BioPlex database
cache_records <- omnipath_cache_search(
    'bioplex',
    ignore.case = TRUE
)
omnipath_cache_remove(names(cache_records))

# remove a record by its URL
regnetwork_url <- 'http://www.regnetworkweb.org/download/human.zip'
omnipath_cache_remove(url = regnetwork_url)

# remove all records older than 30 days
omnipath_cache_remove(max_age = 30)

# for each record, remove all versions except the latest
omnipath_cache_remove(only_latest = TRUE)

## End(Not run)
```

```
bioc_url <- 'https://bioconductor.org/'
version <- omnipath_cache_latest_or_new(url = bioc_url)
httr::GET(bioc_url, httr::write_disk(version$path, overwrite = TRUE))
omnipath_cache_download_ready(version)
key <- omnipath_cache_key(bioc_url)
omnipath_cache_remove(key = key)
```

`omnipath_cache_save` *Saves an R object to the cache*

Description

Exports the object in RDS format, creates new cache record if necessary.

Usage

```
omnipath_cache_save(
  data,
  key = NULL,
  version = NULL,
  url = NULL,
  post = NULL,
  payload = NULL
)
```

Arguments

`data`	An object
`key`	Key of the cache item
`version`	Version of the cache item. If does not exist a new version item will be created
`url`	URL of the downloaded resource
`post`	HTTP POST parameters as a list
`payload`	HTTP data payload

Value

Returns invisibly the data itself.

Invisibly returns the ‘data‘.

See Also

`omnipath_cache_move_in`

Examples

```
mydata <- data.frame(a = c(1, 2, 3), b = c('a', 'b', 'c'))
omnipath_cache_save(mydata, url = 'some_dummy_address')
from_cache <- omnipath_cache_load(url = 'some_dummy_address')
from_cache
#   a b
# 1 1 a
# 2 2 b
# 3 3 c
attr(from_cache, 'origin')
# [1] "cache"

# basic example of saving and loading to and from the cache:
bioc_url <- 'https://bioconductor.org/'
bioc_html <- readChar(url(bioc_url), nchars = 99999)
omnipath_cache_save(bioc_html, url = bioc_url)
bioc_html <- omnipath_cache_load(url = bioc_url)
```

---

omnipath_cache_search *Searches for cache items*

Description

Searches the cache records by matching the URL against a string or regexp.

Usage

```
omnipath_cache_search(pattern, ...)
```

Arguments

pattern	String or regular expression.
...	Passed to grep

Value

List of cache records matching the pattern.

Examples

```
# find all cache records from the BioPlex database
bioplex_cache_records <- omnipath_cache_search(
    'bioplex',
    ignore.case = TRUE
)
```

omnipath_cache_set_ext *Sets the file extension for a cache record*

Description

Sets the file extension for a cache record

Usage

```
omnipath_cache_set_ext(key, ext)
```

Arguments

key	Character: key for a cache item, alternatively a version entry.
ext	Character: the file extension, e.g. "zip".

Value

Returns ‘NULL‘.

Examples

```
bioc_url <- 'https://bioconductor.org/'
version <- omnipath_cache_latest_or_new(url = bioc_url)
version$path
# [1] "/home/denes/.cache/OmnipathR/41346a00fb20d2a9df03-1"
httr::GET(bioc_url, httr::write_disk(version$path, overwrite = TRUE))
key <- omnipath_cache_key(url = bioc_url)
omnipath_cache_set_ext(key = key, ext = 'html')
version <- omnipath_cache_latest_or_new(url = bioc_url)
version$path
# [1] "/home/denes/.cache/OmnipathR/41346a00fb20d2a9df03-1.html"
record <- omnipath_cache_get(url = bioc_url)
record$ext
# [1] "html"
omnipath_cache_remove(url = bioc_url) # cleaning up
```

omnipath_cache_update_status
Updates the status of an existing cache record

Description

Updates the status of an existing cache record

Usage

```
omnipath_cache_update_status(key, version, status,
    dl_finished = NULL)
```

Arguments

key	Key of the cache item
version	Version of the cache item. If does not exist a new version item will be created
status	The updated status value
dl_finished	Timestamp for the time when download was finished, if 'NULL' the value remains unchanged

Value

Character: invisibly returns the version number of the cache version item.

Examples

```
bioc_url <- 'https://bioconductor.org/'
latest_version <- omnipath_cache_latest_or_new(url = bioc_url)
key <- omnipath_cache_key(bioc_url)
omnipath_cache_update_status(
    key = key,
    version = latest_version$number,
    status = 'ready',
    dl_finished = Sys.time()
)
omnipath_cache_remove(url = bioc_url) # cleaning up
```

omnipath_cache_wipe *Permanently removes all the cache contents*

Description

After this operation the cache directory will be completely empty, except an empty cache database file.

Usage

```
omnipath_cache_wipe(...)
```

Arguments

...	Ignored.

Value

Returns 'NULL'.

See Also

omnipath_cache_remove

Examples

```
## Not run:
omnipath_cache_wipe()
# the cache is completely empty:
print(omnipath.env$cache)
# list()
list.files(omnipath_get_cachedir())
# [1] "cache.json"

## End(Not run)
```

omnipath_get_config_path *Current config file path*

Description

Current config file path

Usage

```
omnipath_get_config_path(user = FALSE)
```

Arguments

user	Logical: prioritize the user level config even if a config in the current working directory is available.

Value

Character: path to the config file.

Examples

```
omnipath_get_config_path()
```

---

omnipath_load_config *Load the package configuration from a config file*

Description

Load the package configuration from a config file

Usage

```
omnipath_load_config(path = NULL, title = "default", user = FALSE, ...)
```

Arguments

path	Path to the config file.
title	Load the config under this title. One config file might contain multple configurations, each identified by a title. If the title is not available the first section of the config file will be used.
user	Force to use the user level config even if a config file exists in the current directory. By default, the local config files have prioroty over the user level config.
...	Passed to `yaml::yaml.load_file`.

Value

Invisibly returns the config as a list.

Examples

```
## Not run:
# load the config from a custom config file:
omnipath_load_config(path = 'my_custom_omnipath_config.yml')

## End(Not run)
```

`omnipath_log` *Browse the current OmnipathR log file*

Description

Browse the current OmnipathR log file

Usage

```
omnipath_log()
```

Value

Returns 'NULL'.

See Also

`omnipath_logfile`

Examples

```
## Not run:
omnipath_log()
# then you can browse the log file, and exit with `q`

## End(Not run)
```

`omnipath_logfile` *Path to the current OmnipathR log file*

Description

Path to the current OmnipathR log file

Usage

```
omnipath_logfile()
```

Value

Character: path to the current logfile, or NULL if no logfile is available.

See Also

omnipath_log

Examples

```
omnipath_logfile()
# [1] "/home/denes/omnipathr/omnipathr-log/omnipathr-20210309-1642.log"
```

---

omnipath_msg *Dispatch a message to the OmnipathR logger*

Description

Any package or script can easily send log messages and establish a logging facility with the fantastic 'logger' package. This function serves the only purpose if you want to inject messages into the logger of OmnipathR. Otherwise we recommend to use the 'logger' package directly.

Usage

```
omnipath_msg(level, ...)
```

Arguments

level	Character, numeric or class loglevel. A log level, if character one of the followings: "fatal", "error", "warn", "success", "info", "trace".
...	Arguments for string formatting, passed sprintf or str_glue.

Value

Returns 'NULL'.

Examples

```
omnipath_msg(
    level = 'success',
    'Talking to you in the name of OmnipathR, my favourite number is %d',
    round(runif(1, 1, 10))
)
```

omnipath_reset_config *Restores the built-in default values of all config parameters*

Description

Restores the built-in default values of all config parameters

Usage

```
omnipath_reset_config(save = NULL, reset_all = FALSE)
```

Arguments

save	If a path, the restored config will be also saved to this file. If TRUE, the config will be saved to the current default config path (see omnipath_get_config_path).
reset_all	Reset to their defaults also the options already set in the R options.

Value

The config as a list.

See Also

omnipath_load_config, omnipath_save_config

Examples

```
## Not run:
# restore the defaults and write them to the default config file:
omnipath_reset_config()
omnipath_save_config()

## End(Not run)
```

omnipath_save_config *Save the current package configuration*

Description

Save the current package configuration

Usage

```
omnipath_save_config(path = NULL, title = "default", local = FALSE)
```

Arguments

path	Path to the config file. Directories and the file will be created if don't exist.
title	Save the config under this title. One config file might contain multiple configurations, each identified by a title.
local	Save into a config file in the current directory instead of a user level config file. When loading, the config in the current directory has priority over the user level config.

Value

Returns 'NULL'.

Examples

```
## Not run:
# after this, all downloads will default to commercial licenses
# i.e. the resources that allow only academic use will be excluded:
options(omnipath.license = 'commercial')
omnipath_save_config()

## End(Not run)
```

---

omnipath_set_cachedir *Change the cache directory*

Description

Change the cache directory

Usage

```
omnipath_set_cachedir(path = NULL)
```

Arguments

path	Character: path to the new cache directory. If don't exist, the directories will be created. If the path is an existing cache directory, the package's cache database for the current session will be loaded from the database in the directory. If NULL, the cache directory will be set to its default path.

Value

Returns NULL.

Examples

```
tmp_cache <- tempdir()
omnipath_set_cachedir(tmp_cache)
# restore the default cache directory:
omnipath_set_cachedir()
```

---

omnipath_set_console_loglevel

Sets the log level for the console

Description

Use this method to change during a session which messages you want to be printed on the console. Before loading the package, you can set it also by the config file, with the omnipath.console_loglevel key.

Usage

```
omnipath_set_console_loglevel(level)
```

Arguments

`level`	Character or class ‘loglevel‘. The desired log level.

Value

Returns ‘NULL‘.

See Also

`omnipath_set_logfile_loglevel`

Examples

```
omnipath_set_console_loglevel('warn')
# or:
omnipath_set_console_loglevel(logger::WARN)
```

omnipath_set_logfile_loglevel *Sets the log level for the logfile*

Description

Use this method to change during a session which messages you want to be written into the logfile. Before loading the package, you can set it also by the config file, with the omnipath.loglevel key.

Usage

```
omnipath_set_logfile_loglevel(level)
```

Arguments

level	Character or class 'loglevel'. The desired log level.

Value

Returns 'NULL'.

See Also

omnipath_set_console_loglevel

Examples

```
omnipath_set_logfile_loglevel('info')
# or:
omnipath_set_logfile_loglevel(logger::INFO)
```

omnipath_set_loglevel *Sets the log level for the package logger*

Description

Sets the log level for the package logger

Usage

```
omnipath_set_loglevel(level, target = "logfile")
```

Arguments

level	Character or class 'loglevel'. The desired log level.
target	Character, either 'logfile' or 'console'

Value

Returns 'NULL'.

Examples

```
omnipath_set_loglevel(logger::FATAL, target = 'console')
```

`omnipath_show_db` *Built in database definitions*

Description

Databases are resources which might be costly to load but can be used many times by functions which usually automatically load and retrieve them from the database manager. Each database has a lifetime and will be unloaded automatically upon expiry.

Usage

```
omnipath_show_db()
```

Value

A data frame with the built in database definitions.

Examples

```
database_definitions <- omnipath_show_db()
database_definitions
# # A tibble: 14 x 10
#    name       last_used           lifetime package  loader    loader_p.
#    <chr>      <dttm>                 <dbl> <chr>    <chr>     <list>
#  1 Gene Onto. 2021-04-04 20:19:15      300 Omnipat. go_ontol. <named l.
#  2 Gene Onto. NA                       300 Omnipat. go_ontol. <named l.
#  3 Gene Onto. NA                       300 Omnipat. go_ontol. <named l.
#  4 Gene Onto. NA                       300 Omnipat. go_ontol. <named l.
#  5 Gene Onto. NA                       300 Omnipat. go_ontol. <named l.
# ... (truncated)
# # . with 4 more variables: latest_param <list>, loaded <lgl>, db <list>,
# #   key <chr>
```

omnipath_unlock_cache_db

Removes the lock file from the cache directory

Description

A lock file in the cache directory avoids simulatneous write and read. It's supposed to be removed after each read and write operation. This might not happen if the process crashes during such an operation. In this case you can manually call this function.

Usage

```
omnipath_unlock_cache_db()
```

Value

Logical: returns TRUE if the cache was locked and now is unlocked; FALSE if it was not locked.

Examples

```
omnipath_unlock_cache_db()
```

ontology_ensure_id *Only ontology IDs*

Description

Converts a mixture of ontology IDs and names to only IDs. If an element of the input is missing from the chosen ontology it will be dropped. This can happen if the ontology is a subset (slim) version, but also if the input is not a valid ID or name.

Usage

```
ontology_ensure_id(terms, db_key = "go_basic")
```

Arguments

terms	Character: ontology IDs or term names.
db_key	Character: key to identify the ontology database. For the available keys see omnipath_show_db.

Value

Character vector of ontology IDs.

Examples

```
ontology_ensure_id(c('mitochondrion inheritance', 'GO:0001754'))
# [1] "GO:0000001" "GO:0001754"
```

---

ontology_ensure_name *Only ontology term names*

Description

Converts a mixture of ontology IDs and names to only names. If an element of the input is missing from the chosen ontology it will be dropped. This can happen if the ontology is a subset (slim) version, but also if the input is not a valid ID or name.

Usage

```
ontology_ensure_name(terms, db_key = "go_basic")
```

Arguments

terms	Character: ontology IDs or term names.
db_key	Character: key to identify the ontology database. For the available keys see omnipath_show_db.

Value

Character vector of ontology term names.

Examples

```
ontology_ensure_name(c('reproduction', 'GO:0001754', 'foo bar'))
# [1] "eye photoreceptor cell differentiation" "reproduction"
```

---

ontology_name_id *Translate between ontology IDs and names*

Description

Makes sure that the output contains only valid IDs or term names. The input can be a mixture of IDs and names. The order of the input won't be preserved in the output.

Usage

```
ontology_name_id(terms, ids = TRUE, db_key = "go_basic")
```

Arguments

terms	Character: ontology IDs or term names.
ids	Logical: the output should contain IDs or term names.
db_key	Character: key to identify the ontology database. For the available keys see omnipath_show_db.

Value

Character vector of ontology IDs or term names.

Examples

```
ontology_name_id(c('mitochondrion inheritance', 'reproduction'))
# [1] "GO:0000001" "GO:0000003"
ontology_name_id(c('GO:0000001', 'reproduction'), ids = FALSE)
# [1] "mitochondrion inheritance" "reproduction"
```

---

pathwaycommons_download

Interactions from PathwayCommons

Description

PathwayCommons (http://www.pathwaycommons.org/) provides molecular interactions from a number of databases, in either BioPAX or SIF (simple interaction format). This function retrieves all interactions in SIF format. The data is limited to the interacting pair and the type of the interaction.

Usage

```
pathwaycommons_download()
```

Value

A data frame (tibble) with interactions.

Examples

```
pc_interactions <- pathwaycommons_download()
 pc_interactions
# # A tibble: 1,884,849 x 3
#    from  type                       to
#    <chr> <chr>                      <chr>
#  1 A1BG  controls-expression-of     A2M
#  2 A1BG  interacts-with             ABCC6
#  3 A1BG  interacts-with             ACE2
#  4 A1BG  interacts-with             ADAM10
#  5 A1BG  interacts-with             ADAM17
```

```
# # . with 1,884,839 more rows
```

`pivot_annotations` *Converts annotation tables to a wide format*

Description

Use this method to reconstitute the annotation tables into the format of the original resources. With the 'wide=TRUE' option `import_omnipath_annotations` applies this function to the downloaded data.

Usage

```
pivot_annotations(annotations)
```

Arguments

`annotations`	A data frame of annotations downloaded from the OmniPath web service by `import_omnipath_annotations`.

Value

A wide format data frame (tibble) if the provided data contains annotations from one resource, otherwise a list of wide format tibbles.

See Also

`import_omnipath_annotations`

Examples

```
# single resource: the result is a data frame
disgenet <- import_omnipath_annotations(resources = 'DisGeNet')
disgenet <- pivot_annotations(disgenet)
disgenet
# # A tibble: 126,588 × 11
#    uniprot genesymbol entity_type disease      type  score   dsi   dpi
#    <chr>   <chr>      <chr>       <chr>        <chr> <dbl> <dbl> <dbl>
#  1 P04217  A1BG       protein     Schizophren. dise.  0.3  0.7   0.538
#  2 P04217  A1BG       protein     Hepatomegaly phen.  0.3  0.7   0.538
#  3 P01023  A2M        protein     Fibrosis, L. dise.  0.3  0.529 0.769
#  4 P01023  A2M        protein     Acute kidne. dise.  0.3  0.529 0.769
#  5 P01023  A2M        protein     Mental Depr. dise.  0.3  0.529 0.769
# # . with 126,583 more rows, and 3 more variables: nof_pmids <dbl>,
# #   nof_snps <dbl>, source <chr>

# multiple resources: the result is a list
annotations <- import_omnipath_annotations(
```

```
    resources = c('DisGeNet', 'SignaLink_function', 'DGIdb', 'kinase.com')
)
annotations <- pivot_annotations(annotations)
names(annotations)
# [1] "DGIdb"              "DisGeNet"           "kinase.com"
# [4] "SignaLink_function"
annotations$kinase.com
# # A tibble: 825 x 6
#    uniprot genesymbol entity_type group family subfamily
#    <chr>   <chr>      <chr>       <chr> <chr>  <chr>
#  1 P31749  AKT1       protein     AGC   Akt    NA
#  2 P31751  AKT2       protein     AGC   Akt    NA
#  3 Q9Y243  AKT3       protein     AGC   Akt    NA
#  4 O14578  CIT        protein     AGC   DMPK   CRIK
#  5 Q09013  DMPK       protein     AGC   DMPK   GEK
# # . with 815 more rows
```

`preppi_download` *Interactions from PrePPI*

Description

Retrieves predicted protein-protein interactions from the PrePPI database (http://honig.c2b2.columbia.edu/preppi). The interactions in this table are supposed to be correct with a > 0.5 probability.

Usage

```
preppi_download(...)
```

Arguments

`...`	Minimum values for the scores. The available scores are: str, protpep, str_max, red, ort, phy, coexp, go, total, exp and final. Furthermore, an operator can be passed, either `.op = '&'` or `.op = '|'`, which is then used for combined filtering by multiple scores.

Details

PrePPI is a combination of many prediction methods, each resulting a score. For an explanation of the scores see https://honiglab.c2b2.columbia.edu/hfpd/help/Manual.html. The minimum, median and maximum values of the scores:

Score	Minimum	Median	Maximum
str	0	5.5	6,495
protpep	0	3.53	38,138
str_max	0	17.9	38,138

```
| red     |       0 |     1.25 |              24.4  |
| ort     |       0 |     0    |           5,000    |
| phy     |       0 |     2.42 |              2.42 |
| coexp   |       0 |     2.77 |             45.3  |
| go      |       0 |     5.86 |             181    |
| total   |       0 | 1,292    | 106,197,000,000    |
| exp     |       1 |   958    |           4,626    |
| final   |     600 | 1,778    |            4.91e14 |
```

Value

A data frame (tibble) of interactions with scores, databases and literature references.

See Also

preppi_filter

Examples

```
preppi <- preppi_download()
preppi
# # A tibble: 1,545,710 x 15
#    prot1 prot2 str_score protpep_score str_max_score red_score ort_score
#    <chr> <chr>     <dbl>         <dbl>         <dbl>     <dbl>     <dbl>
#  1 Q131. P146.     18.6           6.45         18.6      4.25      0.615
#  2 P064. Q96N.      1.83         14.3          14.3      4.25      0
#  3 Q7Z6. Q8NC.      4.57          0             4.57     0         0
#  4 P370. P154.    485.            0           485.       1.77      0.615
#  5 O004. Q9NR.     34.0           0            34.0      0.512     0
# # . with 1,545,700 more rows, and 8 more variables: phy_score <dbl>,
# #   coexp_score <dbl>, go_score <dbl>, total_score <dbl>, dbs <chr>,
# #   pubs <chr>, exp_score <dbl>, final_score <dbl>
```

`preppi_filter` *Filter PrePPI interactions by scores*

Description

Filter PrePPI interactions by scores

Usage

```
preppi_filter(data, ..., .op = "&")
```

Arguments

data	A data frame of PrePPI interactions as provided by preppi_download.
...	Minimum values for the scores. The available scores are: str, protpep, str_max, red, ort, phy, coexp, go, total, exp and final. See more about the scores at preppi_download.
.op	The operator to combine the scores with: either '&' or '|'. With the former, only records where all scores are above the threshold will be kept; with the latter, records where at least one score is above its threshold will be kept.

Value

The input data frame (tibble) filtered by the score thresholds.

See Also

preppi_download

Examples

```
preppi <- preppi_download()
preppi_filtered <- preppi_filter(preppi, red = 10, str = 4.5, ort = 1)
nrow(preppi_filtered)
# [1] 8443
```

---

print_bma_motif_es *Prints BMA motifs to the screen from a sequence of edges*

Description

The motifs can be copy-pasted into a BMA canvas.

Usage

```
print_bma_motif_es(edge_seq, G, granularity = 2)
```

Arguments

edge_seq	An igraph edge sequence.
G	An igraph graph object.
granularity	Numeric: granularity value.

Value

Returns ‘NULL‘.

Examples

```
interactions <- import_omnipath_interactions(resources = 'ARN')
graph <- interaction_graph(interactions)
print_bma_motif_es(igraph::E(graph)[1], graph)
# {"Model": {
#     "Name": "Omnipath motif",
#     "Variables":[{
#         "Name":"ULK1",
#         "Id":1,
#         "RangeFrom":0,
#         "RangeTo":2,
#         "Formula":""
#     },
#     {
#         "Name":"ATG13",
#         ...
#     }],
# ... (truncated)
# }}
```

`print_bma_motif_vs` *Prints BMA motifs to the screen from a sequence of nodes*

Description

The motifs can be copy-pasted into a BMA canvas.

Usage

```
print_bma_motif_vs(node_seq, G)
```

Arguments

`node_seq`	An igraph node sequence.
`G`	An igraph graph object.

Value

Returns 'NULL'.

Examples

```
interactions <- import_omnipath_interactions(resources = 'ARN')
graph <- interaction_graph(interactions)
print_bma_motif_vs(
    igraph::all_shortest_paths(
        graph,
        from = 'ULK1',
```

```
        to = 'ATG13'
    )$res,
    graph
)
```

---

print_interactions *Print OmniPath interactions*

Description

Prints the interactions or enzyme-substrate relationships in a nice format.

Usage

```
print_interactions(interDF, writeRefs = FALSE)
```

Arguments

interDF	data.frame with the interactions generated by any of the following functions: • import_omnipath_enzsub • import_omnipath_interactions • import_pathwayextra_interactions • import_kinaseextra_interactions • import_ligrecextra_interactions • import_post_translational_interactions • import_dorothea_interactions • import_tf_target_interactions • import_transcriptional_interactions • import_mirnatarget_interactions • import_all_interactions
writeRefs	[FALSE] writes also the PubMed IDs if available

Value

Returns ‘NULL‘.

Examples

```
enzsub <- import_omnipath_enzsub()
print_interactions(head(enzsub))
print_interactions(tail(enzsub), writeRefs = TRUE)
print_interactions(
    dplyr::filter(
        enzsub,
        enzyme_genesymbol == 'MAP2K1',
```

```
        substrate_genesymbol == 'MAPK3'
    )
)

signor <- import_omnipath_interactions(resources = 'SIGNOR')
print_interactions(head(signor))
#            source interaction            target n_resources
# 6 MAPK14 (Q16539)  ==( + )==> MAPKAPK2 (P49137)          23
# 4  TRPM7 (Q96QT4)  ==( + )==>    ANXA1 (P04083)          10
# 1  PRKG1 (Q13976)  ==( - )==>    TRPC3 (Q13507)           8
# 2  PTPN1 (P18031)  ==( - )==>    TRPV6 (Q9H1D0)           6
# 5 PRKACA (P17612)  ==( - )==>   MCOLN1 (Q9GZU1)           6
# 3  RACK1 (P63244)  ==( - )==>    TRPM6 (Q9BX84)           2
```

`print_path_es` *Prints network paths in an edge sequence*

Description

Pretty prints the interactions in a path.

Usage

```
print_path_es(edgeSeq, G)
```

Arguments

edgeSeq	edge sequence
G	igraph object (from ptms or any interaction dataset)

Value

Returns ‘NULL‘.

See Also

- print_path_vs

Examples

```
interactions <- import_omnipath_interactions(resources = c('SignaLink3'))
OPI_g <- interaction_graph(interactions = interactions)
print_path_es(
    suppressWarnings(igraph::shortest_paths(
        OPI_g,
        from = 'TYRO3',
        to = 'STAT3',
        output = 'epath'
    ))$epath[[1]],
```

```
    OPI_g
)
```

---

print_path_vs *Print networks paths given by node sequence*

Description

Prints the interactions in the path in a nice format.

Usage

```
print_path_vs(nodeSeq, G)
```

Arguments

nodeSeq	node sequence
G	igraph object (from ptms or interactions)

Value

Returns ‘NULL‘.

See Also

print_path_es

Examples

```
interactions <- import_omnipath_interactions(resources=c('SignaLink3'))
OPI_g <- interaction_graph(interactions = interactions)
print_path_vs(
    igraph::all_shortest_paths(
        OPI_g,
        from = 'TYRO3',
        to = 'STAT3'
    )$vpath,
    OPI_g
)
enzsub <- import_omnipath_enzsub(resources=c('PhosphoSite', 'SIGNOR'))
enzsub_g <- enzsub_graph(enzsub)
print_path_vs(
    igraph::all_shortest_paths(
        enzsub_g,
        from = 'SRC',
        to = 'STAT1'
    )$res,
    enzsub_g
)
```

pubmed_open *Open one or more PubMed articles*

Description

Open one or more PubMed articles

Usage

```
pubmed_open(pmids, browser = NULL, sep = ";", max_pages = 25L)
```

Arguments

pmids	Character or numberic vector of one or more PubMed IDs.
browser	Character: name of the web browser executable. If 'NULL', the default web browser will be used.
sep	Character: split the PubMed IDs by this separator.
max_pages	Numeric: largest number of pages to open. This is to prevent opening hundreds or thousands of pages at once.

Value

Returns 'NULL'.

Examples

```
interactions <- import_omnipath_interactions()
pubmed_open(interactions$references[1])
```

ramilowski_download *Downloads ligand-receptor interactions from Ramilowski et al. 2015*

Description

Curated ligand-receptor pairs from Supplementary Table 2 of the article "A draft network of ligand-receptor mediated multicellular signaling in human" (https://www.nature.com/articles/ncomms8866).

Usage

```
ramilowski_download()
```

Value

A data frame (tibble) with interactions.

Examples

```
rami_interactions <- ramilowski_download()
rami_interactions
# # A tibble: 2,557 x 16
#    Pair.Name Ligand.Approved. Ligand.Name Receptor.Approv.
#    <chr>     <chr>            <chr>       <chr>
#  1 A2M_LRP1  A2M              alpha-2-ma. LRP1
#  2 AANAT_MT. AANAT            aralkylami. MTNR1A
#  3 AANAT_MT. AANAT            aralkylami. MTNR1B
#  4 ACE_AGTR2 ACE              angiotensi. AGTR2
#  5 ACE_BDKR. ACE              angiotensi. BDKRB2
# # . with 2,547 more rows, and 12 more variables: Receptor.Name <chr>,
# #   DLRP <chr>, HPMR <chr>, IUPHAR <chr>, HPRD <chr>,
# #   STRING.binding <chr>, STRING.experiment <chr>, HPMR.Ligand <chr>,
# #   HPMR.Receptor <chr>, PMID.Manual <chr>, Pair.Source <chr>,
# #   Pair.Evidence <chr>
```

---

regnetwork_directions *Transcription factor effects from RegNetwork*

Description

Transcription factor effects from RegNetwork

Usage

```
regnetwork_directions(organism = "human")
```

Arguments

organism	Character: either human or mouse.

Value

A data frame (tibble) of TF-target interactions with effect signs.

Examples

```
regn_dir <- regnetwork_directions()
regn_dir
# # A tibble: 3,954 x 5
#    source_genesymb. source_entrez target_genesymb. target_entrez
#    <chr>            <chr>         <chr>            <chr>
#  1 AHR              196           CDKN1B           1027
#  2 APLNR            187           PIK3C3           5289
#  3 APLNR            187           PIK3R4           30849
#  4 AR               367           KLK3             354
#  5 ARNT             405           ALDOA            226
```

```
# # . with 3,944 more rows, and 1 more variable: effect <dbl>
```

`regnetwork_download` *Interactions from RegNetwork*

Description

Downloads transcriptional and post-transcriptional regulatory interactions from the RegNetwork database (http://www.regnetworkweb.org/). The information about effect signs (stimulation or inhibition), provided by `regnetwork_directions` are included in the result.

Usage

```
regnetwork_download(organism = "human")
```

Arguments

`organism`	Character: either human or mouse.

Value

Data frame with interactions.

Examples

```
regn_interactions <- regnetwork_download()
regn_interactions
# # A tibble: 372,778 x 7
#    source_genesymb. source_entrez target_genesymb. target_entrez
#    <chr>            <chr>         <chr>            <chr>
#  1 USF1             7391          S100A6           6277
#  2 USF1             7391          DUSP1            1843
#  3 USF1             7391          C4A              720
#  4 USF1             7391          ABCA1            19
#  5 TP53             7157          TP73             7161
# # . with 372,768 more rows, and 3 more variables: effect <dbl>,
# #   source_type <chr>, target_type <chr>
```

relations_list_to_table *Table from a nested list of ontology relations*

Description

Converting the nested list to a table is a more costly operation, it takes a few seconds. Best to do it only once, or pass `tables = TRUE` to `obo_parser`, and convert the data frame to list, if you also need it in list format.

Usage

```
relations_list_to_table(relations, direction = NULL)
```

Arguments

`relations`	A nested list of ontology relations (the "relations" element of the list returned by `obo_parser` in case its argument 'tables' is `FALSE`).
`direction`	Override the direction (i.e. child -> parents or parent -> children). The nested lists produced by functions in the current package add an attribute "direction" thus no need to pass this value. If the attribute and the argument are both missing, the column will be named simply "side2" and it won't be clear whether the relations point from "term" to "side2" or the other way around. The direction should be a character vector of length 2 with the values "parents" and "children".

Value

The relations converted to a data frame (tibble).

See Also

- `swap_relations`
- `relations_table_to_list`
- `obo_parser`

Examples

```
goslim_url <-
    "http://current.geneontology.org/ontology/subsets/goslim_generic.obo"
path <- tempfile()
httr::GET(goslim_url, httr::write_disk(path, overwrite = TRUE))
obo <- obo_parser(path, tables = FALSE)
unlink(path)
rel_tbl <- relations_list_to_table(obo$relations)
```

relations_table_to_graph

Graph from a table of ontology relations

Description

Graph from a table of ontology relations

Usage

```
relations_table_to_graph(relations)
```

Arguments

relations	A data frame of ontology relations (the "relations" element of the list returned by obo_parser in case its argument ‘tables‘ is TRUE).

Details

By default the relations point from child to parents, the edges in the graph will be of the same direction. Use swap_relations on the data frame to reverse the direction.

Value

The relations converted to an igraph graph object.

Examples

```
go <- get_db('go_basic')
go_graph <- relations_table_to_graph(go$relations)
```

relations_table_to_list

Nested list from a table of ontology relations

Description

Nested list from a table of ontology relations

Usage

```
relations_table_to_list(relations)
```

Arguments

relations	A data frame of ontology relations (the "relations" element of the list returned by obo_parser in case its argument ‘tables‘ is TRUE).

Value

The relations converted to a nested list.

See Also

- relations_list_to_table
- swap_relations
- obo_parser

Examples

```
goslim_url <-
    "http://current.geneontology.org/ontology/subsets/goslim_generic.obo"
path <- tempfile()
httr::GET(goslim_url, httr::write_disk(path, overwrite = TRUE))
obo <- obo_parser(path, tables = TRUE)
unlink(path)
rel_list <- relations_table_to_list(obo$relations)
```

---

remap_dorothea_download

Downloads TF-target interactions from ReMap

Description

ReMap (http://remap.univ-amu.fr/) is a database of ChIP-Seq experiments. It provides raw and merged peaks and CRMs (cis regulatory motifs) with their associations to regulators (TFs). TF-target relationships can be derived as it is written in Garcia-Alonso et al. 2019: "For ChIP-seq, we downloaded the binding peaks from ReMap and scored the interactions between each TF and each gene according to the distance between the TFBSs and the genes' transcription start sites. We evaluated different filtering strategies that consisted of selecting only the top-scoring 100, 200, 500, and 1000 target genes for each TF." (https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6673718/#s1title). This function returns the top TF-target relationships as used in DoRothEA: https://github.com/saezlab/dorothea/blob/master/inst/scripts/02_chip_seq.R).

Usage

```
remap_dorothea_download()
```

Value

Data frame with TF-target relationships.

See Also

remap_tf_target_download

Examples

```
remap_interactions <- remap_dorothea_download()
remap_interactions
# # A tibble: 136,988 x 2
#    tf    target
#    <chr> <chr>
#  1 ADNP  ABCC1
#  2 ADNP  ABCC6
#  3 ADNP  ABHD5
#  4 ADNP  ABT1
#  5 ADNP  AC002066.1
# # . with 136,978 more rows
```

---

remap_filtered *Downloads TF-target interactions from ReMap*

Description

Downloads the ReMap TF-target interactions as processed by Garcia-Alonso et al. (https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6673718/#s1title) and filters them based on a score threshold, the top targets and whether the TF is included in the TF census (Vaquerizas et al. 2009). The code for filtering is adapted from DoRothEA, written by Christian Holland.

Usage

```
remap_filtered(score = 100, top_targets = 500, only_known_tfs = TRUE)
```

Arguments

score	Numeric: a minimum score between 0 and 1000, records with lower scores will be excluded. If NULL no filtering performed.
top_targets	Numeric: the number of top scoring targets for each TF. Essentially the maximum number of targets per TF. If NULL the number of targets is not restricted.
only_known_tfs	Logical: whether to exclude TFs which are not in TF census.

Value

Data frame with TF-target relationships.

See Also

- remap_tf_target_download
- remap_filtered
- tfcensus_download

Examples

```
## Not run:
remap_interactions <- remap_filtered()
nrow(remap_interactions)
# [1] 145680

remap_interactions <- remap_filtered(top_targets = 100)
remap_interactions
# # A tibble: 30,330 x 2
#    source_genesymbol target_genesymbol
#    <chr>             <chr>
#  1 ADNP              ABCC1
#  2 ADNP              ABT1
#  3 ADNP              AC006076.1
#  4 ADNP              AC007792.1
#  5 ADNP              AC011288.2
# # . with 30,320 more rows

## End(Not run)
```

`remap_tf_target_download`

Downloads TF-target interactions from ReMap

Description

ReMap (http://remap.univ-amu.fr/) is a database of ChIP-Seq experiments. It provides raw and merged peaks and CRMs (cis regulatory motifs) with their associations to regulators (TFs). TF-target relationships can be derived as it is written in Garcia-Alonso et al. 2019: "For ChIP-seq, we downloaded the binding peaks from ReMap and scored the interactions between each TF and each gene according to the distance between the TFBSs and the genes' transcription start sites. We evaluated different filtering strategies that consisted of selecting only the top-scoring 100, 200, 500, and 1000 target genes for each TF." (https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6673718/#s1title). This function retrieves the full processed TF-target list from the data deposited in https://zenodo.org/record/3713238.

Usage

```
remap_tf_target_download()
```

Value

Data frame with TF-target relationships.

See Also

- remap_dorothea_download
- remap_filtered

Examples

```
## Not run:
remap_interactions <- remap_tf_target_download()
remap_interactions
# # A tibble: 9,546,470 x 4
#    source_genesymbol target_genesymbol target_ensembl     score
#    <chr>             <chr>             <chr>              <dbl>
#  1 ADNP              PTPRS             ENSG00000105426.16  1000
#  2 AFF4              PRKCH             ENSG00000027075.14  1000
#  3 AHR               CTNND2            ENSG00000169862.18  1000
#  4 AR                PDE4D             ENSG00000113448.18  1000
#  5 ARID1A            PLEC              ENSG00000178209.14  1000
# # . with 9,546,460 more rows

## End(Not run)
```

---

resources_colname *Name of the column with the resources*

Description

Unfortunately the column title is different across the various query types in the OmniPath web service, so we need to guess.

Usage

```
resources_colname(data)
```

Arguments

data	A data frame downloaded by any import_... function in the current package.

Value

Character: the name of the column, if any of the column names matches.

Examples

```
co <- import_omnipath_complexes()
resources_colname(co)
# [1] "sources"
```

simplify_intercell_network *Simplify an intercell network*

Description

The intercellular communication network data frames, created by import_intercell_network, are combinations of a network data frame with two copies of the intercell annotation data frames, all of them already having quite some columns. Here we keep only the names of the interacting pair, their intercellular communication roles, and the minimal information of the origin of both the interaction and the annotations. Optionally further columns can be selected.

Usage

```
simplify_intercell_network(network, ...)
```

Arguments

network	An intercell network data frame, as provided by import_intercell_network.
...	Optional, further columns to select.

Value

An intercell network data frame with some columns removed.

See Also

- import_intercell_network
- unique_intercell_network
- filter_intercell_network

Examples

```
icn <- import_intercell_network()
icn_s <- simplify_intercell_network(icn)
```

swap_relations *Reverse the direction of ontology relations*

Description

Reverse the direction of ontology relations

Usage

```
swap_relations(relations)
```

Arguments

relations	The 'relations' component of the data returned by obo_parser or any '...ontology_download' function such as go_ontology_download. Depending on the tables argument of those functions the 'relations' can be a data frame or a nested list.

Value

Same type as the input, but the relations swapped: if in the input these pointed from each child to the parents, in the output they point from each parent to their children, and vice versa.

See Also

- relations_list_to_table
- relations_table_to_list
- obo_parser

Examples

```
goslim_url <-
    "http://current.geneontology.org/ontology/subsets/goslim_generic.obo"
path <- tempfile()
httr::GET(goslim_url, httr::write_disk(path, overwrite = TRUE))
obo <- obo_parser(path)
unlink(path)
rel_swapped <- swap_relations(obo$relations)
```

swissprots_only *Retain only SwissProt IDs*

Description

Retain only SwissProt IDs

Usage

```
swissprots_only(uniprots, organism = 9606)
```

Arguments

uniprots	Character vector of UniProt IDs.
organism	Character or integer: name or identifier of the organism.

Value

Character vector with only SwissProt IDs.

Examples

```
swissprots_only(c("Q05BL1", "A0A654IBU3", "P00533"))
# [1] "P00533"
```

tfcensus_download *Downloads the list of transcription factors from TF census*

Description

Vaquerizas et al. published in 2009 a list of transcription factors. This function retrieves Supplementary Table 2 from the article (http://www.nature.com/nrg/journal/v10/n4/index.html).

Usage

```
tfcensus_download()
```

Value

A data frame (tibble) listing transcription factors.

Examples

```
tfcensus <- tfcensus_download()
tfcensus
# # A tibble: 1,987 x 7
#    Class `Ensembl ID` `IPI ID` `Interpro DBD` `Interpro DNA-b.
#    <chr> <chr>        <chr>    <chr>          <chr>
#  1 a     ENSG0000000. IPI0021. NA             IPR001289
#  2 a     ENSG0000000. IPI0004. IPR000047;IPR. NA
#  3 a     ENSG0000000. IPI0001. IPR001356;IPR. NA
#  4 a     ENSG0000000. IPI0029. IPR000910;IPR. NA
#  5 a     ENSG0000000. IPI0001. IPR007087;IPR. IPR006794
# # . with 1,977 more rows, and 2 more variables: `HGNC symbol` <chr>,
# # `Tissue-specificity` <chr>
```

`translate_ids` *Translate gene and protein identifiers*

Description

Translates a vector of identifiers, resulting a new vector, or a column of identifiers in a data frame by creating another column with the target identifiers.

Usage

```
translate_ids(
  d,
  ...,
  uploadlists = FALSE,
  ensembl = FALSE,
  keep_untranslated = TRUE,
  return_df = FALSE,
  organism = 9606,
  reviewed = TRUE
)
```

Arguments

`d`	Character vector or data frame.
`...`	At least two arguments, with or without names. The first of these arguments describes the source identifier, the rest of them describe the target identifier(s). The values of all these arguments must be valid identifier types as shown in Details. The names of the arguments are column names. In case of the first (source) ID the column must exist. For the rest of the IDs new columns will be created with the desired names. For ID types provided as arguments without names, the name of the ID type will be used for column name.

`uploadlists` Force using the `uploadlists` service from UniProt. By default the plain query interface is used (implemented in `uniprot_full_id_mapping_table` in this package). If any of the provided ID types is only available in the uploadlists service, it will be automatically selected. The plain query interface is preferred because in the long term, with caching, it requires less download and data storage.

`ensembl` Logical: use data from Ensembl BioMart instead of UniProt.

`keep_untranslated` In case the output is a data frame, keep the records where the source identifier could not be translated. At these records the target identifier will be NA.

`return_df` Return a data frame even if the input is a vector.

`organism` Integer, NCBI Taxonomy ID of the organism (by default 9606 for human). Matters only if `uploadlists` is `FALSE`.

`reviewed` Translate only reviewed (`TRUE`), only unreviewed (`FALSE`) or both (`NULL`) UniProt records. Matters only if `uploadlists` is `FALSE`.

Details

This function, depending on the `uploadlists` parameter, uses either the uploadlists service of UniProt or plain UniProt queries to obtain identifier translation tables. The possible values for `from` and `to` are the identifier type abbreviations used in the UniProt API, please refer to the table here: https://www.uniprot.org/help/api_idmapping. In addition, simple synonyms are available which realize a uniform API for the uploadlists and UniProt query based backends. These are the followings:

OmnipathR	**Uploadlists**	**UniProt query**	**Ensembl BioMart**
uniprot	ACC	id	uniprotswissprot
uniprot_entry	ID	entry name	
trembl	*reviewed = FALSE*	*reviewed = FALSE*	uniprotsptrembl
genesymbol	GENENAME	genes(PREFERRED)	external_gene_name
genesymbol_syn		genes(ALTERNATIVE)	external_synonym
hgnc	HGNC_ID	database(HGNC)	hgnc_symbol
entrez	P_ENTREZGENEID	database(GeneID)	
ensembl	ENSEMBL_ID		ensembl_gene_id
ensg	ENSEMBL_ID		ensembl_gene_id
enst	ENSEMBL_TRS_ID	database(Ensembl)	ensembl_transcript_id
ensp	ENSEMBL_PRO_ID		ensembl_peptide_id
ensgg	ENSEMBLGENOME_ID		
ensgt	ENSEMBLGENOME_TRS_ID		
ensgp	ENSEMBLGENOME_PRO_ID		
protein_name		protein names	
pir	PIR	database(PIR)	
ccds		database(CCDS)	
refseqp	P_REFSEQ_AC	database(refseq)	
ipro			interpro
ipro_desc			interpro_description
ipro_sdesc			interpro_short_description
wikigene			wikigene_name

rnacentral			rnacentral
gene_desc			description
wormbase		database(WormBase)	
flybase		database(FlyBase)	
xenbase		database(Xenbase)	
zfin		database(ZFIN)	
pbd	PBD_ID	database(PDB)	pbd

The mapping between identifiers can be ambiguous. In this case one row in the original data frame yields multiple rows or elements in the returned data frame or vector(s).

Value

- Data frame: if the input is a data frame or the input is a vector and `return_df` is `TRUE`.
- Vector: if the input is a vector, there is only one target ID type and `return_df` is `FALSE`.
- List of vectors: if the input is a vector, there are more than one target ID types and `return_df` is `FALSE`. The names of the list will be ID types (as they were column names, see the description of the `...` argument), and the list will also include the source IDs.

See Also

- `uniprot_id_mapping_table`
- `uniprot_full_id_mapping_table`
- `ensembl_id_mapping_table`

Examples

```
d <- data.frame(uniprot_id = c('P00533', 'Q9ULV1', 'P43897', 'Q9Y2P5'))
d <- translate_ids(d, uniprot_id = uniprot, genesymbol)
d
#   uniprot_id genesymbol
# 1     P00533       EGFR
# 2     Q9ULV1       FZD4
# 3     P43897       TSFM
# 4     Q9Y2P5    SLC27A5
```

---

trembls_only *Retain only TrEMBL IDs*

Description

Retain only TrEMBL IDs

Usage

```
trembls_only(uniprots, organism = 9606)
```

Arguments

uniprots	Character vector of UniProt IDs.
organism	Character or integer: name or identifier of the organism.

Value

Character vector with only TrEMBL IDs.

Examples

```
trembls_only(c("Q05BL1", "A0A654IBU3", "P00533"))
# [1] "Q05BL1" "A0A654IBU3"
```

---

trrust_download *Downloads TF-target interactions from TRRUST*

Description

TRRUST v2 (https://www.grnpedia.org/trrust/) is a database of literature mined TF-target interactions for human and mouse.

Usage

```
trrust_download(organism = "human")
```

Arguments

organism	Character: either "human" or "mouse".

Value

A data frame of TF-target interactions.

Examples

```
trrust_interactions <- trrust_download()
trrust_interactions
# # A tibble: 11,698 x 4
#    source_genesymbol target_genesymbol effect reference
#    <chr>             <chr>               <dbl> <chr>
#  1 AATF              BAX                    -1 22909821
#  2 AATF              CDKN1A                  0 17157788
#  3 AATF              KLK3                    0 23146908
#  4 AATF              MYC                     1 20549547
#  5 AATF              TP53                    0 17157788
#  6 ABL1              BAX                     1 11753601
#  7 ABL1              BCL2                   -1 11753601
# # . with 11,688 more rows
```

`uniprot_full_id_mapping_table` *Creates an ID translation table from UniProt data*

Description

Creates an ID translation table from UniProt data

Usage

```
uniprot_full_id_mapping_table(
  to,
  from = "id",
  reviewed = TRUE,
  organism = 9606
)
```

Arguments

`to`	Character or symbol: target ID type. See Details for possible values.
`from`	Character or symbol: source ID type. See Details for possible values.
`reviewed`	Translate only reviewed (`TRUE`), only unreviewed (`FALSE`) or both (`NULL`) UniProt records.
`organism`	Integer, NCBI Taxonomy ID of the organism (by default 9606 for human).

Details

For both source and target ID type, this function accepts column codes used by UniProt and some simple shortcuts defined here. For the UniProt codes please refer to https://www.uniprot.org/help/uniprotkb The shortcuts are entrez, genesymbol, genesymbol_syn (synonym gene symbols), hgnc, embl, refseqp (RefSeq protein), enst (Ensembl transcript), uniprot_entry (UniProtKB AC, e.g. EGFR_HUMAN), protein_name (full name of the protein), uniprot (UniProtKB ID, e.g. P00533). For a complete table please refer to `translate_ids`.

Value

A data frame (tibble) with columns ‘From‘ and ‘To‘, UniProt IDs and the corresponding foreign IDs, respectively.

See Also

- `translate_ids`
- `ensembl_id_mapping_table`
- `uniprot_id_mapping_table`

Examples

```
uniprot_entrez <- uniprot_full_id_mapping_table(to = 'entrez')
uniprot_entrez
# # A tibble: 20,723 x 2
#    From   To
#    <chr>  <chr>
#  1 Q96R72 NA
#  2 Q9UKL2 23538
#  3 Q9H205 144125
#  4 Q8NGN2 219873
#  5 Q8NGC1 390439
# # . with 20,713 more rows
```

---

uniprot_genesymbol_cleanup

TrEMBL to SwissProt by gene names

Description

TrEMBL to SwissProt by gene names

Usage

```
uniprot_genesymbol_cleanup(uniprots, organism = 9606, only_trembls = TRUE)
```

Arguments

uniprots	Character vector possibly containing TrEMBL IDs.
organism	Character or integer: organism name or identifier.
only_trembls	Attempt to convert only known TrEMBL IDs of the organism. This is the recommended practice.

Details

Sometimes one gene or protein is represented by multiple identifiers in UniProt. These are typically slightly different isoforms, some of them having TrEMBL IDs, some of the SwissProt. For the purposes of most systems biology application, the most important is to identify the protein or gene in a way that we can recognize it in other datasets. Unfortunately UniProt or Ensembl do not seem to offer solution for this issue. Hence, if we find that a TrEMBL ID has a gene name which is also associated with a SwissProt ID, we replace this TrEMBL ID by that SwissProt. There might be a minor difference in their sequence, but most of the omics analyses do not even consider isoforms. And it is quite possible that later UniProt will convert the TrEMBL record to an isoform within the SwissProt record. Typically this translation is not so important (but still beneficial) for human, but for other organisms it is critical especially when translating from foreign identifiers.

This function accepts a mixed input of UniProt IDs and provides a distinct translation table that you can use to translate your data.

Value

Data frame with two columns: "input" and "output". The first one contains all identifiers from the input vector ‘uniprots‘. The second one has the corresponding identifiers which are either SwissProt IDs with gene names identical to the TrEMBL IDs in the input, or if no such records are available, the output has the input items unchanged.

Examples

```
## Not run:
uniprot_genesymbol_cleanup('Q6PB82', organism = 10090)
# # A tibble: 1 × 2
#   input  output
#   <chr>  <chr>
# 1 Q6PB82 O70405

## End(Not run)
```

---

uniprot_id_mapping_table

ID translation data from UniProt Uploadlists

Description

Retrieves an identifier translation table from the UniProt Uploadlists service.

Usage

```
uniprot_id_mapping_table(identifiers, from, to, chunk_size = NULL)
```

Arguments

identifiers	Character vector of identifiers
from	Character or symbol: type of the identifiers provided. See Details for possible values.
to	Character or symbol: identifier type to be retrieved from UniProt. See Details for possible values.
chunk_size	Integer: query the identifiers in chunks of this size. If you are experiencing download failures, try lower values.

Details

This function uses the uploadlists service of UniProt to obtain identifier translation tables. The possible values for ‘from‘ and ‘to‘ are the identifier type abbreviations used in the UniProt API, please refer to the table here: https://www.uniprot.org/help/api_idmapping or the table of synonyms supported by the current package: translate_ids. Note: if the number of identifiers is larger than the chunk size the log message about the cache origin is not guaranteed to be correct (most of the times it is still correct).

Value

A data frame (tibble) with columns ‘From‘ and ‘To‘, the identifiers provided and the corresponding target IDs, respectively.

See Also

translate_ids

Examples

```
uniprot_genesymbol <- uniprot_id_mapping_table(
    c('P00533', 'P23771'), uniprot, genesymbol
)
uniprot_genesymbol
# # A tibble: 2 x 2
#   From   To
#   <chr>  <chr>
# 1 P00533 EGFR
# 2 P23771 GATA3
```

---

uniprot_id_type *UniProt identifier type label*

Description

UniProt identifier type label

Usage

```
uniprot_id_type(label)
```

Arguments

label	Character: an ID type label, as shown in the table at translate_ids

Value

Character: the UniProt specific ID type label, or the input unchanged if it could not be translated (still might be a valid identifier name). This is the label that one can use in UniProt REST queries.

See Also

- ensembl_id_type
- uploadlists_id_type

Examples

```
ensembl_id_type("entrez")
# [1] "database(GeneID)"
```

`unique_intercell_network` *Unique intercellular interactions*

Description

In the intercellular network data frames produced by `import_intercell_network`, by default each pair of annotations for an interaction is represented in a separate row. This function drops the annotations and keeps only the distinct interacting pairs.

Usage

```
unique_intercell_network(network, ...)
```

Arguments

`network`	An intercellular network data frame as produced by `import_intercell_network`.
`...`	Additional columns to keep. Note: if these have multiple values for an interacting pair, only the first row will be preserved.

Value

A data frame with interacting pairs and interaction attributes.

See Also

- `import_intercell_network`
- `simplify_intercell_network`
- `filter_intercell_network`

Examples

```
icn <- import_intercell_network()
icn_unique <- unique_intercell_network(icn)
```

uploadlists_id_type *UniProt Uploadlists identifier type label*

Description

UniProt Uploadlists identifier type label

Usage

```
uploadlists_id_type(label)
```

Arguments

label	Character: an ID type label, as shown in the table at translate_ids

Value

Character: the UniProt Uploadlists specific ID type label, or the input unchanged if it could not be translated (still might be a valid identifier name). This is the label that one can use in UniProt Uploadlists (ID Mapping) queries.

See Also

- ensembl_id_type
- uniprot_id_type

Examples

```
ensembl_id_type("entrez")
# [1] "P_ENTREZGENEID"
```

vinayagam_download *Protein-protein interactions from Vinayagam 2011*

Description

Retrieves the Supplementary Table S6 from Vinayagam et al. 2011. Find out more at https://doi.org/10.1126/scisignal.2001699.

Usage

```
vinayagam_download()
```

Value

A data frame (tibble) with interactions.

Examples

```
vinayagam_interactions <- vinayagam_download()
vinayagam_interactions
# # A tibble: 34,814 x 5
#    `Input-node Gen. `Input-node Gen. `Output-node Ge. `Output-node Ge.
#    <chr>                      <dbl> <chr>                      <dbl>
#  1 C1orf103                   55791 MNAT1                       4331
#  2 MAST2                      23139 DYNLL1                      8655
#  3 RAB22A                     57403 APPL2                      55198
#  4 TRAP1                      10131 EXT2                        2132
#  5 STAT2                       6773 COPS4                      51138
# # . with 34,804 more rows, and 1 more variable:
# # `Edge direction score` <dbl>
```

---

walk_ontology_tree *All nodes of a subtree starting from the selected nodes*

Description

Starting from the selected nodes, recursively walks the ontology tree until it reaches either the root or leaf nodes. Collects all visited nodes.

Usage

```
walk_ontology_tree(
  terms,
  ancestors = TRUE,
  db_key = "go_basic",
  ids = TRUE,
  method = "gra",
 relations = c("is_a", "part_of", "occurs_in", "regulates", "positively_regulates",
    "negatively_regulates")
)
```

Arguments

terms	Character vector of ontology term IDs or names. A mixture of IDs and names can be provided.
ancestors	Logical: if FALSE the ontology tree is traversed towards the leaf nodes; if TRUE, the tree is traversed until the root. The former returns the ancestors (parents), the latter the descendants (children).
db_key	Character: key to identify the ontology database. For the available keys see omnipath_show_db.
ids	Logical: whether to return IDs or term names.

method	Character: either "gra" or "lst". The implementation to use for traversing the ontology tree. The graph based implementation is faster than the list based, the latter will be removed in the future.
relations	Character vector of ontology relation types. Only these relations will be used.

Details

Note: this function relies on the database manager, the first call might take long because of the database load process. Subsequent calls within a short period should be faster. See get_ontology_db.

Value

Character vector of ontology IDs. If the input terms are all leaves or roots NULL is returned. The starting nodes won't be included in the result unless they fall onto the traversal path from other nodes.

See Also

- omnipath_show_db
- get_ontology_db

Examples

```
walk_ontology_tree(c('GO:0006241', 'GO:0044211'))
# [1] "GO:0006139" "GO:0006220" "GO:0006221" "GO:0006241" "GO:0006725"
# [6] "GO:0006753" "GO:0006793" "GO:0006796" "GO:0006807" "GO:0008150"
# ... (truncated)
walk_ontology_tree(c('GO:0006241', 'GO:0044211'), ancestors = FALSE)
# [1] "GO:0044210" "GO:0044211"
walk_ontology_tree(
    c('GO:0006241', 'GO:0044211'),
    ancestors = FALSE,
    ids = FALSE
)
# [1] "'de novo' CTP biosynthetic process" "CTP salvage"
```

---

with_extra_attrs *Interaction records having certain extra attributes*

Description

Interaction records having certain extra attributes

Usage

```
with_extra_attrs(data, ...)
```

Arguments

data	An interaction data frame.
...	The name(s) of the extra attributes; NSE is supported.

Value

The data frame filtered to the records having the extra attribute.

See Also

- extra_attrs
- has_extra_attrs
- extra_attrs_to_cols
- filter_extra_attrs
- extra_attr_values

Examples

```
i <- import_omnipath_interactions(fields = 'extra_attrs')
with_extra_attrs(i, Macrophage_type)
```

---

with_references *Interactions having references*

Description

Interactions having references

Usage

```
with_references(data, resources = NULL)
```

Arguments

data	An interaction data frame.
resources	Character: consider only these resources. If 'NULL', records with any reference will be accepted.

Value

A subset of the input interaction data frame.

Examples

```
cc <- import_post_translational_interactions(resources = 'CellChatDB')
with_references(cc, 'CellChatDB')
```

zenodo_download *Retrieves data from Zenodo*

Description

Zenodo is a repository of large scientific datasets. Many projects and publications make their datasets available at Zenodo. This function downloads an archive from Zenodo and extracts the requested file.

Usage

```
zenodo_download(
  path,
  reader = NULL,
  reader_param = list(),
  url_key = NULL,
  zenodo_record = NULL,
  zenodo_fname = NULL,
  url_param = list(),
  url_key_param = list(),
  ...
)
```

Arguments

path	Character: path to the file within the archive.
reader	Optional, a function to read the connection.
reader_param	List: arguments for the reader function.
url_key	Character: name of the option containing the URL
zenodo_record	The Zenodo record ID, either integer or character.
zenodo_fname	The file name within the record.
url_param	List: variables to insert into the URL string (which is returned from the options).
url_key_param	List: variables to insert into the 'url_key'.
...	Passed to archive_extractor

Value

A connection

Examples

```
## Not run:
# an example from the OmnipathR::remap_tf_target_download function:
remap_dorothea <- zenodo_download(
    zenodo_record = 3713238,
```

```
        zenodo_fname = 'tf_target_sources.zip',
        path = (
            'tf_target_sources/chip_seq/remap/gene_tf_pairs_genesymbol.txt'
        ),
        reader = read_tsv,
        reader_param = list(
            col_names = c(
                'source_genesymbol',
                'target_genesymbol',
                'target_ensembl',
                'score'
            ),
            col_types = cols(),
            progress = FALSE
        ),
      resource = 'ReMap'
    )

    ## End(Not run)
```

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